Independence has delegated utilization management of genetic/genomic testing and certain molecular analyses and cytogenetic tests to eviCore. eviCore uses the Lab Management Clinical Guidelines during the precertification and/or prepayment review of delegated services.
Please note the following:

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Dear Provider,

This document provides detailed descriptions of eviCore’s basic criteria for laboratory services. These criteria are used for the certification of requests and administration of laboratory benefits for our clients for a range of laboratory tests some of which are represented by one CPT or HCPCS code and others represented by several codes. They have been carefully researched and are continually updated in order to be consistent with the most current evidence-based guidelines and recommendations for laboratory testing from national and international medical societies and evidence-based medicine research centers. In addition, the criteria are supplemented by information published in peer reviewed literature. If you believe that our criteria require modification, please send suggested changes with supporting references to the Laboratory Management Program at the address listed below.

Our health plan clients review the development and application of these criteria. Every eviCore health plan client develops a unique list of CPT codes that are part of their utilization management programs. Health Plan medical policy supersedes eviCore when there is conflict with the eviCore criteria and the health plan medical policy. If you are unsure of whether or not a specific health plan has made modifications to these basic criteria in their medical policy please contact the plan or access the plan’s website for additional information.

eviCore works hard to make your clinical review experience a pleasant one. For that reason, we have peer reviewers available to assist you should you have specific questions about a procedure. For your convenience, eviCore’s Customer Service support is available from 7 a.m. to 7 p.m. Our toll-free number is 1-800-918-8924.

Gregg P. Allen, M. D. FAAFP

EVP and Chief Medical Officer
General Information About this Guideline Manual

Description

The eviCore healthcare (eviCore) guideline manual contains medical and reimbursement guidelines that are created and approved by eviCore’s Laboratory Management Program personnel and advisors, internal Medical Advisory Committee, and external Medical Advisory Board. eviCore’s guidelines are created using evidence-based medicine including, but not limited to, professional society guidelines, consensus statements, and peer-reviewed literature. eviCore’s guidelines are intended to provide a library for adoption or a basis for development of tailored coverage criteria for a Health Plan.

Purpose

To establish evidence-based definitions, decision support, medical necessity criteria, coverage limitations, and payment rules for molecular and genetic testing.

Organization

This manual is organized into the following sections.

Molecular and Genetic Clinical Use Guidelines

The guidelines in this section are intended to provide general guidance for the common settings and scenarios in which genetic testing is used (e.g. prenatal, diagnostic, cancer). These guidelines address the overarching coverage principles that broadly apply based on the purpose of the test. They also address specific use situations that may apply to many different tests (e.g. predictive testing for a known familial mutation).

Clinical Use guidelines may include a test-specific guidelines section to direct users to any relevant test-specific guidelines. Because tests may be used for multiple indications, the same test-specific guideline may be referenced by more than one Clinical Use guideline. When a test specific guideline is not available, the coverage principles found in these Clinical Use Guidelines will be applied.

Molecular and Genetic Test Specific Guidelines

The guidelines in this section address a test or group of tests that are used to assess some health condition. The purpose of these guidelines is to provide a framework for determining medical necessity and coverage determinations for a specific test, including where more limited testing may be supported by the medical evidence when broader testing is not. These guidelines provide background about each condition, the available tests, the scenarios in which the test may be used, and the evidence used to determine medical necessity criteria.
Administrative Guidelines

If applicable for this plan, administrative guidelines are included that define coding and reimbursement criteria and requirements.

Glossary

This glossary contains definitions for common genetics, medical and laboratory terminology.

Limitations and Restrictions

When using this manual in electronic or printed form, the following restrictions apply:

- Evidence-based genetic testing is defined as the identification of targeted genetic sequences within the genome of an individual with clinically-identified risk factors or traits suspected of being specific to the genetic disorder, condition, or trait under investigation.

- The medical guidelines contained in this manual are the proprietary property of eviCore, for use by its clients only. These medical guidelines may not be posted, shared, altered, cited or reproduced without the express written consent of eviCore. Commercial use of these guidelines is prohibited.

- Medical guidelines are not to be considered medical advice for a specific patient. Guidelines are used in the process of determining whether a service may be medically necessary and eligible for coverage.

- Medical Guidelines are interpreted and applied at the sole discretion of the Health Plan.

- Current Procedural Terminology (CPT®) codes and descriptions are the property of the American Medical Association with all rights reserved.
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Confirmatory Genetic Testing

Description

The Centers for Medicare and Medicaid Services (CMS) developed the Clinical Laboratory Amendments (CLIA) in order to help regulate laboratory tests. CMS intended to use this program as a way to ensure that quality laboratory testing was performed. Laboratories that receive reimbursement from Medicare or Medicaid must be CLIA certified.¹

Most genetic or genomic tests are performed in a CLIA certified laboratory and used for a clear medical purpose. However, some genetic or genomic tests are performed in a research laboratory that is not CLIA certified or as part of a direct to consumer test that is not necessarily performed for a medical purpose.

When genetic testing is performed in a research laboratory or in a laboratory that is not CLIA certified, it is important to confirm any genetic change found prior to using this information to change an individual’s medical treatment.

Criteria

Confirmatory single site genetic testing in a CLIA certified laboratory will be approved when the following criteria are met:

• A disease-causing genetic mutation was identified by a laboratory that is not CLIA certified (e.g. research lab), AND
• Healthcare providers can use the test results to directly impact medical care for the individual (e.g. change in surveillance or treatment plan)

Exclusions

• Confirmatory genetic testing is not considered medically necessary if the original testing was performed in a CLIA certified laboratory.
• Confirmatory genetic testing is not considered medically necessary if healthcare providers cannot use the test results to directly impact medical care for the individual (e.g. APOE).
• Confirmatory genetic testing is not considered medically necessary if testing is considered Investigational/Experimental per eviCore clinical guidelines (e.g. APOE).
• Confirmatory genetic testing is not considered medically necessary for variants of unknown significance (VUS).

References

Genetic Testing by Multigene Panels

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan’s procedure code list for management requirements.

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What are multi-gene panels?

Definition

Various methodologies can be used to identify potential disease-causing gene mutations. Gene sequencing involves evaluating each DNA nucleotide along the length of a gene. Full gene sequencing is the best approach when many different mutations in the same gene can cause the disorder.

- There are two main ways to sequence a gene:
  - Until recently, most sequencing tests used the Sanger sequencing methodology that was originally developed in the 1970s. Sanger sequencing is labor intensive and did not lend itself to high-throughput applications.¹
  - Next generation sequencing (NGS), also called massively parallel sequencing, has been developing since about 2005 to allow larger scale and more efficient gene sequencing. NGS relies on sequencing many copies of small pieces of DNA simultaneously and using bioinformatics to assemble the sequence.¹

- The efficiency of NGS has led to an increasing number of large, multi-gene testing panels.
  - NGS panels are particularly well-suited to conditions caused by more than one gene or where there is considerable clinical overlap between conditions making it difficult to reliably narrow down likely causes.
  - Panels including genes associated with a high risk of a condition are of greatest value since these mutation-positive results often lead to changes in medical management.
- Panels may also include genes believed to be associated with a particular condition, but with a more modest impact on risk. Results for such genes are of less clear value because there often are not clear management recommendation for mutation-positive individuals.

- Laboratories offer panel testing for multiple genes at the same time in an effort to increase the likelihood of finding a causative gene mutation in a more efficient manner. Such testing may be performed for diagnostic or predictive purposes.

- Diagnostic testing is performed in patients with clinical signs or symptoms of a genetic condition. The genetic test may confirm or rule out a clinical diagnosis. However, many genetic conditions have overlapping features, which can make determining appropriate genetic testing difficult. The use of clinical and family history information may not always lead to a likely diagnosis for an individual. In some cases, many genes may be candidates for a person’s symptoms. In these cases, testing one gene at a time may be time-consuming and costly. It may also lead to a situation where a mutation is missed in another gene that was not tested.\(^2\)

- Predictive genetic testing is performed in people known to be at increased risk of developing an inherited condition based on their family history. For some conditions, a positive genetic test predicts with certainty that the person will eventually develop signs and symptoms of a condition. For other conditions, a positive genetic test result indicates an increased risk (susceptibility) for a condition. Without a specific known mutation running in the family, a negative result rarely rules out a condition. Having test results may improve medical management through improved screening, preventive measures (e.g. prophylactic medication, surgery) and other means. In order to better define a person’s risk, it is preferable to first test someone in the family who is affected.

**Test information**

- Multi-gene panel tests, even for similar clinical scenarios, vary considerably in the genes that are included and in technical specifications (e.g. depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis). Therefore, technologies used in multi-gene testing may fail to identify mutations that might be identifiable through single-gene testing.

- If high clinical suspicion remains for a particular syndrome after negative multi-gene test results, consultation with the testing lab and/or additional targeted genetic testing may be warranted.

- Results may be obtained that cannot be adequately interpreted based on the current knowledgebase. When a sequence variation is identified that has not been previously characterized or shown to cause the disorder in question, it is called a variant of uncertain significance (VUS). VUSs are relatively common findings when sequencing large amounts of DNA with NGS.\(^3\)
• Since genes can be easily added or removed from multi-gene tests over time by a given lab, medical records must document which genes were included in the specific multi-gene test used from each patient, and in which labs they were performed.

• Tests should be chosen that maximize the likelihood of identifying mutations in the genes of interest.

Guidelines and evidence

• The American College of Medical Genetics has a policy statement that offers general guidance on the clinical application of large-scale sequencing focusing primarily on whole exome and whole genome testing. However, some of the recommendations regarding counseling around unexpected results and variants of unknown significance and minimum requirements for reporting apply to many applications of NGS sequencing applications.\(^4\)

Criteria

• This guideline applies to multi-gene panel testing, which is defined as any assay that simultaneously tests for more than one gene associated with a condition. The testing may focus on sequence variants and/or deletions/duplications of those genes. Panels vary in scope, such as:
  
  o Panels consisting of multiple genes that are associated with one specific genetic condition (e.g. Noonan syndrome,\(^5\) Stickler syndrome,\(^6\) etc.)
  
  o Panels consisting of multiple genes that are associated with a symptom or non-specific presentation (e.g. epilepsy, intellectual disability,\(^7\) hearing loss, retinal disorders, etc.)

• Coverage determinations generally rely on the medical necessity of the components of a panel. A panel approach to testing is most compelling when:
  
  o Multiple genes are known to cause the same condition and a limited subset of genes does not account for the majority of disease-causing mutations.
  
  o The clinical presentation is highly suspicious for a genetic disorder, but the constellation of findings in the personal or family history does not suggest a specific diagnosis or limited set of conditions.

• Multiple policies may apply, including test-specific policies where they exist or the following clinical use policies:
  
  o Genetic Testing to Diagnose Non-Cancer Conditions
Genetic Testing to Predict Disease Risk

- Panel coding and billing should reflect the efficiency gains for the laboratory in testing multiple candidate genes simultaneously. Currently, laboratories are billing for panels in a variety of ways. When a panel approach to testing is determined to be medically necessary, the following billing guidelines will apply.

  - Panel is to be billed with a single panel-specific code (e.g., Genomic Sequencing Procedure or GSP) or single unit of the unlisted molecular pathology code 81479:
    - The billed amount should not exceed the list price of the test.

  - Panel is to be billed with multiple procedure codes representing individual genes analyzed:
    - If a more specific code exists that adequately describes the requested panel, the panel will be redirected to the more specific code (e.g., a genomic sequencing procedure code), or
    - If no more specific code exists, the panel will be redirected to a single unit of the unlisted molecular pathology code 81479, which can be used to represent a panel in total, or
    - If the laboratory will not accept redirection to a single code, the medical necessity of each billed component procedure will be assessed independently. Only the individual panel components that meet medical necessity criteria as a first tier of testing will be reimbursed. The remaining individual components will not be reimbursable, and
    - The billed amount should not exceed the list price of the test.

- The following general principles apply:
  - Broad symptom-based panels (e.g. comprehensive ataxia panel) are not medically necessary when a narrower panel is available and more appropriate based on the clinical findings (e.g. autosomal dominant ataxia panel).
  - More than one multi-gene panel should not be necessary at the same time. Multi-gene panel testing should be performed in a tiered fashion with independent justification for each panel requested.
  - If more than ten units of any combination of procedure codes will be billed as part of a panel with no stated differential, the panel will be deemed excessive and not medically necessary.
  - Genetic testing is only necessary once per lifetime. Therefore, a single gene included in a panel or a multi-gene panel may not be reimbursed if testing has been performed previously. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.
Whole genome sequencing is considered to be experimental/investigational even when billed as a panel.

This guideline may not apply to multi-gene panel testing for indications that are addressed in test-specific guidelines. Please see the test-specific list of guidelines for a complete list of test-specific panel guidelines.

Other considerations

If a panel was previously performed and an updated, larger panel is being requested, only testing for the medically necessary, previously untested genes will be reimbursable. Therefore, only the most appropriate procedure codes for those additional genes will be considered for reimbursement.

References


2. Memorial Sloan Kettering Cancer Center. When to consider multigene panels. April 23, 2015. Available at: https://www.mskcc.org/blog/should-i-consider-multigene-panel-testing.


Genetic Testing for Cancer Susceptibility and Hereditary Cancer Syndromes

Description

Genetic testing for cancer susceptibility and hereditary cancer syndromes is performed in people with known risk factors for an inherited form of cancer. Testing may be used in people diagnosed with cancer when there are “red flags” in the individual’s personal medical and/or family history for a hereditary form. Predictive genetic testing may also be performed for this group of conditions, in people known to be at increased risk of developing an inherited condition based on their family history. A positive genetic test result increases the risk for cancer (types vary by the gene involved) and, therefore, impacts medical management decisions around screening, prevention, and treatment.

- Tests used to screen for or make a diagnosis of cancer are covered separately as Genetic Testing for the Screening, Diagnosis, and Monitoring of Cancer.
- This policy does not address diagnostic or predictive testing for conditions other than hereditary cancer. Refer to Genetic Testing to Diagnose Non-Cancer Conditions and Genetic Testing to Predict Disease Risk for those purposes.

Criteria

Criteria: General Coverage Guidance

Individuals may be considered for genetic testing for hereditary cancer syndromes when ALL of the following conditions are met:

- **Technical and clinical validity**: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.
- **Clinical utility**: Healthcare providers can use the test results to provide significantly better medical care for the individual.
- **Reasonable use**: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Limits:

- Testing will be considered only for the number of genes or tests necessary to establish carrier status. A tiered approach to testing, with reflex to more detailed testing and/or different genes, will be required when clinically possible.
• Genetic testing is indicated once per lifetime per condition. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

**Criteria: Special Circumstances**

The following policies address a group of tests that are used for similar purposes. Because a variety of tests may be used, but the circumstances that justify testing are the same, individual test-specific policies are not necessary.

**Predictive testing for at-risk people with known familial mutations**

The genetic mutation(s) associated with a hereditary cancer syndrome can often be defined in an affected family member, allowing for testing of at-risk relatives for those specific mutations. Testing for known familial mutations is reasonable when **ALL** of the following conditions are met:

- The mutation(s) in the family have been clearly defined by previous genetic testing and information about those mutations can be provided to the testing lab.
- Technical and clinical validity: The test must be accurate, sensitive and specific to the familial mutation(s).
- Clinical utility: Healthcare providers can use the test results to provide significantly better medical care for the individual.
- Reasonable use: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

**Limits:**

- Testing will be considered only for the known familial mutations when clinically possible.
- Predictive genetic testing is indicated once per lifetime per condition.
- Predictive genetic testing will be considered only for adult individuals (age 18 and over). Exceptions may be considered if there are medical management and/or significant psychosocial benefits to testing prior to adulthood.$^{1,2}$

**Criteria: Test-specific Policies**

Test-specific policies are available for some hereditary cancer syndrome tests. Please see the policy manual for a list of test-specific policies. For tests without a specific policy, use the General Coverage Guidance in Section 1.
References


Genetic Testing for Carrier Status

Introduction

Carrier screening is performed to identify genetic risks that could impact reproductive decision-making for parents or prospective parents. Carriers are generally not affected but have an increased risk to have a child with a genetic condition.

Availability of genetic testing for carrier status

Carrier screening may be available for autosomal recessive conditions, X-linked conditions, and certain chromosome abnormalities. Ideally, carrier screening is performed prior to pregnancy so that a full range of reproductive options are available to an at-risk couple. However, in practice, it is often performed early in pregnancy when prenatal care is established.

Other applications of carrier testing

This guideline does not address prenatal or preimplantation genetic testing. Refer to guidelines on Genetic Testing for Prenatal Screening and Diagnostic Testing and Preimplantation Genetic Screening and Diagnosis for those purposes.

Testing that may identify carriers who have clinical signs and symptoms, such as cystic fibrosis testing for men with congenital absence of the vas deferens or fragile X genetic testing for women with premature ovarian failure, is addressed in Genetic Testing to Diagnose Non-Cancer Conditions.

Criteria

Introduction

Requests for carrier screening are reviewed using these criteria.

Criteria for general coverage guidance

Individuals may be considered for genetic testing for carrier screening when ALL of the following conditions are met:

- **Technical and clinical validity** — The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.
- **Clinical utility** — Healthcare providers can use the test results to provide significantly better medical care and/or assist individuals with reproductive planning.
• **Reasonable use** — The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

**Limits**

- Testing will only be considered for the number of genes or tests necessary to establish carrier status. A tiered approach to testing, with reflex to more detailed testing and/or different genes, will be required when clinically possible.
- Carrier testing will be allowed once per lifetime. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.
- Carrier testing is indicated only in adults. Carrier screening in minor children is not indicated, except in the case of a pregnancy of the minor child.

**Routine carrier screening**

Individuals may be considered for routine carrier screening when testing is supported by evidence-based guidelines from governmental organizations and/or well-recognized professional societies in the United States.

**Carrier screening based on family history**

Individuals may be considered for carrier screening based on a family history of a genetic condition when ALL of the following conditions are met in addition to the general criteria above:

- The diagnosis of a genetic condition in a family member is known.
- The parent(s) or prospective parent(s) are at-risk to be carriers of that condition based on the pattern of inheritance.
- The genetic condition is associated with potentially severe disability or has a lethal natural history.

**Partner testing of known carrier or affected individuals**

Individuals may be considered for carrier screening if their partners are known carrier or affected individuals when all of the following conditions are met in addition to the general criteria above:

- The diagnosis of a genetic condition or carrier status in the partner is known.
- The genetic condition is associated with potentially severe disability or has a lethal natural history.

**Exclusions for multiplex carrier screening tests**

Multiplex carrier screening tests are designed to identify carrier status or predict risk for many genetic diseases (70 or more) in a single test. Several multiplex carrier screening
tests are available now. Others are known to be in development and will come to market in the next few years. Each test has a unique set of diseases included in novel and proprietary genetic testing platforms.

Of the genetic conditions included in the currently available multiplex carrier screening tests, 12 of them are recommended for at least some people based on ethnicity by either the American College of Obstetrics and Gynecology (ACOG) and/or the American College of Medical Genetics (ACMG). However, mutation analysis is not the preferred initial screening test for some.

These tests do not meet the criteria above for technical and clinical validity and clinical utility:

- The technologies used by the multiplex carrier screening tests are novel. Information about the test's performance, if available, is often provided completely by the laboratory marketing the test, which could be subject to bias.

- Some of the commonly included tests, such as beta-thalassemia and Tay-Sachs disease, have inexpensive and reliable screening tests available (CBC with RBC indices and hexosaminidase A enzyme activity, respectively) that are superior to genetic testing.

- Multiplex carrier screening tests typically include carrier screening for many diseases that have not been identified as appropriate for population-based carrier screening. They may also include disorders, such as hereditary hemochromatosis and factor V Leiden, which affect primarily adults and are generally manageable. These kinds of conditions do not meet the requirements for reproductive carrier screening programs.

**Test-specific guidelines**

Test-specific guidelines are available for some tests designed to predict carrier status. Please see the clinical guidelines manual for a list of test-specific guidelines. For tests without a specific guideline, use the General Coverage Guidance in Section 1.
Genetic Testing for Non-Medical Purposes

Description

While most traditional genetic tests are used for clear medical purposes, advances in gene discovery and genetic testing technology allow laboratories to offer genetic testing for other uses. Testing for paternity, ancestry, and non-disease traits such as baldness and eye color may be highly accurate and interesting. However, because these kinds of tests are not useful for medical management in the vast majority of cases, they are typically excluded from consideration.

Criteria

Criteria: General Coverage Guidance

Any genetic test that DOES NOT meet the following criteria is excluded from consideration:

- **Technical and clinical validity**: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.
- **Clinical utility**: Healthcare providers can use the test results to provide significantly better medical care for the individual.
- **Reasonable use**: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Criteria: Test-specific Policies

The following types of testing are specifically excluded from consideration:

- Genome-wide association studies (GWAs): testing a large number of genetic variations spread across the whole genome for disease associations, generally done for information outside of a specific clinical need or context
  - Common trade names: 23andMe, Navigenics, Pathway Genomics, deCODEme
- Paternity testing: testing to establish biological relationships, often between a father and child(ren) but sometimes to determine other kinds of relationships (siblings, grandparents, etc.)
- Ancestry testing: testing that helps people discover more about the genetic make-up of their ancestors, generally used by genealogists and those interested in family history
Common trade names: Ancestry.com, 23andMe, Pathway Genomics, Family Tree DNA, deCODEme

- Non-disease trait testing: testing for physical traits (e.g., eye color, hair color, male pattern baldness, and cellulite) that do not have associated health problems, or can be deemed cosmetic in nature.

- Nutritional testing: for variations in metabolism pathways that may suggest vitamin or other nutritional supplements.
  - Common trade names: MyDNAVitamins, GeneWise

- Athletic ability: Testing to predict athletic performance types.
  - Common trade names: Sports Gene, Athleticode

- Genetic testing related to dating services
  - Common trade names: Scientific Match, GenePartner
Genetic Testing for Prenatal Screening and Diagnostic Testing

Description

Prenatal screening and diagnostic testing is performed during pregnancy to identify fetuses at increased risk for or affected with genetic conditions and birth defects. Screening with ultrasound and maternal serum markers is routinely offered. Prenatal diagnosis by chorionic villus sampling or amniocentesis for chromosome abnormalities is available to all women. However, it is usually offered specifically to those at higher risk because of maternal age, a positive screen result, abnormal ultrasound findings, or known risk of a genetic condition based on family history. Investigations for fetal infection and blood antigen incompatibility may also be performed in the prenatal period. Results of testing are used to guide reproductive decision-making, pregnancy management and anticipatory management of the infant at birth.

Note
This policy does not include prenatal or preconception carrier screening or preimplantation genetic testing. Please refer to Genetic Testing for Carrier Status and Preimplantation Genetic Screening and Diagnosis for those purposes.

Criteria

Criteria: General Coverage Guidance

Individuals may be considered for genetic testing for prenatal screening and diagnostic testing when ALL of the following conditions are met:

- **Technical and clinical validity**: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.
- **Clinical utility**: Healthcare providers can use the test results to provide significantly better medical care and/or assist patients with reproductive planning.
- **Reasonable use**: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Limits:

- Testing will only be covered for the number of genes or tests necessary to establish a prenatal diagnosis. A tiered approach to testing, with reflex to more detailed testing and/or different genes, will be required when clinically possible.
- Prenatal diagnostic testing will be allowed once per pregnancy. Exceptions may be considered if ambiguous results require retesting for clarification.
• If prenatal samples are studied concurrently with a maternal DNA sample to rule out prenatal analytic errors due to maternal cell contamination, a single unit of CPT code 81265 may be approved.

Criteria: Special Prenatal Diagnosis Circumstances

Each of the following policies addresses a group of tests that are used for similar purposes in pregnancy. Because a variety of tests may be used, but the circumstances that justify testing are the same, individual test-specific policies are not necessary.

Prenatal diagnostic testing based on family history

Prenatal genetic testing, generally by amniocentesis or CVS, for the diagnosis of a genetic condition is reasonable when the following conditions are met:

- The pregnancy is at an increased risk for a genetic disease because of ANY of the following:
  - At least one parent is known or suspected to be a carrier of a genetic condition based on the family history and/or previous carrier testing results; or
  - One or both parent(s) are affected with a genetic condition; or
  - A sibling is affected with a genetic condition; AND
- The genetic condition is associated with potentially severe disability or has a lethal natural history.

Fetal infectious disease testing

Genetic testing may be used for the diagnosis of an infectious disease (e.g., cytomegalovirus, toxoplasmosis, parvovirus B19, and varicella zoster) in a fetus according to current guidelines from the American College of Obstetricians and Gynecologists (ACOG). Prenatal testing, generally by amniocentesis or CVS, is reasonable when ANY of the following conditions are met:

- Clinical signs and symptoms of a current infection in the mother; OR
- Serologic evidence of a current or recent infection in the mother (with or without clinical signs); OR
- Fetal abnormalities identified on ultrasound indicating an increased risk for a congenital infection

Blood antigen incompatibility testing

Prenatal genetic testing, generally by amniocentesis, for the determination of blood antigen genotype is supported by current evidence-based recommendations from the American College of Obstetricians and Gynecologists. Fetal antigen genotyping is reasonable when the following conditions are met:
A positive erythrocyte antibody screen in the mother; AND EITHER

- The father’s blood antigen genotype is known and indicates a risk for the fetus to be positive; OR
- The father’s blood antigen genotype is not known and unavailable

Criteria: Test-specific Policies

- Test-specific policies are available for some prenatal screening tests and diagnostic tests. Please see the policy manual for a list of test-specific policies. For tests without a specific policy, use the General Coverage Guidance in Section 1.

References

Genetic Testing for the Screening, Diagnosis, and Monitoring of Cancer

Description

Genetic testing for screening, diagnosis and monitoring of cancer refers to molecular diagnostic tests whose purposes include identifying the possible presence of cancer in asymptomatic, average risk individuals; confirming the absence or presence of cancer; and monitoring the absence or presence of cancer after a prior diagnosis and treatment.

Screening

The goal of cancer screening is to identify the possible presence of cancer before symptoms appear. Screening tests cannot diagnose cancer, but typically determine if there is an increased chance cancer is present, and triages individuals for more invasive, diagnostic testing. Most cancer screening does not include genetic testing, but instead relies on physical exam, radiological exams, or non-genetic laboratory tests. Advances in human genetics, however, have identified several molecular diagnostic tests that may provide clues for early cancer detection.

Diagnosis

When cancer is suspected because of an abnormal screening test or symptoms, blood tests for tumor markers or molecular testing on tissue samples can aid in confirming a diagnosis of cancer. These tests may contribute information to helping the clinician understand prognosis and treatment options.

Monitoring

During treatment, or after an apparently successful treatment, active monitoring is often recommended to identify if the cancer is responding to treatment or has returned or spread, before any symptoms appear. Monitoring may include increased surveillance or routine blood tests for tumor markers, and increasingly, molecular genetic tests.

- Tests used to determine hereditary cancer risk are covered separately as Genetic Testing for Cancer Susceptibility and Hereditary Cancer Syndromes.
- This policy does not address drug response to cancer, or testing to determine which therapies to use. Please refer to Pharmacogenomic Testing for Drug Toxicity and Response for that purpose.
- This policy does not address molecular tumor marker testing in solid tumors. Please refer to Tumor Marker Testing—Solid Tumors and Liquid Biopsy Testing – Solid Tumors for that purpose.
• This policy does not address diagnostic or predictive testing for conditions other than non-inherited cancer. Refer to Genetic Testing to Diagnose Non-Cancer Conditions and Genetic Testing to Predict Disease Risk for those purposes.

Criteria

Criteria: General Coverage Guidance

Individuals may be considered for genetic testing for screening, diagnosing, or monitoring cancer when ALL of the following conditions are met:

• Technical and clinical validity: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.

• Clinical utility: Healthcare providers can use the test results to provide significantly better medical care for the individual.

• Reasonable use: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Limits:

• Testing will be considered only for the number of genes or tests necessary. A tiered approach to testing, with reflex to more detailed testing and/or different genes, will be required when clinically possible.

• For tests that look for changes in germline DNA (i.e., not tumor DNA or viral DNA), testing will be allowed once per lifetime per gene. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

Criteria: Test-specific Policies

Test-specific policies are available for some tests designed to screen for, diagnose, or monitor cancer. Please see the policy manual for a list of test-specific policies. For tests without a specific policy, use the General Coverage Guidance in Section 1.
Genetic Testing to Diagnose Non-Cancer Conditions

Description

Diagnostic testing is performed in patients with clinical signs or symptoms of a non-cancer genetic condition. The genetic test may confirm or rule out a clinical diagnosis. In some cases, genetic testing is the gold standard for making a diagnosis based on evidence- or consensus-based guidelines. In others, it may be used to confirm a clinical diagnosis, offer prognostic information that impacts management, or rule out a diagnosis in the differential. Often, diagnostic testing of an affected individual will offer results that are relevant to the testing of other family members.

- This guideline does not include risk assessment or predictive testing for at-risk, asymptomatic individuals. Please refer to Genetic Testing to Predict Disease Risk for that purpose.
- Diagnostic testing of a pregnancy or an embryo is addressed by guidelines on Genetic Testing for Prenatal Screening and Diagnostic Testing and Preimplantation Genetic Screening and Diagnosis, respectively.
- In addition, testing for hereditary cancer syndromes is addressed separately under Genetic Testing for Cancer Susceptibility and Hereditary Cancer Syndromes.

Criteria

Criteria: General Coverage Guidance

Individuals may be considered for diagnostic genetic testing when ALL of the following conditions are met:

- Clinical signs and symptoms in the individual are consistent with the diagnosis in question.
- Technical and clinical validity: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.
- Clinical utility: Healthcare providers can use the test results to provide significantly better medical care for the individual.
- Reasonable use: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Limits:
• Testing will be considered only for the number of genes or tests necessary to establish carrier status. A tiered approach to testing, with reflex to more detailed testing and/or different genes, will be required when clinically possible.

• Diagnostic genetic testing will be allowed once per lifetime per condition. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

Criteria: Special Circumstances

Diagnostic testing of a child to inform reproductive planning and testing for parents or testing for siblings.

Diagnostic genetic testing may be requested in a symptomatic child with a known genetic condition. While diagnostic testing may not impact management of the affected child, the information gained from genetic testing may be needed to perform accurate carrier testing in the child’s parent(s) and/or genetic diagnosis in a sibling.*

In these circumstances, diagnostic genetic testing in the child may be considered when **ALL** of the following conditions are met:

• The diagnosis of the disease in the affected child is **certain or highly probable** based on clinical signs and symptoms, history, imaging, and/or results of other laboratory testing.

• The results of the genetic test in the symptomatic child must be **required** in order to perform accurate carrier testing in the child’s parent(s) and/or genetic diagnosis in a sibling.

• **Technical and clinical validity**: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.

• **Clinical utility**: Healthcare providers can use the test results to provide informative genetic testing for the child’s sibling, child’s parents, and/or for a current or future at-risk pregnancy.

• **Reasonable use**: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Limits:

• Testing will be indicated only for the number of genes or tests necessary to establish the familial mutation(s). A tiered approach to testing, with reflex to more detailed testing and/or different genes, will be required when clinically possible.

• Diagnostic genetic testing will be allowed once per lifetime per condition. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

*Parent or sibling must also be a covered member under the same health plan.
Criteria: Test-specific Policies

Test-specific policies are available for some tests designed to diagnosis non-cancer conditions. Please see the policy manual for a list of test-specific policies. For tests without a specific policy, use the General Coverage Guidance in Section 1.
Genetic Testing to Predict Disease Risk

Description

Predictive genetic testing is performed in people known to be at increased risk of developing an inherited non-cancer condition (for the purposes of this guideline) based on their family history. For some conditions, a positive genetic test predicts with certainty that the person will eventually develop signs and symptoms of a condition. For other conditions, a positive genetic test result indicates an increased risk (susceptibility) for a condition. A negative result may rule out a condition, or lower the risk significantly. Having test results may improve medical management through improved screening, preventive measures, prophylactic medication, and other means.

- This guideline does not include testing of a symptomatic individual. Please refer to Genetic Testing to Diagnose Non-Cancer Conditions for that purpose.
- Predictive testing for hereditary cancer syndromes is addressed separately under Genetic Testing for Cancer Susceptibility and Hereditary Cancer Syndromes.

Criteria

Criteria: General Coverage Guidance

Individuals may be considered for predictive genetic testing when ALL of the following conditions are met:

- The individual is known to be at-risk for developing inherited condition because a parent, sibling, or child is affected by or known to be a carrier of a genetic disease.
- Technical and clinical validity: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.
- Clinical utility: Healthcare providers can use the test results to provide significantly better medical care for the individual.
- Reasonable use: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Limits:

- Testing will be considered only for the number of genes or tests necessary to establish carrier status. A tiered approach to testing, with reflex to more detailed testing and/or different genes, will be required when clinically possible.
- Predictive genetic testing will be allowed once per lifetime per condition. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.
• Predictive testing will be considered only for adult individuals (age 18 and over). Exceptions may be considered if there are medical management and/or significant psychosocial benefits to testing prior to adulthood.¹,²

Criteria: Special circumstances

Testing for Known Familial Mutations

The genetic mutation(s) associated with a genetic disease can often be defined in an affected family member, allowing for testing of at-risk relatives for those specific mutations. Testing for known familial mutations may be considered when ALL of the following conditions are met:

- The mutations in the family have been clearly defined by previous genetic testing and information about those mutations can be provided to the testing lab.
- Technical and clinical validity: The test must be accurate, sensitive and specific to the familial mutations.
- Clinical utility: Healthcare providers can use the test results to provide significantly better medical care for the individual.
- Reasonable use: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Limits:

- Testing will be considered only for the known familial mutations when clinically possible.
- Predictive genetic testing will be allowed once per lifetime per condition.
- Predictive testing will be considered only for adult individuals (age 18 and over). Exceptions may be considered if there are medical management and/or significant psychosocial benefits to testing prior to adulthood.¹,²

Criteria: Test-specific Policies

Test-specific policies are available for some tests designed to predict disease risk. Please see the policy manual for a list of test-specific policies. For tests without a specific policy, use the General Coverage Guidance.

References

# Hereditary (Germline) Testing After Tumor (Somatic) Testing

## Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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### What is germline hereditary cancer testing following somatic tumor testing

#### Definition

Most cancer is sporadic and due to the acquisition of somatic variants. In addition, up to 10% of cancer has a hereditary etiology due to constitutional germline variants.¹

- In oncology, next generation sequencing (NGS) technology makes it feasible to catalog the DNA sequence variations within a person’s cancer (i.e., somatic mutation profiling). This helps define therapeutic targets which might improve outcomes through the use of specific medications directed at those mutations.²
- Germline variants can also be identified as an ancillary finding during primary tumor profiling to identify somatic mutations. “In the course of analyzing tumor DNA (without matched normal DNA), sequencing can identify constitutional (germline) DNA variations that are associated with disease or susceptibility to disease as well as carrier states for Mendelian disorders. Centers may use matched tumor-normal
sequencing to facilitate more accurate calling of somatic mutations by using the normal DNA to exclude germline variants from the tumor cells.”

- In a study by Schrader et al, “Targeted tumor sequencing with a panel of 341 genes and matched normal DNA in 1566 individuals with advanced malignant neoplasms revealed presumed pathogenic germline variants (PPGVs) in about 16% of individuals. Most PPGVs (80.5%, 95% CI, 75.1%-85.0%) were in genes related to cancer susceptibility. The PPGVs in genes previously designated as clinically actionable cancer targets were seen in 5.0% (95% CI, 4.1%-6.2%) of individuals. Most cancer-susceptibility PPGVs were retained in the tumor (91.9%; 95% CI, 87.3%-95.0%). This study is in line with other published studies investigating the prevalence of incidental findings with somatic tumor profiling.”

- The debate continues regarding whether there is an obligation to test for and report these germline findings which are secondary to the original purpose of somatic tumor profiling. In making this determination, pre-test informed consent is of utmost importance. “Honoring patient preferences requires oncology providers to communicate the potential for incidental and secondary germline information specific to the test being offered, the relevance and potential benefits of this information for patients and their relatives, and the limitations and risks of receiving incidental and secondary germline information.”

**Test information**

- Testing to investigate somatic and constitutional DNA variants has become more common as sequencing technology has evolved from the more labor intensive Sanger sequencing to NGS. “NGS is a powerful technology that permits the characterization of large amounts of DNA sequence much quicker and at lower cost than traditional Sanger sequencing.”

- Laboratories performing somatic mutation profiling may include paired germline testing, not in an effort to identify hereditary etiologies but to identify variants that are genetic “drivers” of the individual’s malignancy.

- Laboratories may also use bioinformatics to subtract the inherited variants from the somatic tumor profiling findings. Germline variants may be missed during this process without performing further analysis.

**Guidelines and evidence**

- The National Comprehensive Cancer Network (NCCN, 2017) states the following regarding germline testing following somatic tumor testing for BRCA1/2 mutations.

  - BRCA1/2 germline mutation testing should be performed when a BRCA1/2 mutation is detected by tumor profiling.
There have been various peer-reviewed publications that reviewed pre- and post-test considerations for germline testing following somatic tumor testing.

**Pre-test considerations:**

- Somatic tumor-only NGS testing is used to guide treatment for an affected person. The testing is not designed to elucidate a hereditary etiology. A germline variant may not be detected (due to differences in coverage in the testing, cellularity of the sample, allelic loss of the germline mutation) or may not be reported by the somatic testing laboratory.\(^2,3,10\)

- Directed germline genetic testing can be ordered to identify a potential hereditary etiology for the person’s tumor. Referrals to oncology genetic counselors or other specialized healthcare providers should occur if the individual’s personal and/or family history meets established criteria to warrant a more detailed discussion.\(^9,11,12\)

- Ancillary findings from somatic or germline testing may include variants in genes that cause a hereditary cancer syndrome, a non-oncologic hereditary syndrome, or identify carrier status for Mendelian disease. Specific findings are dependent on specific testing performed by the laboratory.\(^2,3,10\)

- Many patients undergoing somatic tumor profiling have advanced stage disease. Centers performing somatic tumor profiling should consider obtaining a surrogate individual to receive results in the event that the proband has passed away or is otherwise unable to receive the results.\(^2,3,10\)

**Post-test considerations:**

- Clinicians must determine the technical specifications of the laboratory used for somatic tumor profiling and determine if this includes paired germline testing. Some laboratories may not report germline variants.\(^2,3,13\)

- Tumor profiling variant interpretation may differ from the variant interpretation process for germline mutations. For example, a laboratory profiling a somatic tumor may classify a certain variant as pathogenic whereas a laboratory testing a germline mutation may classify that same variant as a variant of uncertain significance (VUS).\(^2,3,13\)

- Referrals to oncology genetic counselors or other specialized healthcare providers should occur if the individual’s personal and/or family history meets established criteria to warrant a more detailed discussion, regardless of somatic tumor profiling results.\(^9,11,12\)

**Criteria**

- Requests for single-site or full-gene sequence germline tumor testing following somatic tumor analysis will be considered medically necessary when at least one of the following criteria is met:
The individual's personal or family history is suggestive of a germline mutation, a specific germline variation is identified by somatic tumor testing, and the individual meets the published test-specific criteria to test for that variant, OR

One of the identified variants is a highly-recurrent or founder mutation (i.e., BRCA1 c185delAG or the recurrent inversion of MSH2 seen in some families with Lynch syndrome), OR

The tumor profile shows thousands of somatic variants, suggesting a germline mutation in a DNA mismatch repair gene or in the POLE proofreading domain, OR

Two separate primary tumors are sequenced and both harbor the same genetic variant, OR

The individual's tumor harbors a mutation in BRCA1/2, OR

Patient does not meet published criteria for germline testing, but variant(s) in genes that commonly harbor somatic mutations (including but not limited to TP53, APC, CDH1) are identified and the variant allele frequency in the tumor is at least 50%.

Exclusions and other considerations

- Germline testing of somatic variants of uncertain significance (VUS) is not considered medically necessary.
- Germline testing for asymptomatic individuals based solely on a family member’s somatic testing result is not considered medically necessary.
- In individuals meeting criteria for germline DNA testing, analysis of the entire gene, as opposed to single site testing, is recommended.
- Resources, such as ClinVar, should be used by the provider to determine if a pathogenic variant classification provided by germline testing laboratories is consistent with independent assessments of that variant.

References


Proceedings of the 105th Annual Meeting of the American Association for Cancer Research; 2014 Apr 5-9; San Diego, CA. Philadelphia (PA): AACR; Cancer Res. 2014;74(19 Suppl):Abstract nr 1893. Available at: http://cancerres.aacrjournals.org/content/74/19_Supplement/1893

Investigational and Experimental Molecular and Genomic Testing

Introduction

Investigational and experimental (I&E) molecular and genomic testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures address by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
</table>
| Investigational and experimental tests that make use of molecular and genomic technologies | 81479  
84999  
81599, and  
Others |

What is I&E molecular and genomic testing

Definition

An investigational and experimental (I&E) procedure is the use of a service, supply, drug, or device that is not recognized as standard medical care for the condition, disease, illness, or injury. Treatment is determined by the health plan based on an independent, peer review of literature and scientific data. I&E molecular and genomic tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact.

Investigational and experimental determinations

Molecular and genomic tests are routinely released to market that make use of novel technologies or have a novel clinical application. These tests are often available on a clinical basis long before the required evidence to support clinical validity and clinical utility are established. Typically, there is insufficient data to support that the test

- accurately assesses the outcome of interest, analytical and clinical validity
- significantly improves health outcomes, clinical utility, and
• performs better than an existing standard of care medical management option.

Because these tests are often proprietary, there may be no independent test evaluation data available in the early stages to support the laboratory's claims regarding test performance and utility.

As new molecular and genomic tests become commercially available, the evidence base is reviewed. Tests determined to be I&E by the Health Plan are addressed by this guideline or a test-specific guideline and are not eligible for reimbursement.

FDA clearance

In the case of molecular and genomic testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight. FDA clearance often does not assess clinical utility.

Criteria

Introduction

This sectioncatalogues some, but not all, molecular and genomic tests that have been determined to be investigational and experimental (I&E). I&E tests may also be addressed in test-specific guidelines and the reader is referred to those documents for additional information. New I&E tests may not yet be specifically listed in this guideline, but such decisions will be made using the following criteria.

Criteria: General Coverage Guidance

Molecular and genomic tests are only eligible for reimbursement when ALL of the following conditions are met:

• Technical and clinical validity: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test.
• Clinical utility: Healthcare providers can use the test results to provide significantly better medical care for the individual.
• Reasonable use: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

Novel oncology molecular and genomic tests

The following tests used in the screening, diagnosis, prognostication, and/or treatment decision-making for various neoplasms do not meet the above criteria and are not eligible for reimbursement.

Gene Expression Assays

• BluePrint Molecular Subtyping Profile [Proprietary 80-gene expression signature to classify Basal-type, Luminal-type and ERBB2-type breast cancers from Agendia]
• BreastOncPx [Proprietary 14-gene signature to assess breast cancer prognosis from LabCorp]
• ColonSentry [Proprietary 7-gene signature to detect colorectal cancer from Innovative Diagnostic Laboratory]
• ColoPrint [Proprietary 18-gene signature to assess colon cancer recurrence risk from Agendia]
• DecisionDx- Cutaneous Melanoma assay [Proprietary 31-gene signature to assess melanoma metastatic risk from Castle Biosciences]
• ExosomeDx®Prostate(IntelliScore) [Oncology (prostate) gene expression profile by real-time RT-PCR of 3 genes (ERG, PCA3, and SPDEF), urine, algorithm reported as risk score from Exosome Diagnostics, Inc.] CPT 0005U
• GeneFx Colon assay [Proprietary 482-gene signature to assess 5 year colon cancer recurrence risk from Helomics]
• myPath Melanoma [Proprietary 23-gene expression assay to assess the risk of malignant melanoma when a result cannot be obtained by clinical assessment and/or histopathology alone from Myriad Genetics]
• NexCourse Breast [Proprietary IHC assay of hormone receptor (ER/PR), HER2, and Ki-67 to determine recurrence risk from Genoptix]
• OncoDefender - CRC [Proprietary 5-gene expression assay to predict recurrence risk in early stage colorectal cancer within 3 years after surgery from Everist Genomics]
• OncoTarget/OncoTreat [Oncology, RNA, gene expression by whole transcriptome sequencing, formalin-fixed paraffin embedded tissue or fresh frozen tissue, predictive algorithm reported as potential targets for therapeutic agents from Columbia University Department of Pathology and Cell Biology, Darwin Health] CPT 0019U
• Pervenio Lung NGS Test [Proprietary 25-gene expression assay for risk stratification of early stage NSCLC from Life Technologies]

Other Novel Assays

• DEPArray [Oncology (breast cancer), ERBB2 (HER2) copy number by FISH, tumor cells from formalin fixed paraffin embedded tissue isolated using image-based dielectrophoresis (DEP) sorting, reported as ERBB2 gene amplified or non-amplified from PacificDx] CPT 0009U
• HERmark Breast Cancer Assay [Proprietary test designed to evaluate Her-2 total proteins in formalin-fixed, paraffin-embedded (FFPE) tissue specimens from LabCorp]
• Knowerror [Proprietary test for DNA based specimen provenance confirmation from Strand Diagnostics]
• MatePair Targeted Rearrangements, Oncology, [Oncology (solid organ neoplasia), gene rearrangement detection by whole genome next-generation sequencing,
DNA, fresh or frozen tissue or cells, report of specific gene rearrangement(s) from Mayo Clinic CPT 0013U

- MatePair Targeted Rearrangements, Hematologic, [Hematology (hematolymphoid neoplasia), gene rearrangement detection by whole genome next generation sequencing, DNA, whole blood or bone marrow, report of specific gene rearrangement(s) from Mayo Clinic] CPT 0014U

- mi-KIDNEY Cancer Test [Proprietary microRNA-based assay that differentiates 4 main histological types of primary kidney tumors from Rosetta Genomics]

- mi-LUNG Cancer Test [Proprietary microRNA-based assay that identifies four main subtypes of lung cancer from Rosetta Genomics]

- miRlnform Pancreas Test [Proprietary score based on expression levels of seven microRNAs to differentiate pancreatic ductal adenocarcinoma from chronic pancreatitis from Asuragen]

- PancraGen (previously PathFinderTG) [Proprietary topographic genotyping assay to be used when a definitive pathologic diagnosis cannot be made from RedPath Integrated Pathology]

- Prostate Core Mitomic Test [Proprietary test using mitochondrial DNA to detect prostate cancer not identified by standard biopsy pathology from MDNA Life Sciences]

- ProstaVysion [Proprietary panel of two biomarkers designed to predict prostate cancer prognosis from Bostwick Laboratories]

**Cardiovascular molecular and genomic tests**

The following tests used to predict cardiovascular disease and/or direct therapy do not meet the above criteria and are not eligible for reimbursement.

- 4q25-AF Risk Genotype Test (rs2200733 allele)
- 9p21 Genotype Test (rs10757278 and rs1333049 alleles)
- Apolipoprotein E Genotype (APOE)
- C-GAAP (Clopidogrel Genetic Absorption Activation Panel) [Proprietary test from Transgenomic Lab, includes ABCB1 and CYP2C19 gene variants]
- KIF6 Genotype Test
- LPA-Aspirin Genotype Test(4399Met allele)
- LPA-Intron 25 Genotype Test
- PAI-1 Testing for Cardiovascular Disease Risk Assessment
• Statin Induced Myopathy Genotype (SLCO1B1) CPT 81328

**Gene variant or marker risk assessment tests**

The following tests that make use of inherited genomic information to assess disease risk, prognosis, or subtyping do not meet the above criteria and are not eligible for reimbursement.

• ARISk Autism Risk Assessment Test [Proprietary test from IntegraGen]
• BREVAGen [Proprietary sporadic breast cancer risk based on genetic markers from Phenogen Sciences]
• Cardiac DNA Insight [Proprietary test from Pathway Genomics that assesses genetic markers for cardiac-related conditions]
• Crohn's prognostic test [NOD2/CARD15 gene variant testing]
• IBD sgi Diagnostic [Proprietary test from Prometheus with genomic components including ATG16L1, STAT3, NKX2-3, and ECM1 gene variants.]
• LactoTYPE [Proprietary test from Prometheus that assesses the hypolactasia C/T genetic variant]
• Macula Risk [Proprietary test from ArcticDx to predict risk of age-related macular degeneration progression]
• Pathway Fit [Proprietary test from Pathway Genomics that focuses on metabolism, diet, and exercise traits]
• RetnaGene AMD [Proprietary test from Sequenom CMM to predict risk of wet AMD progression]

**Whole genome sequencing**

• CPT codes 81425, 81426, 81427

**Pharmacogenomic panels**

• 5-Fluorouracil (5-FU) Toxicity and Chemotherapeutic Response [Proprietary panel of DPYD and TYMS gene variants to assess risk of 5-fluorouracil toxicity from ARUP Laboratory] CPT 81232 and 81346
• Genecept Assay [Proprietary panel of biomarker tests to predict response to different psychiatric treatments from Genomind]
• GeneSightRx ADHD [Proprietary test from AssureRx assessing three genes]
• GeneSightRx Analgesic [Proprietary test from AssureRx assessing two genes]
• GeneSightRx Psychotropic [Proprietary test from AssureRx assessing six genes]
• Mental Health DNA Insight [Proprietary test from Pathway Genomics]
• OneOme RightMed, [Drug metabolism (adverse drug reactions), DNA, 22 drug metabolism and transporter genes, real-time PCR, blood or buccal swab, genotype and metabolizer status for therapeutic decision support from OneOme, LLC]

• Pain Medication DNA Insight [Proprietary test from Pathway Genomics]

**Non-cancer gene expression assays**

• Renal Transplant Monitoring (FOXP3, Granzyme B, Perforin, IP10) [Gene expression panel that is an indirect indicator of immune response designed to detect or monitor renal transplant rejection from Quest Diagnostics]

**Infectious disease assays**

• AmHPR Helicobacter pylori Antibiotic Resistance Next Generation Sequencing Panel, [Helicobacter pylori detection and antibiotic resistance, DNA, 16S and 23S rRNA, gyrA, pbp1, rdxA and rpoB, next generation sequencing, formalin-fixed paraffin embedded or fresh tissue, predictive, reported as positive or negative for resistance to clarithromycin, fluoroquinolones, metronidazole, amoxicillin, tetracycline and rifabutin from American Molecular Laboratories, Inc.] CPT 0008U

• Bacterial Typing by Whole Genome Sequencing, [Infectious disease (bacterial), strain typing by whole genome sequencing, phylogenetic-based report of strain relatedness, per submitted isolate from Mayo Clinic] CPT 0010U
Pharmacogenomic Testing for Drug Toxicity and Response

For the purposes of this guideline, pharmacogenomic tests are those performed to predict or assess an individual's response to therapy as well as the risk of toxicity from drug treatment. Testing may be performed prior to treatment in order to determine if the individual has genetic mutations that could affect drug response and/or increase the risk for adverse drug reactions. Testing may also be performed during treatment to assess whether an individual is having an adequate response or investigate the cause of an unexpected or adverse reaction.

Companion diagnostics are assays that help determine whether a drug may be safe or effective for a particular patient. Companion assays are evaluated as part of the Food & Drug Administration's (FDA's) development and approval process for the new drug. According to the FDA, “A companion diagnostic is a medical device, often an in vitro device, which provides information that is essential for the safe and effective use of a corresponding drug or biological product. The test helps a health care professional determine whether a particular therapeutic product’s benefits to patients will outweigh any potential serious side effects or risks.”  Although specific companion diagnostic tests may be identified in the FDA label for a new drug approval, similar laboratory-developed tests (LDTs) performed by a CLIA-certified laboratory are generally accepted as alternatives that can typically provide the required information.

Complementary diagnostics are assays that were developed and in use prior to the FDA's approval of a new drug. They are not evaluated through the FDA's development and approval process for new drugs. Complementary diagnostics are used to help provide additional information about how a drug might be used, or whether someone should receive a certain class of drugs. These tests are not specifically required for the safe and effective use of a drug, which is part of what differentiates them from companion diagnostics. As with companion diagnostics, LDTs that are similar to the defined complementary diagnostic, when performed by a CLIA-certified laboratory, are able to provide the same information.

Criteria

Criteria: General Coverage Guidance

Pharmacogenomic tests may be indicated when ALL of the following conditions are met:

- The individual is currently taking or considering treatment with a drug potentially affected by a known mutation that can be detected by a corresponding test.
- Technical and clinical validity: The test must be accurate, sensitive, and specific, based on sufficient, quality scientific evidence to support the claims of the test.
• Clinical utility: Healthcare providers can use the test results to guide changes in
drug therapy management that will improve patient outcomes.

• Reasonable use: The usefulness of the test is not significantly offset by negative
factors, such as expense, clinical risk, or social, or ethical challenges.

Criteria: Companion Diagnostic Testing

Single gene testing for purposes of medication usage will be approved when the
following criteria are met:

• Testing is being performed in a CLIA-certified laboratory, AND
• Testing is NOT being performed as part of a panel of genes, AND
• Testing of the requested gene has not previously been performed, AND
• A medication’s FDA label requires results from the genetic test to effectively or
safely use the therapy in question, AND
• Healthcare providers can use the test results to directly impact medical care for the
individual, OR
• The member meets all criteria in a test-specific guideline, if available

Criteria: Limits

Testing will be covered only for the number of genes or tests necessary to establish
drug response. When available and cost-efficient, a tiered approach to testing, with
reflex to more detailed testing and/or different genes, is recommended.

For pharmacogenomic tests that look for changes in germline DNA (i.e., not tumor DNA
or viral DNA), testing will be allowed once per lifetime per gene. Exceptions may be
considered if technical advances in testing or the discovery of novel genetic variants
demonstrate significant advantages that would support a medical need to retest.

Testing performed in a CLIA-certified laboratory will be considered for coverage. The
use of a specific FDA approved companion diagnostic is not necessary for coverage to
be considered.

Criteria: Exclusions

Some pharmacogenomic tests are considered investigational and experimental, as
they are not supported by existing evidence, professional guidelines and/or the FDA, or
their use in medical management is deemed to be still evolving.

The following pharmacogenomic tests are considered investigational and
experimental. This list is not intended to be all inclusive.

• 5HT2C (Serotonin Receptor) gene variants
• Ankyrin G gene variants
• COMT (Catechol Methyl Transferase) gene variants
• Catechol-O-Methyltransferase (COMT) Genotype from Mayo Clinic (CPT 0032U)
• CYP450 gene variants (including, but not limited to CYP1A2, CYP2D6, CYP2C9, CYP2C19, CYP3A4 [CPT 81230], CYP3A5 [CPT 81231]) for psychotherapeutic, cardiovascular, or general drug response
• Cytochrome P450 1A2 Genotype from Mayo Clinic (CPT 0031U)
• CYP2C19 testing for the management of H. pylori
• DRD2 (Dopamine Receptor) gene variants
• Focused Pharmacogenomics Panel from Mayo Clinic (CPT 0029U)
• IFNL3 rs12979860 gene variant (CPT 81283)
• KIF6 gene variants
• KRAS testing for response to erlotinib in non-small cell lung cancer
• MTHFR gene variants
• NAT2 gene variants
• OPRM1 gene variants
• Serotonin Receptor Genotype (HTR2A and HTR2C) from Mayo Clinic (CPT 0033U)
• SLC6A4 (5-HTTLPR) serotonin transporter variants
• Thiopurine Methyltransferase (TPMT) and Nudix Hydrolase (NUDT15) Genotyping from Mayo Clinic (CPT 0034U)
• Warfarin Response Genotype from Mayo Clinic (CPT 0030U)

Criteria: Test-specific Guidelines

Test-specific guidelines are available for some pharmacogenomic tests. Please see the guidelines manual for a list of test-specific guidelines. For tests without a specific guideline, use the General Coverage Guidance provided.

References

1. Companion diagnostics. U.S. Food & Drug Administration website. Available at: https://www.fda.gov/MedicalDevices/ProductsandMedicalProcedures/InVitroDiagnostics/ucm407297.htm


Preimplantation Genetic Screening and Diagnosis

Description

Preimplantation Genetic Diagnosis (PGD) and Preimplantation Genetic Screening (PGS) are used to detect genetic conditions, chromosome abnormalities, and fetal sex during assisted reproduction with in vitro fertilization (IVF). PGD refers to embryo testing that is performed when one or both parents have a known genetic abnormality. This includes single-gene mutations and chromosome rearrangements. PGS refers to screening an embryo for aneuploidy when both parents are chromosomally normal. Genetic testing is performed on cells from the developing embryo prior to implantation. Only those embryos not affected with a genetic condition are implanted. PGD may allow at-risk couples to avoid a pregnancy affected with a genetic condition. The use of PGS has been reviewed by the American College of Obstetricians and Gynecologists (ACOG). The Society for Assisted Reproductive Technology and the American Society for Reproductive Medicine published a joint practice committee opinion to address the safety, accuracy, and overall efficacy of PGD and PGS.

- This guideline does not include prenatal or preconception carrier screening. Please refer to Genetic Testing for Carrier Status for that purpose.
- This guideline does not include prenatal genetic testing. Please see Genetic Testing for Prenatal Screening and Diagnostic Testing for genetic testing done during pregnancy.

Criteria

Criteria: General coverage guidance

Preimplantation genetic diagnosis may be considered when ALL of the following conditions are met:

- **Technical and clinical validity**: The test must be accurate, sensitive and specific, based on sufficient, quality scientific evidence to support the claims of the test. In the case of PGD, the mutation(s) or translocation(s) to be tested in the embryo should first be well-characterized in the parent(s) AND the embryonic test results must be demonstrated to be highly accurate.

- **Clinical utility**: Healthcare providers can use the test results to provide significantly better medical care and/or assist patients with reproductive planning.

- **Reasonable use**: The usefulness of the test is not significantly offset by negative factors, such as expense, clinical risk, or social or ethical challenges.

AND THE FOLLOWING APPLY:
• The couple is known to be at-risk to have child with a genetic condition because of ANY of the following:
  
  o Both parents are known carriers of a recessive genetic condition; OR
  o One parent is affected by or known to be a carrier of a dominant condition; OR
  o The female contributing the egg is known to be a carrier of an X-linked condition; OR
  o One or both parents are carriers of a structural chromosome rearrangement (e.g., translocation or inversion); OR
  o One or both parents have a known chromosome microdeletion (e.g. 22q11 deletion – DiGeorge syndrome, 7q11.23 deletion – Williams syndrome);

  AND

• The genetic condition is associated with potentially severe disability or has a lethal natural history.

---

**Note**  This guideline ONLY addresses the genetic testing component of PGS or PGD. Coverage of any procedures, services, or tests related to assisted reproduction is subject to any applicable plan benefit limitations.

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**Criteria: Special circumstances**

**Sex determination**

• PGD for sex (X and Y chromosome testing) is considered medically necessary only for identification of potentially affected embryos for gender-related conditions.

**HLA typing**

• PGD for human leukocyte antigen (HLA) typing for transplant donation is considered medically necessary only if:
  
  o A couple has child with a bone marrow disorder needing a stem cell transplant; AND
  o The only potential source of a compatible donor is an HLA-matched sibling

**Chromosome abnormality screening**

• PGS for de novo chromosome abnormalities is not considered medically necessary. This includes the following indications:¹
  
  o Maternal age alone
  o To improve in vitro success rates
For recurrent unexplained miscarriage and/or recurrent implantation failures

References


Molecular and Genetic Test Specific Guidelines
ABL Tyrosine Kinase Sequencing for Chronic Myeloid Leukemia

Introduction

ABL tyrosine kinase sequencing for chronic myeloid leukemia is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
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<tr>
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<tr>
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<td>81401</td>
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<tr>
<td>ABL1 Tyrosine Kinase Domain Sequencing</td>
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What are CML and BCR-ABL

Definition

Chronic myelogenous leukemia (CML) is a hematopoietic stem cell disease that results in overgrowth of white blood cells in the bone marrow. It is defined by the presence of the Philadelphia chromosome (Ph), a translocation between chromosomes 9 and 22 that results in the fusion of two genes known as BCR and ABL.1,2 Acute lymphoblastic leukemia (ALL) is a different form of leukemia, but may also be positive for the Philadelphia chromosome (Ph+). About 3% of pediatric ALL and 25% of adult ALL is Ph+.3

Diagnosis

Detection of the BCR-ABL fusion gene is diagnostic for CML and Ph+ ALL and can be established by fluorescent in situ hybridization (FISH) or quantitative real-time polymerase chain reaction (qPCR).2

Symptoms

The three phases of CML are chronic, accelerated and blastic. In the chronic phase, there are few symptoms and most people are diagnosed after a routine blood test reveals the characteristic blood count and differential. If not treated, the disease will
progress to the accelerated and blastic phases, symptoms of which include fever, bone pain, splenomegaly, fatigue and weakness.¹

**Treatment**

First-line treatment for CML and some Ph+ ALL is with a class of drugs called tyrosine kinase inhibitors (TKIs), which block the production of the BCR-ABL fusion gene protein product. Three TKI therapies are available as first-line therapies: imatinib (Gleevec®), nilotinib (Tasigna®), and dasatinib (Sprycel®). These TKI therapies have all demonstrated proven benefit, and median survival is expected to approach normal life expectancy for most patients with CML.¹,²

**Monitoring**

Monitoring of patients for treatment response to TKIs includes routine measurement of the BCR-ABL fusion gene protein product via qPCR prior to initiation of treatment and during treatment every 3 months. After CCyR has been achieved, measurement of the BCR-ABL fusion gene product is recommended every 3 months for 2 years and every 3 to 6 months thereafter.²

**Treatment resistance**

For individuals who display apparent treatment resistance, consideration of alternative treatment options (or enrollment in a clinical trial) may be appropriate.² Treatment resistance in both CML and ALL can be caused by mutations in the BCR-ABL kinase domain.²,³

**Test information**

**Introduction**

Testing for CML ABL1 tyrosine kinase domain may include targeted mutation analysis or sequence analysis.

**Genetic testing**

ABL1 tyrosine kinase domain mutation analysis is performed on a blood or bone marrow aspirate sample. Testing is performed by either

- targeted mutation analysis for specific resistance variants, such as T315I,
• sequencing of the entire ABL1 tyrosine kinase domain.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to when BCR-ABL kinase domain analysis should be performed.

National Comprehensive Cancer Network

The National Comprehensive Cancer Network (NCCN, 2017)\(^2\) for CML states:

• BCR-ABL kinase domain analysis should be performed when:
  o "Chronic phase:
    ▪ Inadequate initial response to TKI therapy (lack of PCyR or BCR-ABL1 > 10\% (IS) at 3 and 6 months or less than a CCyR or BCR-ABL1 > 1\% (IS) at 12 months).
    ▪ Any sign of loss of response (defined as hematologic or cytogenetic relapse)
    ▪ 1-log increase in BCR-ABL1 transcript levels and loss of MMR
  o Disease progression to accelerated or blast phase.”

• “Mutational analysis is helpful in the selection of subsequent TKI therapy for patients with inadequate initial response to first-line or second-line TKI therapy. Mutational analysis would also be helpful to identify a subgroup of patients who demand careful monitoring (as these patients are at a higher risk of progression) and the subset of patients who will be eligible for allogeneic HSCT.”

• These recommendations are category 2A: “based on lower-level evidence and there is non-uniform NCCN consensus (but no major disagreement)”

The National Comprehensive Cancer Network (NCCN, 2017)\(^3\) for ALL states:

• ABL gene mutation testing should be considered for all Ph+ ALL in adolescents, young adults, and adults (AYA).

• These recommendations are category 2A: “based on lower-level evidence and there is non-uniform NCCN consensus (but no major disagreement)”

Criteria

Introduction

Requests for ABL Tyrosine Kinase analysis will be reviewed using these criteria.
Criteria

BCR-ABL kinase domain mutation analysis is indicated in:

- Individuals with CML who have:
  - Inadequate initial response to TKI therapy (lack of PCyR or BCR-ABL1 > 10% (IS) at 3 and 6 months or less than a CCyR or BCR-ABL1 > 1% (IS) at 12 months), or
  - Any sign of loss of response (hematologic or cytogenetic relapse), or
  - A 1-log increase in BCR-ABL1 transcript levels and loss of MMR, or
  - Disease progression to accelerated or blast phase, OR
- Individuals with Ph+ ALL.

Note: BCR-ABL kinase domain mutation analysis is not indicated in other cancer types for which tyrosine kinase inhibitor therapy may be considered.

References

Introduction

These references are cited in this guideline.

## Acute Myeloid Leukemia (AML) Genetic Testing

**Procedures addressed**

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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## What is Acute Myeloid Leukemia

### Definition

Acute myeloid leukemia (AML) is a neoplasm resulting from the clonal expansion of myeloid blasts in the peripheral blood (PB), bone marrow (BM), or other tissues. It is a heterogeneous disease clinically, morphologically and genetically.

- The required blast percentage for a diagnosis of AML is \( \geq 20\% \) myeloblasts and/or monoblasts/promonocytes and/or megakaryoblasts in the PB or BM. The diagnosis of AML can also be made when the blasts percentage is <20\% if the increase blast count is associated with: \( t(8;21)(q22;q22.1) \), \( \text{inv}(16)(p13.1q22) \) or \( t(16;16)(p13.1;q22) \), or \( t(15;17)(q24.1;q21.2) \).

- A large number of recurrent cytogenetic abnormalities and mutated genes are recognized in AML. Some of these genetic abnormalities are associated with unique phenotype and prognostic features and are classified under acute myeloid leukemia with recurrent genetic abnormalities. The AMLs with no recurrent genetic abnormalities are classified under acute myeloid leukemia, not otherwise specified.

- In AML, the pretreatment cytogenetic abnormalities represent the single most important prognostic factor for predicting remission rates, relapse risk, and overall survival rate.\(^1\) Even in de novo AMLs with no chromosomal abnormalities the clinical outcome is heterogeneous.
• Studies has shown that molecular abnormalities including NPM1, FLT3-ITD, CEBPA, IDH1/2, DNMT3A, KIT and other mutations are important for prognostication not only in AMLs with a normal karyotype but also in other AML subsets.2-5 Some of these molecular abnormalities also affect the choice of treatment for patients with AML.

• After treatment is selected and initiated, treatment response can be monitored by assessing the blast cell percentage in bone marrow using morphology and immunophenotyping. Individuals who have chromosomal abnormalities at initial diagnosis are monitored for disappearance (indicates remission) and re-emergence (indicates relapse) of these abnormalities.

Test information

• The various diagnostic modalities that are utilized for front-end diagnostics are specified by the National Comprehensive Cancer Network (NCCN, 2017).6
  o These components include standard morphologic examinations on the blood and bone marrow, immunohistochemical staining, cytogenetics, flow cytometry, etc. However, this policy only pertains to those molecular techniques, which have either prognostic and/or predictive (that is, influencing chemotherapy selection with improved outcomes) implications.

• The specific methodology used to identify molecular markers is dependent upon the type of marker being investigated.
  o DNA mutations are generally detected through direct analysis of hotspots or sequencing either parts of the gene or the whole gene.
  o Chromosome abnormalities, such as translocations or deletions, may be detected through direct visualization of the chromosomes (karyotyping), in situ hybridization of probes (e.g., FISH) to detect deletions or duplications that are too small to see directly, or DNA-based methods that identify deletions or translocation breakpoints.
  o Gene expression profiling simultaneously measures the amount of RNA or protein being made by many genes. Expression patterns may be used to predict the type of cancer present, tumor aggressiveness, and therapy needs.
  o Whole exome sequencing is used to identify multiple molecular markers, at the same time, which may be present in a tumor.

• The efficiency of next generation sequencing (NGS) has led to an increasing number of large, multi-gene molecular marker panels. Given that hematological malignancies can have multiple and unexpected genetic changes, these panels may provide physicians with information about therapeutic targets that would not otherwise be considered.

Guidelines and evidence

• The National Comprehensive Cancer Network (NCCN, 2017) states the following in regards to genetic testing in individuals with AML.6
“The ability to identify mutations that carry prognostic impact is increasing with the use of molecular profiling. Thus, in addition to basic cytogenetic analysis, new molecular markers can help refine prognostic groups, particularly in patients with a normal karyotype. These markers include NPM1, FMS-like tyrosine kinase 3 (FLT3), CEBPA, isocitrate dehydrogenase 1 and 2 (IDH1/2), DNA (cytosine-5)-methyltransferase 3A (DNMT3A), and KIT gene mutations. Tests for these molecular markers are becoming more common in commercial reference laboratories and in referral centers. Therefore, it is important for physicians to confer with the local pathologists on how to optimize sample collection from the time of diagnosis for subsequent molecular diagnostic tests. Testing for additional mutations may also be recommended.”

- Some FDA labels require results from molecular marker tests to effectively or safely use the therapy for a specific cancer type. A list of all Pharmacogenomic Biomarkers included in FDA labeling and associated implications can be found here.
- Whereas the above mentioned biomarkers constitute established evaluation pathways for AML, there are many emerging mutations that might have clinical relevance to various types of AML that are not yet considered standard of care.

Criteria

Introduction

Medical necessity criteria differ based on the type of testing being performed (i.e., individual tumor markers separately chosen based on the cancer type versus pre-defined panels of tumor markers).

Single gene testing for AML

- The member has AML and will benefit from information provided by the requested molecular marker test based on at least one of the following:
  - An oncology therapy FDA label requires results from the marker test to effectively or safely use the therapy for the member’s AML, or
  - NCCN guidelines include the tumor marker test in the management algorithm for AML and all other requirements are met (specific pathology findings, staging, etc.); however, the tumor marker must be explicitly included in the guidelines and not simply included in a footnote as an intervention that may be considered, or
The NCCN Biomarker Compendium has a level of evidence of at least 2A for the tumor marker’s application to AML

**Panel testing for AML**

Gene panels that are specific to hematological cancers and include the following genes will be eligible for reimbursement according to the criteria outlined in this policy: NPM1, FLT3, CEBPA, IDH1, IDH2, DNMT3A, KIT and TP53. This sequencing panel will only be considered for reimbursement when billed with the appropriate panel CPT code: 81450.

- Genetic Counseling
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous panel testing for AML, AND
- Diagnostic Testing for Symptomatic Individuals:
  - Member has a diagnosis of AML, and
  - The results of the test will directly impact the diagnostic and treatment options that are recommended for the patient, AND
- Rendering laboratory is a qualified provider for service per Health Plan policy

**Billing and reimbursement considerations**

- When multiple CPT codes are billed for components of a panel and there is a more appropriate CPT code representing the panel, eviCore will redirect to the panel code(s).
- Panels of over 50 genes billed with CPT code 81455 are considered excessive in individuals with AML and will not be reimbursed.

**References**


7. US Food and Drug Administration. Table of Pharmacogenomic Biomarkers in Drug Labeling. Available at: http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm
Afirma Gene Expression Classifier for Thyroid Cancer

Introduction

Afirma Gene Expression Classifier testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan’s procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Afirma Gene Expression Classifier</td>
<td>81545</td>
</tr>
<tr>
<td>BRAF V600 Targeted Mutation Analysis</td>
<td>81210</td>
</tr>
</tbody>
</table>

What is the Afirma Gene Expression Classifier for thyroid cancer

Definition

The Afirma® Gene Expression Classifier (GEC) is a test that uses fine needle aspiration (FNA) samples for determining the risk of malignancy in thyroid nodules previously diagnosed as cytologically indeterminate that would otherwise be recommended for diagnostic thyroid surgery.¹

Indications

Palpable thyroid nodules are often evaluated using fine needle aspiration (FNA) to rule out malignancy. In 15-30% of cases, the result is indeterminate: not clearly benign or malignant.² Cytologically indeterminate nodules may then be referred for diagnostic surgery. However, 70-80% have benign results.³ ⁴

To help avoid unnecessary diagnostic surgeries, gene expression testing may be used to further characterize these nodules as benign or suspicious for cancer.

The Afirma GEC is intended for

• cytologically indeterminate FNA biopsy samples including atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS), and
• follicular or Hürthle Cell Neoplasms.⁵

When indicated, the Afirma GEC must be used in conjunction with cytopathology, ultrasound assessment, and other clinical factors to determine an individual’s risk of thyroid cancer and the necessity of thyroid surgery.⁵

Results

Afirma GEC results correlate with the postoperative surgical pathology,⁵ which may guide the decision to observe the individual’s nodule in lieu of surgical resection.⁶

Based on the results of the Afirma GEC, additional testing may be requested or performed, such as the Afirma Malignancy Classifiers, BRAF and MTC. See the Test Information section for more information regarding these tests.

Test Information

Introduction

Afirma GEC testing may include a combination of cytopathology and gene expression testing. This guideline addresses only the gene expression testing component.

When GEC testing is performed

A FNA sample can be submitted for cytopathology assessment.

<table>
<thead>
<tr>
<th>If the cytopathology assessment is ...</th>
<th>Then ...</th>
</tr>
</thead>
<tbody>
<tr>
<td>benign or malignant</td>
<td>the analysis is complete.</td>
</tr>
<tr>
<td>indeterminate</td>
<td>the GEC is performed.</td>
</tr>
</tbody>
</table>

Afirma GEC

The Afirma GEC measures the gene expression levels of 142 genes from FNA biopsy specimens.¹ These 142 genes are correlated with histologically benign thyroid nodules that were previously diagnosed as cytologically indeterminate in two prospective multicenter clinical validation studies.⁵ ⁷ A retrospective multicenter study confirmed originally published Afirma GEC test performance.⁸

Results

There are two possible results from the Afirma GEC test:

○ **Benign** — This result has a negative predictive value of 95%, which means a risk of malignancy of 5% or less.
- **Suspicious for malignancy** — This result has a positive predictive value for malignancy of 38%. \(^1\)

### Afirma Malignancy Classifiers

The Afirma Malignancy Classifiers, BRAF and MTC, are intended to help guide surgical decisions when the cytopathology or Afirma GEC result suggests the individual should be considered for surgery. \(^9,10\)

<table>
<thead>
<tr>
<th>Test</th>
<th>Detection</th>
<th>When test is performed</th>
</tr>
</thead>
<tbody>
<tr>
<td>Afirma BRAF</td>
<td>BRAF V600E mutation</td>
<td>This test is performed when there is • malignant or suspicious for malignancy cytopathology, or • a suspicious GEC result.(^1)</td>
</tr>
<tr>
<td>Afirma Medullary Thyroid Cancer (MTC)</td>
<td>Medullary thyroid carcinoma before surgery</td>
<td>This test is performed • when there is malignant or suspicious for malignancy cytopathology, or • in conjunction with the GEC at the time of an indeterminate result.(^1)</td>
</tr>
</tbody>
</table>

### Guidelines and evidence

**Introduction**

This section includes relevant guidelines and evidence pertaining to Afirma GEC testing.

**National Comprehensive Cancer Network**

The National Comprehensive Cancer Network (NCCN, 2017) Thyroid Carcinoma Guidelines incorporate the use of molecular tests in the evaluation of indeterminate thyroid nodules (category 2B). For FNA results consistent with Follicular or Hürthle Cell Neoplasms, or atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS) with a “High clinical suspicion of malignancy”, they state:\(^6\)

“The diagnosis of follicular carcinoma or Hürthle cell carcinoma requires evidence of either vascular or capsular invasion, which cannot be determined by FNA. Molecular diagnostics may be useful to allow reclassification of follicular lesions (i.e. follicular neoplasm, atypia of undetermined significance (AUS), follicular lesions of undetermined significance (FLUS)) as either more or less likely to be benign or malignant based on the genetic profile….If molecular testing, in conjunction with clinical and ultrasound features, predicts a risk of malignancy comparable to the risk of
malignancy seen with a benign FNA cytology (approximately 5% or less), consider observation.”

Criteria

Introduction

Requests for Afirma GEC testing are reviewed using these criteria.

Afirma Gene Expression Classifier (GEC)

• Testing Multiple Samples:
  o The Afirma GEC is reimbursed only once per date of service regardless of the number of nodules submitted for testing, and
  o The Afirma GEC is indicated only once per thyroid nodule per lifetime.

• Required Clinical Characteristics:
  o Afirma GEC is indicated for thyroid nodules with indeterminate FNA results that are included in the following cytopathology categories:
    ▪ Atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS), or
    ▪ Follicular or Hürthle cell neoplasm, and
  o The patient is not undergoing thyroid surgery for diagnostic confirmation.

• Required Testing Process:
  o If FNA of a nodule is indicated to evaluate for malignancy, and the sample is sent to Veracyte for cytopathology, the gene expression classifier is only indicated when the result is indeterminate, and
  o Supporting documentation of an appropriate indeterminate cytology result will be required for payment.

Afirma Malignancy Classifiers

• Afirma MTC
  o Afirma MTC testing will be reimbursed if it is performed as part of the GEC as outlined above, and
- The Afirma MTC testing must be billed as part of the Afirma GEC. The Afirma MTC may not be billed separately using an additional unit or procedure code.

- **Afirma BRAF V600E**
  - Afirma BRAF testing may be considered for either GEC or FNA suspicious or malignant results. See *Tumor Marker Testing – Solid Tumors* guideline for criteria.
  - Afirma BRAF testing in conjunction with a GEC Indeterminate result will not be reimbursed.

## References

### Introduction

These references are cited in this guideline.


AlloMap Gene Expression Profiling for Heart Transplant Rejection

Introduction

AlloMap Gene Expression Profiling is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan’s procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedure addressed by this guideline</th>
<th>Procedure code</th>
</tr>
</thead>
<tbody>
<tr>
<td>AlloMap</td>
<td>81595</td>
</tr>
</tbody>
</table>

What is AlloMap

Definition

AlloMap is a non-invasive blood test that is designed to help identify heart transplant recipients with stable allograft function who have a low probability of moderate/severe acute cellular rejection at the time of testing.¹

Current uses

AlloMap is designed to help providers obtain this information without the use of endomyocardial biopsy. While endomyocardial biopsy is currently the standard of care for heart transplant recipients, it is an invasive procedure with associated risks.

Description

AlloMap is a panel of 20 genes. The assay uses gene expression of RNA isolated from peripheral blood mononuclear cells.¹

Results

Using data from the gene expression of these genes, an AlloMap score is calculated. The lower the score, the lower the probability of acute cellular rejection at the time of testing.¹
**Intended use**

AlloMap is intended for use in heart transplant recipients 15 years of age or older who are at least 2 months post heart transplant.¹

**Test information**

**Introduction**

The AlloMap assay measures the gene expression of RNA of 20 genes. 11 of these genes are thought to be informative for the assay, while the remaining 9 are used for quality control.¹

**Risk score**

The data collected from these genes is translated into a risk score. Scores range from 0-40 and are compared to post-transplant patients in the same post-transplant period. The lower the score, the lower the probability of acute cellular rejection at the time of testing.¹

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to AlloMap testing.

**International Society of Heart and Lung Transplantation**

The International Society of Heart and Lung Transplantation (2010)² stated the following:

“Gene Expression Profiling (AlloMap) can be used to rule out of the presence of acute cellular rejection (ACR) of grade 2R or greater in appropriate low risk patients, between 6 months and 5 years after HT.”

**Class IIa**

Class IIa: Weight of evidence/opinion is in favor of usefulness/efficacy.

Level of evidence: B – data derived from a single randomized clinical trial or large non-randomized studies.

**U.S. Food and Drug Administration (FDA)**

In 2008, the U.S. Food and Drug Administration (FDA) cleared AlloMap as a Class II Medical Device.³
**EIMAGE (Early Invasive Monitoring Attenuation through Gene Expression)**

The EIMAGE (Early Invasive Monitoring Attenuation through Gene Expression) study (2015)\(^4\) was conducted as a single-center randomized parallel 2-arm interventional study (n=60). This study compared AlloMap with heart biopsy in the first year post transplant.

- Study population consisted of 60 patients aged 18 years or older and at least 55 days post-transplant.
- Incidence of composite primary outcome in both groups was not statistically significant.
- The need for biopsy was reduced in the AlloMap monitoring group: 42 biopsies were performed in the AlloMap group vs. 253 in the biopsy group. 29 out of 42 of the biopsies performed in the AlloMap group were a direct result of the elevated AlloMap score.

**IMAGE (Invasive Monitoring Attenuation through Gene Expression) study**

The IMAGE (Invasive Monitoring Attenuation through Gene Expression) study (2010)\(^5\) serves as the first randomized, prospective trial (n=602) comparing AlloMap head-to-head with rejection monitoring by endomyocardial biopsy, the current standard of care. The study included patients who were clinically stable, 18 years of age or older, and at least 6 months post-transplant. Results of this study indicated:

- Rates of adverse events (primary outcome: rejection, graft dysfunction, death) were the same in low-risk patients monitored with AlloMap vs. traditional graft biopsy.
- The need for biopsy was reduced in the AlloMap monitoring group (since those with low scores did not get biopsies): 409 biopsies were performed in the AlloMap group vs. 1249 in the biopsy group.

However, limitations of this study were acknowledged by the authors. These limitations include:\(^5\)

- a study population that was likely significantly skewed toward patients at lower risk of rejection, since only patients who had received a cardiac transplant more than 6 months previously were eligible for enrollment
- wide statistical margins for comparing AlloMap vs. biopsy, and
- primary endpoint measures that included events that may not have been due to rejection.

The authors conclude that “gene expression profiling of peripheral blood specimens may offer a reasonable alternative to routine biopsies, for monitoring cardiac-transplant recipients for rejection if the interval since transplantation is at least 6 months and the patient is considered to be low risk for rejection.”\(^5\)
Crespo-Leiro et al

Crespo-Leiro et al (2015)\(^6\) conducted a study to examine the ability of AlloMap score variability to predict future events. They found that at a score variability of 0.6, the negative predictive value increased to 97% and the positive predictive value decreased to 23.3%. The authors concluded that “GEP score variability may be helpful in estimating probability of future events of death, re-transplantation or graft failure in heart transplant recipients.”

Deng et al

Deng et al (2014)\(^7\) conducted a study to examine the use of AlloMap score variability to predict clinical events in heart transplant recipients. They found that AlloMap score variability had a predictive accuracy of 0.69. They concluded, “the variability of gene expression profiling scores from an individual may help predict the risk of clinically defined future allograft dysfunction or death in the individual.”

Limitations of the Crespo-Leiro and Deng studies

These studies attempt to demonstrate the value of AlloMap scores for prognostic purposes. However, the studies are retrospective analyses of prospective studies and have notable limitations which marginalize their ability to effectively demonstrate clinical utility.

Criteria

Introduction

Requests for AlloMap Gene Expression Profiling are reviewed using these criteria.

Criteria

AlloMap is considered medically necessary when ALL of the following criteria are met:

- Member is not acutely symptomatic,\(^2\) and
- Member does not have recurrent rejection,\(^2\) (defined as having a documented prior rejection and currently having signs/symptoms of rejection), and
- Member is not currently receiving 20 mg or more of daily oral prednisone,\(^2\) and
- Member has not received high-dose intravenous corticosteroids or myeloablative therapy in the past 21 days,\(^2\) and
- Member has not received blood products or hematopoietic growth factors in the past 30 days,\(^2\) and
- Member is not pregnant,\(^2\) and
- Member is at least 2 months post-transplant,\(^2,4\) and
• Member is less than 5 years post-transplant, and
• Member is at least 15 years of age

**Recommended frequency of AlloMap testing**

This table describes the recommended frequency of AlloMap testing.

<table>
<thead>
<tr>
<th>Months post-transplant</th>
<th>Frequency of AlloMap testing</th>
</tr>
</thead>
<tbody>
<tr>
<td>2 to 6 months</td>
<td>every 2 to 4 weeks</td>
</tr>
<tr>
<td>6 to 12 months</td>
<td>every 2 months</td>
</tr>
<tr>
<td>12 to 24 months</td>
<td>every 3 months</td>
</tr>
<tr>
<td>24 months to 60 months</td>
<td>every 6 months</td>
</tr>
<tr>
<td>greater than 60 months</td>
<td>every 12 months</td>
</tr>
</tbody>
</table>

**Exceptions to testing frequency**

AlloMap may be used as a substitute for endomyocardial biopsy in surveillance of stable patients. Exceptions to the above testing frequencies may be considered as warranted by an individual patient’s clinical presentation, such as current signs or symptoms of rejection. AlloMap testing is not routinely covered in individuals greater than 5 years post-transplant. Requests for exceptions to this criteria will be evaluated on a case by case basis.

**Exclusions**

Coverage for AlloMap testing has some exclusions.

**Exclusion for prognostic purposes**

The use of AlloMap for prognostic purposes is specifically excluded by this guideline. Studies on the ability of the test to predict future clinical events do not provide enough evidence to warrant coverage at this time.

**References**

**Introduction**

These references are cited in this guideline.


Introduction

Alpha-1 antitrypsin deficiency (AATD) testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Protease Inhibitor (PI) Typing</td>
<td>82104</td>
</tr>
<tr>
<td>SERPINA1 Targeted Mutation Analysis</td>
<td>81332</td>
</tr>
<tr>
<td>SERPINA1 Sequencing</td>
<td>81479</td>
</tr>
</tbody>
</table>

What is alpha-1 antitrypsin deficiency

Definition

Alpha-1 antitrypsin deficiency (AATD) is a disorder that is inherited in an autosomal recessive manner. AATD results from mutations in the SERPINA1 gene, which codes for the enzyme alpha-1 antitrypsin (AAT).¹

Prevalence

It is estimated that 1 in 5000 to 1 in 7000 people in North America have AATD. AATD commonly afflicts individuals of Northern European heritage. This disorder is most common in Scandinavia, occurring in approximately 1 in 1500 to 1 in 3000 individuals there.¹ However, AATD is an under-recognized condition, with estimates that only 10% of those affected are actually diagnosed.²

Symptoms

The most common clinical manifestation is chronic obstructive pulmonary disease (COPD), particularly emphysema.¹⁻³ Smoking is a major environmental risk factor for lung disease in AATD, increasing the risk for emphysema by 1000-fold.³

AATD also increases the risk for neonatal or childhood liver disease, manifested by obstructive jaundice and hyperbilirubinemia, and early onset adult liver disease, usually cirrhosis and fibrosis.¹
Inheritance

Alpha-1 antitrypsin deficiency (AATD) is inherited in an autosomal recessive manner.\(^1\)

Diagnosis

AATD may first be suspected based on reduced serum levels of alpha-1 antitrypsin. Confirmatory testing includes either protease inhibitor typing or genetic testing for common mutations.\(^1\)

Test information

Introduction

Testing for alpha-1 antitrypsin deficiency may include protease inhibitor typing, SERPINA1 targeted mutation analysis, or SERPINA1 sequencing.

Protease Inhibitor typing

Protease Inhibitor (PI) typing by isoelectric focusing to determine phenotype (PI*Z, PI*S).\(^1\) PI typing is considered the gold standard for diagnosing AATD, as it can detect normal as well as variant alleles, but cannot detect null alleles\(^1,2\) If PI typing is ambiguous, mutation testing should be performed.\(^1\)

SERPINA1 targeted mutation analysis

SERPINA1 targeted mutation analysis tests for the two common mutations in the gene (Z and S), which make up greater than 95% of the mutations.\(^1\) The Z allele is by far the most common and more severe variant.\(^3\)

SERPINA1 sequencing

SERPINA1 sequencing is available, but only appropriate in limited situations. Sequencing will identify 99% of mutations associated with AATD.\(^1\)

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to alpha-1 antitrypsin deficiency testing.
American Thoracic Society and the European Respiratory Society recommendations

The American Thoracic Society and the European Respiratory Society states that testing for AATD is recommended for the following indications (quoted directly):³

• symptomatic adults with emphysema, chronic obstructive pulmonary disease (COPD), or asthma with airflow obstruction that is incompletely reversible after aggressive treatment with bronchodilators

• individuals with unexplained liver disease, including neonates, children, and adults, particularly the elderly

• asymptomatic individuals with persistent obstruction on pulmonary function tests with identifiable risk factors, examples include cigarette smoking and occupational exposure

• adults with necrotizing panniculitis, and

• siblings of an individual with AATD.

Other recommendations

The following sections outline recommendations from other authorities. However, these sources do not specifically comment on the use of SERPINA1 sequencing in the diagnostic work-up. When ambiguous results are obtained between quantification, genotype or phenotype assays, gene sequencing can identify rare variants or null alleles that would otherwise be missed.

Prins et al. (2008)

Prins et al. (2008) sequenced exons 2, 3, and 5 of the SERPINA1 gene from 66 patients with AAT concentration less than or equal to 1.0 g/L. They predicted that up to 22% of the disease-associated AAT deficiency alleles could be missed by S and Z genotyping or by phenotyping. They also identified rare alleles Mprocida, Mpalermo, M6passau, Mwurzburg, Mheerlen and the previously undescribed null alleles Q0soest and Q0amersfoort.

They found pathogenic variants in 22% of those who had negative PI and targeted mutation testing. The authors recommend direct sequencing of the coding regions of the SERPINA1 gene for patients with suspected AATD based on a serum AAT concentration ≤1.0 g/L.⁴

Ferrarotti et al. (2007)

Ferrarotti et al. (2007) described a protocol they developed to optimize AAT deficiency diagnosis from dried blood spot samples. The protocol has an initial screen using quantification of AAT and genotyping for the S and Z deficiency alleles. Discordant samples are then reflexed to PI typing.
Sequencing is used for any samples in which the plasma AAT level is low (<70 mg/dL), and the genotype/phenotype results are PI*MS or PI*MZ. Specific testing for the Q0_{isola di procida} allele is also performed, which results from a deletion and therefore cannot be detected by sequencing. While this report described the protocol used, it did not comment on the sensitivity or specificity of this approach.\(^5\)

**Graham, et al. (2015)**

Graham, et al. (2015) found pathogenic variants with sequencing after PI and targeted mutation analysis were performed. They support full gene sequencing when there is discrepancies between clinical presentation and genotyping after PI and targeted mutation analysis.\(^6\)

**Criteria**

**Introduction**

Requests for alpha-1 antitrypsin deficiency (AATD) testing are reviewed using these criteria.

**Criteria**

Protease inhibitor (PI) typing or SERPINA1 common mutation analysis (S, Z) may be considered in individuals who meet the following criteria:\(^1,3\)

- Abnormally low (less than 120mg/dL) or borderline (90-140mg/dL) alpha-1 antitrypsin (AAT) levels; AND
- At least one of the following:
  - Symptomatic adults with emphysema, chronic obstructive pulmonary disease (COPD), or asthma with airflow obstruction that is incompletely reversible after aggressive treatment with bronchodilators; or
  - Individuals of any age with unexplained liver disease (including obstructive liver disease in infancy); or
  - Asymptomatic individuals with persistent obstruction on pulmonary function tests who have identifiable risk factors (e.g., cigarette smoking, occupational exposure); or
  - C-ANCA positive vasculitis; or
  - Adults with necrotizing panniculitis; or
  - Siblings of an individual with AATD

Sequencing of the SERPINA1 gene may be considered in individuals who meet the following criteria:\(^1\)
• There are discrepancies between clinical presentation, serum alpha-1 antitrypsin quantification, targeted mutation analysis, and/or PI typing; OR
• The presence of rare variants or null alleles (which cannot be identified by other methods) is suspected.

References

Introduction

These references are cited in this guideline.


Amyotrophic Lateral Sclerosis (ALS) Genetic Testing

Introduction

ALS genetic testing is addressed by this guideline.

Procedures addressed

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<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>ALS Known Familial Mutation Analysis</td>
<td>81403</td>
</tr>
<tr>
<td>Genetic Testing for ALS</td>
<td>S3800</td>
</tr>
<tr>
<td>ALS Gene Analysis</td>
<td>81400-81408</td>
</tr>
<tr>
<td>ALS Gene Analysis</td>
<td>81479</td>
</tr>
</tbody>
</table>

What is amyotrophic lateral sclerosis

Definition

Amyotrophic lateral sclerosis (ALS) is a disease caused by the progressive degradation of motor neurons (nerve cells that control muscle movement).¹ ALS may initially present with muscle weakness, twitching, cramping, or slurred speech.¹ Symptoms worsen over time and include muscle atrophy and difficulty swallowing.¹

Diagnosis

Most cases of suspected ALS are diagnosed based on a unique combination of symptoms and the exclusion of similar disorders. The Escorial Criteria were developed in 2000 to standardize the clinical diagnosis of ALS.² These criteria include:

• the presence of upper and lower motor neuron deterioration
• the progressive spread of symptoms, and
• no clinical evidence of other diseases with similar symptoms.

Causes of ALS

There are more than 20 genes known to cause familial Amyotrophic Lateral Sclerosis (FALS). Genetic testing for many of the genes is clinically available.\(^1,3,4\) FALS subtypes are named based on the causative gene. For example, ALS1 subtype is caused by SOD1 gene mutations.

A pathogenic mutation can be identified in 60-70% of cases of FALS. The majority of combined ALS/FTD cases with a family history of either disorder are caused by C9orf72 repeat expansions.\(^4\)

Genes associated with familial ALS

Genetic testing for many of the genes is available clinically.\(^1,3,4\)

<table>
<thead>
<tr>
<th>Gene symbol</th>
<th>FALS subtype</th>
<th>% of individuals with FALS</th>
<th>Inheritance</th>
</tr>
</thead>
<tbody>
<tr>
<td>SOD1</td>
<td>ALS1</td>
<td>20%</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>C9orf72</td>
<td>ALS/FTD</td>
<td>23%-30%</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>FUS/TLS</td>
<td>ALS6</td>
<td>~4%</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>TARDBP</td>
<td>ALS10</td>
<td>1%-4%</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>18q21</td>
<td>ALS3</td>
<td>Rare</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>SETX</td>
<td>ALS4 (Motor neuropathy with pyramidal features)</td>
<td>Rare</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>20p13</td>
<td>ALS7</td>
<td>Rare</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>VAPB</td>
<td>ALS8 (Finkel type SMA or SMA IV)</td>
<td>Rare</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>FIG4</td>
<td>ALS11</td>
<td>Rare</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>17q</td>
<td>ALS/FTD</td>
<td>Rare</td>
<td>Autosomal dominant</td>
</tr>
<tr>
<td>ANG</td>
<td>ALS9</td>
<td>Rare</td>
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<tr>
<td>VCP</td>
<td>ALS14</td>
<td>1-2%</td>
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<td>HNRNPA1</td>
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<tr>
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<td>ALS17</td>
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<td>Unknown</td>
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### Gene symbol | FALS subtype | % of individuals with FALS | Inheritance
--- | --- | --- | ---
TUBA4A | ALS22 | Rare | Autosomal dominant
SQSTM1 | ALS/FTD | ~1% | Autosomal dominant
TBK1 | ALS/FTD | Rare | Autosomal dominant
ALS2 | ALS2 | Rare | Autosomal dominant
ALS7 (20p13) | FALS | Rare | Autosomal dominant
CHCHD10 | ALS/FTD | Rare | Autosomal dominant
SPG11 | ALS5 | Rare | Autosomal recessive
OPTN | ALS12 | Rare | Autosomal recessive
SPG20 | SPG20 | Rare | Autosomal recessive
UBQLN2 | ALS15 | Rare | X-linked dominant

### Inheritance

Most people with FALS have an autosomal dominant form, meaning only one mutation is required to cause disease. In this case, children of an affected person have a 50% chance of inheriting the disease-causing mutation.

There are rare autosomal recessive forms of ALS as well as one X-linked form. Two mutations are required to cause autosomal recessive types, usually only siblings are affected, and there is no parent-to-child transmission.

### Onset

The average age of ALS onset is 56 years if the affected individual has no family history, and 46 years old if there is a family history of ALS. However, there are infantile and juvenile onset forms that should also prompt consideration of a genetic etiology.

### Survival

ALS is fatal. The average survival after diagnosis is 3 years, but can vary widely. Treatment focuses on slowing progression with medication and therapy.

### Prevalence

Between 4 and 8 out every 100,000 people develop ALS. About 90% of ALS cases are sporadic, and the remaining 10% of individuals have familial ALS (FALS).
Test information

Introduction

Testing for Familial Amyotrophic Lateral Sclerosis (FALS) may include gene sequencing or known familial mutation analysis.

Sequence analysis

Genetic testing for FALS is usually done by gene sequencing because mutations are diverse. Sequencing is generally >99% accurate for identifying mutations in the coding region of a gene.

Laboratories may offer individual gene sequencing or multi-gene panels for FALS.

Sequential genetic testing approach

An expert-authored review makes the following suggestions when pursuing a sequential individual gene approach:¹

- “SOD1 testing is appropriate in any individual with ALS who has another affected family member or an incomplete family history, including the early death of a close relative from any cause. Approximately 20% of individuals with FALS have ALS1 with an identified disease-causing mutation in SOD1. Interpretation of the significance of a SOD1 mutation regarding disease severity and progression depends on the specific mutation identified because of wide variability in genotype/phenotype correlations. Failure to detect a SOD1 mutation does not rule out FALS. Up to 3% of individuals with ALS with no family history of ALS have SOD1 mutations. Because data on penetrance of many mutations are limited, establishing the risk to other family members of developing clinical symptoms can be difficult.”

- “SETX testing is appropriate in kindreds with adolescent-onset spinal muscular atrophy with pyramidal features.”

- “VAPB testing should be pursued in the context of clinical symptoms of primarily adult-onset spinal muscular atrophy.”

- “FUS/TLS, TARDBP, and ANG testing should be considered for SOD1-negative individuals with FALS.”

- “ALS2 testing is appropriate for those with childhood-onset UMN-predominant ALS.”

- “VCP testing should be considered for individuals with a family history of ALS with or without symptoms of inclusion body myopathy, Paget disease and/or frontotemporal dementia.”
“OPTN testing may be considered for individuals with a family history consistent with autosomal dominant or autosomal recessive inheritance, including simplex cases who do not have a mutation in more common ALS-related genes.”

**Known familial mutation analysis**

Known familial mutation analysis can provide predictive information about the risk to develop ALS. It can also be used to diagnose ALS when the patient does not yet meet the full ALS diagnostic criteria.\(^5\)

Once a mutation has been identified through sequencing in an affected family member, it is straightforward to test at-risk relatives for that one mutation. The involved gene and precise mutation name/location must be known.

The detection rate for a known familial mutation is greater than 99%.

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to ALS genetic testing.

**World Federation of Neurology Research Group on Motor Neuron Diseases**


- These revised criteria still do not specify when genetic testing should be done, but they do state “If a pathogenic mutation in a disease-causing gene is found in the patient and segregates with the disease the term hereditary or primary genetic ALS (HALS/GALS) should be used. The finding of a pathogenic mutation in a known gene can substitute for either lower or upper motor neuron signs, so that diagnosis of ALS can be made on the basis of UMN or LMN signs in one body region, associated with a positive genetic test.”
- “ALS can be defined as Mendelian in inheritance if a disease-causing gene variant can be shown to segregate within a family. In such cases the genetic variant can serve as a substitute for upper motor neuron deficits or a second limb or region (rule of two).”

**Expert-authored review**

A 2015 expert-authored review states: “Presymptomatic testing for a TARDBP mutation is complicated because the penetrance is unknown, the age of onset is not predictable, and preventative measures do not exist. Because of the individualized nature of predictive testing, consultation with a genetic counselor and a psychologist to obtain informed consent is recommended. At this time, no established testing protocol (e.g., as
in Huntington disease) exists, although establishment of such protocols has been suggested. However, to err on the side of caution, testing centers often follow a similar protocol.”

Identifying a SOD1 mutation in a pre-symptomatic individual can impact future management and overall prognosis of ALS. However, it is considered controversial because of reduced penetrance, which means that not everyone with a mutation will necessarily develop symptoms. It also lacks overall intervention or prevention strategies and has an inability to predict the age of onset.1,3

**European Federation of Neurological Societies**

A European Federation of Neurological Societies Task Force (EFNS, 2012) addressed presymptomatic testing in its diagnosis and management guidelines: “Presymptomatic genetic testing should only be performed in first-degree adult blood relatives of patients with a known gene mutation. Testing should only be performed on a strictly voluntary basis as outlined (see Table 7 in the original guideline document) and should follow accepted ethical principles.”

**European Federation of Neurological Societies**

Guidelines from the European Federation of Neurological Societies (EFNS, 2012) address molecular testing of ALS:8

“Clinical deoxyribonucleic acid (DNA) analysis for gene mutations should only be performed in cases with a known family history of ALS, and in sporadic ALS cases with the characteristic phenotype of the recessive D90A mutation.”

“Clinical DNA analysis for gene mutations should not be performed in cases with sporadic ALS with a typical classical ALS phenotype.”

“In familial or sporadic cases where the diagnosis is uncertain, SMN, androgen receptor, or TARDBP, FUS, ANG, or SOD1 DNA analysis may accelerate the diagnostic process.”

“Before blood is drawn for DNA analysis, the patient should receive genetic counselling. Give the patient time for consideration. DNA analysis should be performed only with the patient’s informed consent.”

**European Federation of Neurological Societies**

Guidelines from the European Federation of Neurological Societies (EFNS, 2011) address the molecular diagnosis of ALS and other neurogenetic disorders. They state:9

“Currently, molecular diagnosis mainly has implications for genetic counseling rather than for therapy. However, when more directed causal therapies become available in the future, establishing a correct genetic diagnosis in a given patient will be essential. Despite the rather low prevalence sequencing of the small SOD1 gene should be considered in patients with ALS with dominant inheritance to offer presymptomatic or prenatal diagnosis, if this is requested by the family (Level B).”
World Federation of Neurology Research Group on Motor Neuron Diseases


This group doesn't specify when genetic testing should be done, but they do state “The demonstration of the presence of a pathogenetically relevant gene mutation can assist in the diagnosis of ALS (such as SOD1).”

These criteria set a lower threshold for diagnosis when an ALS-causing mutation is known in the family. For example, a patient may be diagnosed as “Clinically Definite Familial ALS — Laboratory-supported” with evidence of only upper or lower motor neuron disease in one region; whereas a definite diagnosis without genetic test results requires upper and lower motor neuron disease in three regions.

Criteria

Introduction

Requests for ALS genetic testing are reviewed using these criteria.

Known familial mutation testing

• Genetic Counseling
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous full gene sequencing and/or large rearrangement testing of the gene with the known familial mutation, AND
  o Known familial mutation in a gene that causes amyotrophic lateral sclerosis (e.g., SOD1, C9orf72, FUS, TARDBP) identified in a 1st, 2nd, or 3rd degree relative(s), AND
  o Age 18 years or older, AND
  o Rendering laboratory is a qualified provider of service per the Health Plan policy.

Full sequence and deletion/duplication analysis

• Full gene sequencing and deletion/duplication analysis for ALS are considered investigational and experimental and, therefore, not eligible for reimbursement.
References

Introduction

These references are cited in this guideline.


Angelman Syndrome Testing

Introduction

Angelman syndrome testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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What is Angelman syndrome

Definition

Angelman syndrome (AS) is a genetic disorder that can cause intellectual disability, severe speech impairment, tremors, seizures, microcephaly, and decreased need for sleep.

Symptoms

Angelman syndrome (AS) is characterized by:\(^1\)

- Severe developmental delay or intellectual disability by age 6-12 months
- Severe speech impairment — usually with minimal or no word use
- Gait ataxia and limb tremors
- Seizures and microcephaly
• Happy demeanor with hand flapping, and
• Decreased need for sleep.

Causes

Features of Angelman syndrome are caused by a missing or defective UBE3A gene inherited from the individual's mother.²

A missing or defective UBE3A gene can be caused by a gene deletion, gene mutation, uniparental disomy (two copies of paternal chromosome), imprinting defect, or a chromosome rearrangement.²³

Test information

Introduction

Testing for Angelman syndrome may include SNRPN/UBE3A methylation analysis, FISH analysis for 15q11-q13 deletion, chromosome 15 uniparental disomy (UPD), imprinting center defect analysis, UBE3A sequencing, or known familial mutation analysis.

SNRPN/UBE3A methylation analysis

This test is typically the first test in the evaluation of both Angelman syndrome (AS) and Prader-Willi syndrome (PWS). It will detect about 80% of patients with AS and >99% of patients with PWS. However, DNA methylation analysis does not identify the underlying cause, which is important for determining the risk to future siblings. This risk ranges from less than 1% to up to 50%, depending on the genetic mechanism. Follow-up testing for these causes may be appropriate.

FISH analysis for 15q11-q13 deletion

If DNA methylation analysis for Angelman (AS) or Prader-Willi syndrome (PWS) is abnormal, deletion analysis is typically the next step. Approximately 70% of cases of both AS and PWS have a deletion in one copy of chromosome 15 involving the 15q11.2-q13 region. When looking specifically for this deletion, FISH (fluorescence in situ hybridization) analysis is most commonly performed. However, chromosome microarray can also detect such deletions (see that guideline for guidance). If chromosomal microarray (CMA, array CGH) has already been done, FISH is not likely to be necessary.

Chromosome 15 uniparental disomy (UPD)

If DNA methylation analysis is abnormal but deletion analysis is normal, UPD analysis may be an appropriate next step for evaluation of both Angelman (AS) and Prader-Willi syndrome (PWS). About 28% of PWS cases are due to maternal UPD (both chromosome 15s are inherited from the mother). About 7% of cases of AS are due to
paternal UPD (both chromosome 15s are inherited from the father). Both parents must be tested to diagnose UPD.

**Imprinting center defect analysis**

This test may be considered in the evaluation of Angelman syndrome (AS) and Prader-Willi syndrome (PWS) when methylation is abnormal, but FISH (or array CGH) and UPD studies are normal. Individuals with such results are presumed to have an imprinting defect. An abnormality in the imprinting process has been described in a minority of cases. However, imprinting center deletions may be familial, and if familial, the recurrence risk can be up to 50%.

**UBE3A sequencing**

If DNA methylation analysis is normal, UBE3A gene mutations should be suspected. Such mutations are found in 11% of Angelman syndrome patients and can only be detected by sequencing the entire gene.¹ These mutations can be carried by the mother of an affected individual and pose up to a 50% risk of recurrence in her other children, and an increased risk to other family members.

**Known familial mutation analysis**

If a UBE3A gene mutation has been identified in an affected individual through sequencing, testing for just the known familial mutation in UBE3A can be performed for at-risk relatives, including at-risk pregnancies.

If a mutation in the imprinting center has been identified in an affected family member, testing for just the known familial mutation in the imprinting center can be performed for at-risk relatives, including at-risk pregnancies.

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to Angelman syndrome testing.

**American College of Medical Genetics and American Society of Human Genetics**

Consensus guidelines from the American College of Medical Genetics and American Society of Human Genetics (2006) recommend two equally-accepted tiered approaches to testing.³

- **Approach one**
  - Start with UBE3A methylation analysis.
    - If abnormal (only paternal alleles are present), a diagnosis is confirmed.
Consider the following to identify the underlying cause for recurrence risk counseling.

- **FISH 15q11-q13 (deletion analysis)**
  - If FISH testing is abnormal, FISH testing on the mother should be done to rule out an inherited chromosome abnormality (rare)
  - If FISH testing is normal, consider UPD analysis.

- **Uniparental Disomy (UPD) analysis of chromosome 15** to determine whether the proband inherited both copies of chromosome 15 from the father.
- If FISH and UPD analysis are normal, an imprinting center mutation is a likely cause and should be evaluated (which may carry a higher recurrence risk than other causes).

**Approach two**

- Start with FISH 15q11-q13 (deletion analysis)
  - If abnormal, a diagnosis is confirmed
  - If normal, then proceed to methylation analysis.

- **UBE3A Methylation analysis**
  - If methylation analysis is abnormal, the diagnosis is confirmed, but UPD testing may be done to better estimate recurrence risk

- **Uniparental Disomy (UPD) analysis of chromosome 15**

- If methylation analysis is abnormal, but FISH and UPD analysis are normal, an imprinting center mutation is a likely cause and should be evaluated (which may carry a higher recurrence risk than other causes).

**Expert-authored review**

An expert-authored review (2011) comments on the utility of familial mutation analysis:¹

- “Individuals with an imprinting center (IC) deletion can have a phenotypically normal mother who also has an IC deletion. If a proband's mother has a known IC deletion, the risk to the sibs is 50%.”
- “UBE3A mutations can be inherited or de novo. In addition, several cases of mosaicism for a UBE3A mutation have been noted. If a proband's mother has a UBE3A mutation, the risk to the sibs is 50%.”

• “If a proband’s mother carries a known IC deletion or UBE3A mutation, the mother's sisters are also at risk of carrying the IC deletion or the mutation. Each child of the unaffected sisters who are carriers is at a 50% risk of having AS. Unaffected maternal uncles of the proband who are carriers are not at risk of having affected children, but are at risk of having affected grandchildren through their unaffected daughters who have inherited the IC deletion or UBE3A mutation from them.”

Criteria

Introduction

Requests for Angelman syndrome testing are reviewed using these criteria.

SNRPN/UBE3A Methylation Analysis

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Testing:
  o No previous SNRPN/UBE3A methylation analysis, AND
• Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay by age 6-12 months, typically severe to profound, without loss of milestones, and
  o Some combination of the following:
    ▪ Movement or balance disorder, typically with ataxia, or
    ▪ Frequent laughter/smiling, apparent happy demeanor; easily excitable personality (often with uplifted hand-flapping, or waving movements), or hypermotric behavior, or
    ▪ Speech impairment with no or minimal number of words, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

FISH Analysis for 15q11-q13 Deletion

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Testing:
  o No previous chromosomal microarray, and
• Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay by age 6-12 months, typically severe to profound, without loss of milestones, and
  o Some combination of the following:
    ▪ Movement or balance disorder, typically with ataxia, or
    ▪ Frequent laughter/smiling, apparent happy demeanor; easily excitable personality (often with uplifted hand-flapping, or waving movements), or hypermotoric behavior, or
    ▪ Speech impairment with no or minimal number of words, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Chromosome 15 Uniparental Disomy

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o SNRPN/UBE3A methylation analysis results are abnormal, and
  o 15q11-q13 deletion analysis is negative, and
  o No previous chromosome 15 UPD studies, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay by age 6-12 months, typically severe to profound, without loss of milestones, and
  o Some combination of the following:
    ▪ Movement or balance disorder, typically with ataxia, or
    ▪ Frequent laughter/smiling, apparent happy demeanor; easily excitable personality (often with uplifted hand-flapping, or waving movements), or hypermotoric behavior, or
    ▪ Speech impairment with no or minimal number of words, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Imprinting Center Defect Analysis

• Genetic Counseling:
o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  o SNRPN/UBE3A methylation analysis results are abnormal, and
  o 15q11-q13 deletion analysis is negative, and
  o Previous chromosome 15 UPD testing is negative, and
  o No previous imprinting center (IC) analysis, AND

- Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay by age 6-12 months, typically severe to profound, without loss of milestones, and
  o Some combination of the following:
    - Movement or balance disorder, typically with ataxia, or
    - Frequent laughter/smiling, apparent happy demeanor; easily excitable personality (often with uplifted hand-flapping, or waving movements), or hypermotoric behavior, or
    - Speech impairment with no or minimal number of words, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**UBE3A Sequencing**

- Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  o SNRPN/UBE3A methylation analysis results are normal, and
  o No previous sequencing of UBE3A, AND

- Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay by age 6-12 months, typically severe to profound, without loss of milestones, and
  o Movement or balance disorder, typically with ataxia, and
  o Frequent laughter/smiling, apparent happy demeanor; easily excitable personality (often with uplifted hand-flapping, or waving movements), or hypermotoric behavior, and
• Speech impairment with no or minimal number of words, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**UBE3A Deletion/Duplication Analysis**

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o SNRPN/UBE3A methylation analysis results are normal, and
  o Normal UBE3A sequencing, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay by age 6-12 months, typically severe to profound, without loss of milestones, and
  o Movement or balance disorder, typically with ataxia, and
  o Frequent laughter/smiling, apparent happy demeanor; easily excitable personality (often with uplifted hand-flapping, or waving movements), or hypermotoric behavior, and
  o Speech impairment with no or minimal number of words, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Known Familial Mutation Analysis**

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous UBE3A sequencing or imprinting center defect analysis testing, AND

• Family History:
  o Known familial UBE3A mutation in a blood relative, or
- Known familial imprinting center defect mutation in a blood relative, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

References

Introduction

These references are cited in this guideline.


APOE Variant Analysis for Alzheimer Disease Testing

Introduction

APOE variant analysis for Alzheimer disease testing is addressed by this guideline.

Procedures addressed

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What is Alzheimer disease

Definition

Alzheimer disease (AD) is characterized by an adult-onset, progressive dementia with cerebral cortical atrophy and beta amyloid plaque formation.¹

Prevalence

The general population lifetime risk of AD is about 10%. First-degree relatives, siblings or offspring of a single person in the family with AD have a 20-25% lifetime risk.²

Familial AD

Of all people with AD, about 25% have at least two affected people in the family. This is referred to as familial AD.²

Approximately 95% of people with familial AD develop symptoms after 65.² This is called late-onset familial AD. Late-onset familial AD is believed to have complex inheritance with multiple susceptibility genes and environmental factors playing a role.²

In about 5% of familial cases, symptoms consistently start before 65.² This is called early onset familial Alzheimer disease (EOFAD). EOFAD is an autosomal dominant inherited disorder caused by different genes than those that may predispose to late-onset AD.¹

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Symptoms
Common findings include memory loss, confusion, speech issues, hallucinations, and personality and behavioral changes such as poor judgment, agitation, and withdrawal.\textsuperscript{2,3}

Onset
Symptoms of AD usually start after 60-65 years old.\textsuperscript{2}

APOE variants
There are three common versions of the APOE gene: e2, e3, and e4.

\textbf{e4 variant}

The e4 variant is significantly associated with Alzheimer disease.\textsuperscript{2}

People with AD, and especially late-onset familial AD, are more likely to have one or two copies of APOE e4. For example, less than 1\% of unaffected people have two copies of e4 (e4/e4), but nearly 19\% of people with familial AD have two copies of e4.\textsuperscript{2}

APOE e4 is not necessary to develop AD and having no copies of e4 does not rule out the disease.\textsuperscript{2,4} APOE e4 appears to cause susceptibility to AD, but the reason is unclear.\textsuperscript{2,5}

Test information
Introduction
Testing for APOE gene variants is available clinically.

APOE variant clinical testing
Many laboratories in the U.S. directly test for these three variants (e2, e3, e4) to assist diagnosis or predict risk of Alzheimer disease.

Guidelines and evidence
Introduction
This section includes relevant guidelines and evidence pertaining to APOE variant analysis for AD.
The American College of Medical Genetics and The National Society of Genetic Counselors (2011)

“Genetic testing for susceptibility loci (e.g., APOE) is not clinically recommended due to limited clinical utility and poor predictive value.”

“Because the ε4 allele is neither necessary nor sufficient to cause AD, there have been numerous consensus statements and articles that have recommended against using APOE genotyping for predicting AD risk.”

European Federation of Neurological Societies (2010)

“The ApoE e4 allele is the only genetic factor consistently implicated in late-onset AD, but it is neither necessary nor sufficient for development of the disease. Hence, there is no evidence to suggest ApoE testing is useful in a diagnostic setting.”


“Insofar as patients with AD are more likely to have an APOE-e4 allele than are patients with other forms of dementia or individuals without dementia, physicians may choose to use APOE genotyping as an adjunct to other diagnostic tests for AD.”

“Since genotyping cannot provide certainty about the presence or absence of AD, it should not be used as the sole diagnostic test.”

“The use of APOE genotyping to predict future risk of AD in symptom-free individuals is not recommended at this time.”

Criteria

Introduction

Requests for APOE variant analysis for AD are reviewed using these criteria.

Criteria

• This test is considered investigational and/or experimental
  o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
• In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility

References

Introduction

These references are cited in this guideline.


## Introduction

Ashkenazi Jewish carrier screening is addressed by this guideline.

## Procedures addressed

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<td>Molecular pathology procedure, Level 3</td>
<td>81402</td>
</tr>
</tbody>
</table>
What is Ashkenazi Jewish carrier screening

Definition

Ashkenazi Jewish carrier screening is available for certain genetic conditions that are more common or have superior mutation detection rates in the Ashkenazi Jewish population. “Ashkenazi” refers to someone whose Jewish ancestors originally came from Central or Eastern Europe, such as Russia, Poland, Germany, Hungary, Lithuania. Most Jewish people in the US are of Ashkenazi descent. There are regional differences in the number and types of tests commonly offered. Individuals and providers may choose all or a subset of these conditions.

Inheritance

These Jewish genetic diseases are inherited in an autosomal recessive manner. An affected individual must inherit a gene mutation from both parents.

- Individuals who inherit only one mutation are called carriers. Carriers do not show symptoms of the disease, but have a 50% chance of passing on the mutation to their children.
- Two carriers of the same disease have a 25% chance of having a child with the disorder.

Prevalence

While these genetic diseases are individually rare, the overall chance for an individual of Ashkenazi Jewish descent to be a carrier for one of these genetic diseases is 1 in 4 to 1 in 5. An individual can also be a carrier of more than one condition.
People from other ethnic backgrounds can be carriers of these conditions, but it is generally less common. The test is typically not as effective at identifying carrier status in individuals of non-Ashkenazi Jewish descent.

**Test information**

**Introduction**

Ashkenazi Jewish carrier screening can be offered to couples or individuals of Ashkenazi Jewish descent when they are planning a pregnancy (preconceptional) or during a pregnancy (prenatal).¹-³

**One member of couple is Jewish**

If only one member of the couple is Ashkenazi Jewish, carrier screening should start with the Ashkenazi Jewish partner. Both parents must be carriers to have an affected child, so reproductive partners of known carriers should also be offered testing even if not Jewish. In some cases, full gene sequencing would be most appropriate for testing of a non-Jewish partner.

**Purpose of test**

Carrier screening generally looks for a small number of gene mutations that are particularly common in the Ashkenazi Jewish population, although an increasing number of full gene sequencing panels are becoming available.

In addition, enzyme analysis is particularly effective for Tay-Sachs disease and is generally preferred to mutation testing.

**Detection rate**

The carrier detection rate is greater than 95% in the Ashkenazi Jewish population for most diseases.³

The detection rate for these tests in the non-Ashkenazi population is unknown for most conditions, but generally low. Exceptions include cystic fibrosis and Tay-Sachs enzyme analysis, which each have good detection rates in non-Jewish populations.

A negative test result in one or both partners significantly lowers the chance of an affected child, but does not eliminate it.²

**Commonly tested conditions**

The genes included in carrier screening panels vary widely between laboratories. The following table includes the most commonly tested conditions.
<table>
<thead>
<tr>
<th>Ashkenazi Jewish genetic disease</th>
<th>Ashkenazi carrier frequency</th>
<th>What the test looks for</th>
<th>Chance of correctly finding an Ashkenazi Jewish carrier</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bloom syndrome</td>
<td>1/107</td>
<td>1 mutation (2281del6ins7)</td>
<td>Greater than 99%</td>
</tr>
<tr>
<td>Canavan disease</td>
<td>1/41</td>
<td>2 mutations (E285A, Y231X)</td>
<td>97.4%</td>
</tr>
<tr>
<td>Cystic fibrosis</td>
<td>1/29</td>
<td>23 most common mutations in several ethnic groups</td>
<td>97%</td>
</tr>
<tr>
<td>Dihydrolipoamide dehydrogenase deficiency</td>
<td>1/107</td>
<td>2 mutations (G229C and Y35X)</td>
<td>Greater than 95%</td>
</tr>
<tr>
<td>Familial dysautonomia</td>
<td>1/31</td>
<td>2 mutations (2507+6TtoC, R696P)</td>
<td>Greater than 99%</td>
</tr>
<tr>
<td>Familial hyperinsulinism</td>
<td>1/68</td>
<td>2 mutations (c.3989-9G&gt;A and Phel1387del)</td>
<td>90%</td>
</tr>
<tr>
<td>Fanconi anemia group C</td>
<td>1/89</td>
<td>1 mutation (IVS4+4AtoT)</td>
<td>Greater than 99%</td>
</tr>
<tr>
<td>Gaucher disease</td>
<td>1/18</td>
<td>4 mutations (N370S, 84GG, L444P, IVS2+1GtoA)</td>
<td>Up to 94.6%</td>
</tr>
<tr>
<td>Glycogen storage disease type 1A (GSD1A)</td>
<td>1/71</td>
<td>1 mutation (R83C)</td>
<td>93% to 100%</td>
</tr>
<tr>
<td>Joubert syndrome</td>
<td>1/92</td>
<td>1 mutation (R12L)</td>
<td>99%</td>
</tr>
<tr>
<td>Maple syrup urine disease (MSUD)</td>
<td>1/80</td>
<td>3 mutations (R183P, G278S, E372X)</td>
<td>About 99%</td>
</tr>
<tr>
<td>Mucolipidosis IV</td>
<td>1/127</td>
<td>2 mutations (IVS3–2AtoG, Del6.4kb)</td>
<td>95%</td>
</tr>
<tr>
<td>Nemaline myopathy</td>
<td>1/168</td>
<td>1 mutation (R2478_D2512del)</td>
<td>Greater than 95%</td>
</tr>
<tr>
<td>Niemann-Pick disease type A</td>
<td>1/90</td>
<td>3 mutations (R496L, L302P, fsP330)</td>
<td>97%</td>
</tr>
<tr>
<td>Tay-Sachs disease</td>
<td>1/90</td>
<td>Mutation analysis: 3 mutations (1278insTATC,</td>
<td>92-94%</td>
</tr>
</tbody>
</table>
### Guidelines and evidence

#### Introduction

This section includes relevant guidelines and evidence pertaining to Ashkenazi Jewish carrier screening.

**American College of Obstetrics and Gynecology and American College of Medical Genetics and Genomics**

The American College of Obstetrics and Gynecology (ACOG, 2009) and the American College of Medical Genetics and Genomics (ACMG, 2008) recommend carrier screening for a group of disorders when at least one member of a couple is Ashkenazi Jewish and that couple is pregnant or planning pregnancy.

Both ACOG and ACMG agree that testing should be offered for cystic fibrosis, Canavan disease, familial dysautonomia, and Tay-Sachs.

ACMG also recommends routine testing for Fanconi anemia, Niemann-Pick, Bloom syndrome, mucolipidosis IV, and Gaucher disease; while ACOG states “individuals of Ashkenazi Jewish descent may inquire about the availability of carrier screening for other disorders” and educational materials may be provided to assist informed decision making about additional tests.

**ACMG guidance for additional carrier screening**

Carrier screening for common Ashkenazi Jewish mutations that cause many other conditions is now clinically available, but these tests are not specifically addressed in current carrier screening guidelines. However, the 2008 ACMG guidelines outline the criteria for recommending additional carrier screening in the Ashkenazi Jewish population as new tests become available:

- the natural history must be well understood
- people affected with the disorder must have significant morbidity and mortality, and

<table>
<thead>
<tr>
<th>Ashkenazi Jewish genetic disease</th>
<th>Ashkenazi carrier frequency</th>
<th>What the test looks for</th>
<th>Chance of correctly finding an Ashkenazi Jewish carrier</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>1421+1GtoC, G269S OR</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Hexosaminidase A enzyme analysis</td>
<td>About 98%</td>
</tr>
<tr>
<td>Usher syndrome III4</td>
<td>1/120</td>
<td>1 mutation (N48K)</td>
<td>Greater than 95%</td>
</tr>
</tbody>
</table>
• the test must have greater than 90% detection OR the allele frequency must be at least 1%.

Conditions that meet these criteria
The following conditions meet these criteria:
  o dilipoamide dehydrogenase deficiency
  o familial hyperinsulinism
  o glycogen storage disease type 1a
  o Joubert syndrome
  o maple syrup urine disease
  o nemaline myopathy, and
  o Usher syndrome type III.

Criteria
Introduction
Requests for Ashkenazi Jewish Carrier Screening are reviewed using these criteria.

Ashkenazi Jewish Genetic Diseases Carrier Screening Panels

• Testing may be considered for carrier screening for all or any desired subset of the Ashkenazi Jewish genetic diseases eligible for coverage per the Coverage Guidance table when the following criteria are met:
  o The individual is planning a pregnancy or currently pregnant; and
  o At least one partner of a couple is Ashkenazi Jewish (NOTE: Detection rates for testing are higher in people with Ashkenazi Jewish ancestry. If only one partner of a couple is Ashkenazi Jewish, testing should start in that person when possible.); AND

• Testing will be billed using the procedure code 81412 that will represent all tests performed for the assessment of carrier status based on Ashkenazi Jewish ancestry and no additional tests for this purpose will be separately billed by the same lab for the same test date; or
• Testing for separate genes will be billed because the panel code is not more appropriate (e.g., fewer than the 9 stated genes will be assessed or a different methodology is used), in which case individual gene test coverage will be assessed based on the guidance provided in the Coverage Guidance table.

**Single Ashkenazi Jewish Genetic Diseases Carrier Screening Tests**

Testing may be considered for carrier screening of a single Ashkenazi Jewish disease, regardless of ethnicity or reproductive plans, if either of the following are met:

• The individual has a family history of one of these conditions; OR
• The individual’s partner is a known carrier or affected with any of these conditions

**Other considerations**

If an Ashkenazi Jewish carrier screening panel was previously performed and an updated, larger panel is being requested, only testing for the medically necessary, previously untested genes will be reimbursable. Therefore, only the most appropriate procedure codes for those additional genes will be considered for reimbursement.

**Coverage Guidance for Genes Included in Ashkenazi Jewish Carrier Screening Tests**

<table>
<thead>
<tr>
<th>Condition</th>
<th>Gene</th>
<th>CPT</th>
<th>Required claim code</th>
<th>Coverage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bloom syndrome</td>
<td>BLM</td>
<td>81209</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Canavan disease</td>
<td>ASPA</td>
<td>81200</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Cystic fibrosis</td>
<td>CFTR</td>
<td>81220</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Dihydrolipoamide dehydrogenase deficiency</td>
<td>DLD</td>
<td>81479</td>
<td>DLD</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Familial dysautonomia</td>
<td>IKBKAP</td>
<td>81260</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Familial hyperinsulinism</td>
<td>ABCC8</td>
<td>81401</td>
<td>ABCC8</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Fanconi anemia, type C</td>
<td>FANCC</td>
<td>81242</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Gaucher disease, type 1</td>
<td>GBA</td>
<td>81251</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Glycogen storage</td>
<td>G6PC</td>
<td>81250</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
</tbody>
</table>
### Ashkenazi Jewish Carrier Screening

<table>
<thead>
<tr>
<th>Condition</th>
<th>Gene</th>
<th>CPT</th>
<th>Required claim code</th>
<th>Coverage</th>
</tr>
</thead>
<tbody>
<tr>
<td>disease, type 1A</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Joubert syndrome, type 2</td>
<td>TMEM216</td>
<td>81479</td>
<td>TMEM216</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Maple syrup urine disease, type 1b</td>
<td>BCKDHB</td>
<td>81205</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Mucolipidosis, type IV</td>
<td>MCOLN1</td>
<td>81290</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Nemaline myopathy, type 2</td>
<td>NEB</td>
<td>81400</td>
<td>NEB</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Niemann-Pick disease, type A</td>
<td>SMPD1</td>
<td>81330</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Tay-Sachs disease</td>
<td>HEXA</td>
<td>81255</td>
<td>NONE</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Usher syndrome, type 1F</td>
<td>PCDH15</td>
<td>81400</td>
<td>PCDH15</td>
<td>MOL.TS.129</td>
</tr>
<tr>
<td>Usher syndrome, type 3</td>
<td>CLRN1</td>
<td>81400</td>
<td>CLRN1</td>
<td>MOL.TS.129</td>
</tr>
</tbody>
</table>

**Note** Other tests may be eligible for coverage under the above criteria if the condition is associated with significant morbidity and mortality, the allele frequency is >1% in the Ashkenazi Jewish population, and the selected test method has >90% detection rate for disease-causing mutations.

### References

**Introduction**

These references are cited in this guideline.


Ataxia-Telangiectasia Testing

Introduction

Ataxia-telangiectasia (A-T) testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>ATM known familial mutation analysis</td>
<td>81403</td>
</tr>
<tr>
<td>ATM sequencing</td>
<td>81408</td>
</tr>
<tr>
<td>ATM deletion and duplication analysis</td>
<td>81479</td>
</tr>
</tbody>
</table>

What is Ataxia-telangiectasia

Definition

Ataxia-telangiectasia (A-T) is a progressive neurological disorder that is caused by mutations in the ATM gene.

Prevalence

The prevalence of A-T is approximately 1 in 40,000 to 1 in 100,000 live US births.\textsuperscript{1,2} It is the most common cause of childhood progressive cerebellar ataxia in most countries.\textsuperscript{3}

Symptoms

Signs and symptoms of A-T include\textsuperscript{1}

- truncal and gait ataxia
- ocular apraxia
- slurred speech
- head tilting, after the age of 6 months
- conjunctival telangiectasias
- immunodeficiencies
- malignancies, especially leukemias and lymphomas, and
• radiation sensitivity.

Onset

The onset for A-T is typically between the ages of 1 and 4 years.

Related conditions

ATM has been implicated as causing an increased risk for breast cancer, especially in women with a strong family history of breast cancer.\textsuperscript{4,7} Epidemiological data has also suggested an increased risk for cardiovascular disease in carriers as well.\textsuperscript{6,7} Therefore, carriers of ATM mutant alleles may need to be screened for breast cancer and cardiovascular disease.

Inheritance

A-T is inherited in an autosomal recessive inheritance pattern. Males and females are equally likely to be affected. If both parents are carriers of A-T, the risk for a pregnancy to be affected is 1 in 4, or 25%. Preimplantation and prenatal diagnosis are available for couples known to be at-risk.

Prognosis

Although individuals with A-T live to adulthood, they are at an increased risk for early death. Currently, most individuals live beyond 25 years, with some surviving into their 50s. Cause of death is associated with A-T associated cancers, infection, and pulmonary failure.\textsuperscript{1}

Test information

Introduction

Testing for Ataxia-telangiectasia may include sequence analysis, deletion/duplication analysis, or known familial mutation analysis.

Sequence analysis

Sequence analysis of the ATM gene can identify 90-95% of A-T mutations in affected individuals.\textsuperscript{1}

Deletion and duplication analysis

Deletion and duplication analysis of the ATM gene can identify another 1-2% of mutations.\textsuperscript{1}
Known familial mutation analysis

Once a deleterious mutation has been identified, relatives of affected individuals can undergo tests. Detection of at-risk individuals affects medical management in the case of breast cancer screening and cardiovascular disease screening.

Prenatal testing is available to individuals with a known family mutation. Genetic testing can be performed on amniocytes obtained through amniocentesis or chorionic villi obtained through a chorionic villus sampling.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to Ataxia-telangiectasia testing.

International Workshop on A-T

The Eighth International Workshop on Ataxia-telangiectasia (A-T) was convened in 1999. The workshop described ATM mutations and cancer risk in heterozygotes, and potential therapeutic approaches. Genetic testing strategies were not described.

Criteria

Introduction

Requests for Ataxia-telangiectasia testing are reviewed using these criteria.

ATM known familial mutation analysis

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous genetic testing of ATM, AND
- Carrier Screening Individuals:
  - Known family mutation in ATM in 1st, 2nd, or 3rd degree biologic relative(s), OR
- Prenatal Testing for At-Risk Pregnancies:
  - ATM mutations identified in both biologic parents.
ATM sequencing

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous ATM gene sequencing, and
  o No known ATM mutation in family, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Elevated Alpha-fetoprotein (AFP) levels, or
  o Decreased ATM protein detected by immunoblotting, and
  o Progressive cerebellar ataxia, or
  o Truncal and gait ataxia, or
  o Oculomotor apraxia, OR

• Diagnostic Testing for Carriers:
  o One mutation detected by targeted mutation analysis, and
  o Elevated Alpha-fetoprotein (AFP) levels, or
  o Decreased ATM protein detected by immunoblotting, OR

• Testing for Individuals with Family History or Partners of Carriers:
  o 1st, 2nd, or 3rd, degree relative diagnosed with Ataxia-Telangiectasia clinical diagnosis, family mutation unknown, and testing unavailable, or
  o Partner is monoallelic or biallelic for ATM mutation, and
  o Has living children with this partner, or
  o Has the potential and intention to reproduce

ATM duplication and deletion analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous deletion/duplication analysis of ATM, and
- No mutations detected in full sequencing, or
- Heterozygous for mutation and elevated alpha-fetoprotein levels or decreased ATM protein detected by immunoblotting

References

Introduction

These references are cited in this guideline.


BCR-ABL Negative Myeloproliferative Neoplasm Testing

Introduction

BCR-ABL negative myeloproliferative neoplasm (MPN) testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan’s procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>JAK2 V617F Mutation Analysis</td>
<td>81270</td>
</tr>
<tr>
<td>JAK2 Exon 12 Mutation Analysis</td>
<td>81403</td>
</tr>
<tr>
<td>CALR Exon 9 Mutation Analysis</td>
<td>81219</td>
</tr>
<tr>
<td>MPL Mutation Analysis</td>
<td>81402</td>
</tr>
<tr>
<td>ASXL1 Mutation Analysis</td>
<td>81479</td>
</tr>
<tr>
<td>EZH2 Mutation Analysis</td>
<td>81479</td>
</tr>
<tr>
<td>TET2 Mutation Analysis</td>
<td>81479</td>
</tr>
<tr>
<td>IDH1 Mutation Analysis</td>
<td>81403</td>
</tr>
<tr>
<td>IDH2 Mutation Analysis</td>
<td>81403</td>
</tr>
<tr>
<td>SRSF2 Mutation Analysis</td>
<td>81479</td>
</tr>
<tr>
<td>SF3B1 Mutation Analysis</td>
<td>81479</td>
</tr>
</tbody>
</table>

What are BCR-ABL negative myeloproliferative neoplasms

Definition

Primary myelofibrosis (PMF), polycythemia vera (PV) and essential thrombocythemia (ET) are a group of heterogeneous disorders of the hematopoietic system collectively known as Philadelphia chromosome-negative MPN.
Prevalence

The following table describes the prevalence of Philadelphia chromosome-negative MPNs in the U.S.¹

<table>
<thead>
<tr>
<th>Disorder</th>
<th>Prevalence in the U.S.</th>
</tr>
</thead>
<tbody>
<tr>
<td>PMF</td>
<td>13,000</td>
</tr>
<tr>
<td>ET</td>
<td>134,000</td>
</tr>
<tr>
<td>PV</td>
<td>148,000</td>
</tr>
</tbody>
</table>

Symptoms

Symptoms vary among the subtypes, but generally include
- constitutional symptoms
- fatigue
- pruritus
- weight loss
- symptoms of splenomegaly, and
- variable lab abnormalities, including
  - erythrocytosis
  - thrombocytosis, and
  - leukocytosis.¹

Risks

Individuals with MPNs are at risk of the condition transforming into acute myeloid leukemia (AML), which is associated with a poor response to therapy and short survival. These disorders are also associated with an increased risk of major bleeding and thrombosis or thromboembolism compared to the general population.¹

Diagnosis

The diagnosis and management of patients with MPN has evolved since the identification of mutations that activate the JAK pathway, including JAK2, CALR, and MPL. The development of targeted therapies has resulted in significant improvements in disease-related symptoms and quality of life.¹

- **JAK2 V617F mutations** — JAK2 V617F mutations account for the majority of patients with PV (greater than 90%), ET or PMF (60%). Most of the mutations occur in exon 14 with rare insertions/deletions in exon 12.¹
- **JAK2 exon 12 mutations** — JAK2 exon 12 mutations have been seen in approximately 2-3% of patients with PV.¹
• **MPL mutations** — MPL mutations have been reported in 5-8% of patients with PMF and 1-4% of patients with ET. MPL mutations are associated with lower hemoglobin levels at diagnosis and increased risk of transfusion dependence in patients with PMF.¹

• **CALR mutations** — CALR mutations are reported in approximately 20-35% of patients with ET and PMF, accounting for approximately 60-80% of patients with JAK2/MPL-negative ET and PMF. CALR deletion mutations are more commonly seen in patients with PMF and are associated with a significantly higher risk of myelofibrosis transformation in ET. CALR insertion mutations are associated with ET, low risk of thrombosis and an indolent course. CALR mutations are associated with a lower hemoglobin level, lower WBC count, higher platelet count and lower incidence of thrombosis than the JAK2 V617F mutation.¹

**Test information**

**Introduction**

Testing for BCR-ABL negative MPN may include cytogenetic testing, single gene mutation analysis, or panel testing.

**Types of tests**

There are various methods used to test for the cytogenetic and molecular abnormalities associated with MPN.¹,² Tests for the cytogenetic and molecular abnormalities include:

- bone marrow cytogenetics: karyotype, with or without FISH
- single gene mutation analysis for JAK2, MPL, and CALR, and
- panel testing using next generation sequencing.

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to BCR-ABL negative MPN testing.

**World Health Organization: PMF**

The World Health Organization (WHO, 2016) has established diagnostic criteria for PMF.²
### Pre Primary Myelofibrosis (prePMF)
[Diagnosis requires meeting all 3 major criteria, and at least 1 minor criterion]

**Major criteria:**
- Megakaryocytic proliferation and atypia, without reticulin fibrosis > grade 1, accompanied by increased age-adjusted BM cellularity, granulocytic proliferation, and often decreased erythropoiesis
- Not meeting WHO criteria for BCR-ABL1+ CML, PV, ET, myelodysplastic syndromes, or other myeloid neoplasms
- Presence of JAK2, CALR, or MPL mutation or in the absence of these mutations, presence of another clonal marker, or absence of minor reactive BM reticulin fibrosis

**Minor criteria:**
Presence of at least one of the following, confirmed in 2 consecutive determinations:
- Anemia not attributed to a comorbid condition
- Leukocytosis $\geq 11 \times 10^9/L$
- Palpable splenomegaly
- LDH increased to above upper normal limit of institutional reference range

### Overt Primary Myelofibrosis (overt PMF)
[Diagnosis requires meeting all 3 major criteria, and at least 1 minor criterion]

**Major criteria:**
- Megakaryocytic proliferation and atypia, accompanied by either reticulin and/or collagen fibrosis grades 2 or 3
- Not meeting WHO criteria for BCR-ABL1+ CML, PV, ET, myelodysplastic syndromes, or other myeloid neoplasms
- Presence of JAK2, CALR, or MPL mutation or in the absence of these mutations, presence of another clonal marker, or absence of reactive BM myelofibrosis

**Minor criteria:**
Presence of at least one of the following, confirmed in 2 consecutive determinations:
- Anemia not attributed to a comorbid condition
- Leukocytosis $\geq 11 \times 10^9/L$
- Palpable splenomegaly
- LDH increased to above upper normal limit of institutional reference range
- Leukoerythroblastosis

### Absence of 3 major clonal mutations

In the absence of any of the 3 major clonal mutations, the search for the most frequent accompanying mutations help determine the clonal nature of the disease. Examples of the most frequent accompanying mutations include:

- ASXL1
- EZH2

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www.eviCore.com
• TET2
• IDH1
• IDH2
• SRSF2, or
• SF3B1.

World Health Organization: PV

The World Health Organization (WHO, 2016) has established diagnostic criteria for PV.²

Polycythemia Vera (PV)
[Diagnosis requires meeting either all 3 major criteria, or the first 2 major criteria and the minor criterion]

Major criteria:
• Hemoglobin > 16.5 g/dL in men, > 16.0 g/dL in women OR Hematocrit >49% in men, >48% in women OR Increased red cell mass (RCM), defined as >25% above the mean normal predicted value
• Bone marrow biopsy showing hypercellularity for age with trilineage growth (panmyelosis) including prominent erythroid, granulocytic, and megakaryocytic proliferation with pleomorphic, mature megakaryocytes (difference in size)
• Presence of JAK2 V617F or JAK2 exon 12 mutation

Minor criteria:
• Subnormal serum EPO level

BM biopsy not required in some cases

A bone marrow biopsy may not be required in cases with sustained absolute erythrocytosis; hemoglobin levels >18.5 g/dL in men (hematocrit, 55.5%) or >16.5 g/dL in women (hematocrit, 49.5%) if major criterion 3 and the minor criterion are present. However, initial myelofibrosis (present in up to 20% of patients) can only be detected by performing a BM biopsy; this finding may predict a more rapid progression to overt myelofibrosis (post-PV PMF).

World Health Organization: ET

The World Health Organization (WHO, 2016) has established diagnostic criteria for ET.²
Essential Thrombocythemia (ET)

[Diagnosis requires meeting all 4 major criteria or the first 3 major criteria and the minor criterion]

Major criteria:
- Platelet count ≥ 450 x 10^9/L
- Bone marrow biopsy showing proliferation mainly of the megakaryocyte lineage with increased numbers of enlarged, mature megakaryocytes with hyperlobulated nuclei. No significant increase or left shift in neutrophil granulopoiesis or erythropoiesis and very rarely minor (grade 1) increase in reticulin fibers
- Not meeting WHO criteria for BCR-ABL1+ CML, PV, PMF, myelodysplastic syndromes, or other myeloid neoplasms
- Presence of JAK2, CALR, or MPL mutation

Minor criteria:
- Presence of a clonal marker or absence of evidence for reactive thrombocytosis

National Comprehensive Cancer Network

The National Comprehensive Cancer Network (NCCN, 2017) evidence and consensus-based guidelines recommend the following initial laboratory evaluations for individuals suspected to have MPN:¹

- “Laboratory evaluations should include complete blood count (CBC), microscopic examination of the peripheral smear, comprehensive metabolic panel with serum uric acid, serum LDH, liver function tests, serum EPO level and serum iron studies.”
- “Fluorescence in situ hybridization (FISH) or a reverse transcriptase polymerase chain reaction (RT-PCR) on a peripheral blood specimen to detect BCR-ABL1 transcripts and exclude the diagnosis of CML is recommended for all patients, especially those with left-shifted leukocytosis and/or thrombocytosis with basophilia.”
- “Molecular testing for JAKV617F mutations should be performed in all patients. If JAKV617F mutation is negative, molecular testing for MPL and CALR mutations should be performed for patients with MF and ET; molecular testing for JAK2 exon 12 mutation should be done for those with PV.”
- “In the absence of JAK2, CALR and MPL mutations, the presence of another clonal marker is included as one of the major diagnostic criteria for PMF. Additional mutations in ASXL1, EZH2, TET2, IDH1, IDH2, SRSF2, and SF3B1 genes are noted to be of help in determining the clonal nature of the disease.”
• “Bone marrow aspirate and biopsy with trichrome and reticulin stain and bone marrow cytogenetics (karyotype, with or without FISH) is necessary to accurately distinguish the bone marrow morphological features between the disease subtypes (early/prefibrotic PMF, ET and masked PV).”

Criteria

JAK2 V617F Mutation Analysis

• Member does not meet WHO criteria for BCR-ABL1+ CML, myelodysplastic syndromes, or other myeloid neoplasms, AND
• Member meets at least ONE of the following diagnostic criteria for MPN:
  o Bone marrow biopsy results that are consistent with WHO diagnostic criteria for prePMF, overt PMF, ET, or PV, or
  o Platelet count $\geq 450 \times 10^9$/L, or
  o Hemoglobin > 16.5 g/dL in men, > 16.0 g/dL in women, or
  o Hematocrit >49% in men, >48% in women, or
  o Increased red cell mass (RCM), defined as $>25\%$ above the mean normal predicted value, or
  o A combination of two of the following symptoms:
    ▪ Anemia not attributed to a comorbid condition, or
    ▪ Leukocytosis $\geq 11 \times 10^9$/L, or
    ▪ Palpable splenomegaly, or
    ▪ LDH increased to above upper normal limit of institutional reference range, or
    ▪ Leukoerythroblastosis, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

JAK2 Exon 12 Analysis

• Member does not meet WHO criteria for BCR-ABL1+ CML, myelodysplastic syndromes, or other myeloid neoplasms, AND
• JAK2 V617F mutation analysis is negative, AND
• Member meets at least ONE of the following diagnostic criteria for PV:
  o Bone marrow biopsy results that are consistent with WHO diagnostic criteria for PV, or
- Hemoglobin > 16.5 g/dL in men, > 16.0 g/dL in women, or
- Hematocrit >49% in men, >48% in women, or
- Increased red cell mass (RCM), defined as >25% above the mean normal predicted value, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

CALR Exon 9 and MPL Mutation Analysis

- Member does not meet WHO criteria for BCR-ABL1+ CML, PV, myelodysplastic syndromes, or other myeloid neoplasms, AND
- JAK2 V617F mutation analysis is negative, AND
- Member meets at least ONE of the following diagnostic criteria for ET or PMF:
  - Bone marrow biopsy results that are consistent with WHO diagnostic criteria for prePMF, overt PMF, or ET, or
  - Platelet count ≥ 450 x 10^9/L, or
  - A combination of two of the following symptoms:
    - Anemia not attributed to a comorbid condition, or
    - Leukocytosis ≥ 11 x 10^9/L, or
    - Palpable splenomegaly, or
    - LDH increased to above upper normal limit of institutional reference range, or
    - Leukoerythroblastosis, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

Analysis of ASXL1, EZH2, TET2, IDH1, IDH2, SRSF2, and /or SF3B1

- Member does not meet WHO criteria for BCR-ABL1+ CML, PV, ET, myelodysplastic syndromes, or other myeloid neoplasms, AND
- JAK2, CALR, and MPL mutation analyses are all negative, AND
- Member meets at least ONE of the following diagnostic criteria for PMF:
  - Bone marrow biopsy results that are consistent with WHO diagnostic criteria for prePMF or overt PMF, or
  - A combination of two of the following symptoms:
    - Anemia not attributed to a comorbid condition, or
    - Leukocytosis ≥ 11 x 10^9/L, or
- Palpable splenomegaly, or
- LDH increased to above upper normal limit of institutional reference range, or
- Leukoerythroblastosis, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

References

Introduction

These references are cited in this guideline.


BCR-ABL Testing for Chronic Myelogenous Leukemia

Introduction

BCR-ABL chronic myelogenous leukemia (CML) testing is addressed by this guideline.

Procedures addressed

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What is chronic myelogenous leukemia

Definition

Chronic myelogenous leukemia (CML) is a hematopoietic stem cell disease that results in overgrowth of white blood cells in the bone marrow. It is defined by the presence of the chromosome (Ph), a translocation between chromosomes 9 and 22 that results in the fusion of two genes known as BCR and ABL.1,2

CML phases

The three phases of CML are chronic, accelerated, and blastic. In the chronic phase, there are few symptoms and most people are diagnosed after a routine blood test reveals the characteristic blood count and differential. If not treated, the disease will progress to the accelerated and blastic phases, symptoms of which include

- fever
- bone pain
- splenomegaly
- fatigue, and
• weakness.¹

Diagnosis

Detection of the BCR-ABL fusion gene is diagnostic for CML and Ph+ ALL and can be established by fluorescent in situ hybridization (FISH) or quantitative real-time polymerase chain reaction (QPCR).²

Acute lymphoblastic leukemia (ALL) is a different form of leukemia, but may also be positive for the chromosome (Ph+). About 3% of pediatric ALL and 25% of adult ALL is Ph+.³

Treatment

First-line treatment for CML and some Ph+ ALL is with a class of drugs called tyrosine kinase inhibitors (TKIs), which block the production of the BCR-ABL fusion gene protein product. Three TKI therapies are available as first-line therapies:

• imatinib (Gleevec®)
• nilotinib (Tasigna®), and
• dasatinib (Sprycel®).

These TKI therapies have all demonstrated proven benefit, and median survival is expected to approach normal life expectancy for most patients with CML.¹²

Treatment response

Monitoring of patients for treatment response to TKIs includes routine measurement of the BCR-ABL fusion gene protein product via QPCR prior to initiation of treatment and during treatment every 3 months.²

Treatment resistance

For individuals who display apparent treatment resistance, consideration of alternative treatment options may be appropriate.² Treatment resistance in both CML and ALL can be caused by mutations in the BCR-ABL kinase domain.²³

Test information

Introduction

Testing for chronic myelogenous leukemia (CML) may include qPCR for BCR-ABL transcript levels or FISH for t(9;22) BCR-ABL.
qPCR for BCR-ABL transcript levels

Bone marrow cytogenetics and measurement of BCR-ABL transcript levels by quantitative polymerase chain reaction is recommended before initiation of treatment as well as for assessing response to therapy.

FISH for t(9;22) BCR-ABL

If collection of bone marrow is not feasible, fluorescence in situ hybridization (FISH) on peripheral blood specimen using dual probes for the BCR and ABL genes is an acceptable method of confirming the diagnosis of CML.4

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to BCR-ABL testing for chronic myeloid leukemia.

National Comprehensive Cancer Network

The National Comprehensive Cancer Network (NCCN, 2017) recommends bone marrow cytogenetics to confirm a diagnosis of CML. If bone marrow is not available, FISH on a peripheral blood specimen using probes for both BCR and ABL can confirm the diagnosis.2

The NCCN recommends BCR-ABL transcript levels be obtained by quantitative RT-PCR in the following scenarios:

• at diagnosis
• every three months after initiating treatment. After a patient reaches complete cytogenetic response, every 3 months for two years, and every 3-6 months thereafter
• If a patient has a rising level of BCR-ABL transcripts (1 log increase), repeat testing in 1–3 months

These recommendations are category 2A which is “Based upon lower-level evidence, there is uniform NCCN consensus that the intervention is appropriate.”

Criteria

BCR-ABL transcript level testing is indicated in individuals at the initiation of treatment and at regular intervals (ranges from every month to once every 3-6 months) during treatment with ANY of the following drug therapies:

• Imatinib (Gleevec®)
• Nilotinib (Tasigna®)
• Dasatinib (Sprycel®)

References

Introduction

These references are cited in this guideline.


Introduction

Bloom syndrome testing is addressed by this guideline.

Procedures addressed

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What is Bloom syndrome

Definition

Bloom syndrome is a genetic disorder in which an individual's chromosomes contain large breaks, gaps, or rearrangements.¹

Symptoms

Affected individuals are usually smaller than average and suffer from a variety of symptoms:¹-²

- Intrauterine growth retardation that persists into childhood.
- Long, narrow face, small lower jaw, and prominent nose and ears.
- Sensitivity to sunlight. Exposure to sunlight causes a characteristic butterfly-shaped rash on the face.
- Chronic lung problems, diabetes, and immune deficiencies.
- Cancer diagnosis at an early age.
Learning disabilities.

Prevalence

Fewer than 300 cases of Bloom syndrome have been reported since the disease was first described over 50 years ago.\(^2,3\)

About 1 in 48,000 Ashkenazi Jewish individuals have Bloom syndrome, and 25% of all affected individuals have Ashkenazi Jewish ancestry.\(^2\)

Prognosis

There is no cure for Bloom syndrome. Treatment involves continuous monitoring by multiple physicians and specialists.\(^1,3\)

Cause

Bloom syndrome is caused by a genetic mutation in the BLM gene.\(^1,3,4\)

BLM is essential to maintaining the stability of chromosomes during DNA replication and cell division.\(^3,4\)

Mutations in BLM lead to mistakes during cellular replication.\(^3,4\)

Individuals with Bloom syndrome have multiple breaks, gaps, and genetic rearrangements in their chromosomes, leading to a unique combination of signs and symptoms.\(^3,4\)

Diagnosis

A diagnosis of Bloom syndrome is suspected when the patient presents with the characteristic suite of signs and symptoms. This diagnosis can be confirmed by genetic testing and may be needed to differentiate between other disorders with overlapping symptoms. There are several types of tests available for diagnostic purposes.

Inheritance

Bloom syndrome is an autosomal recessive disorder, meaning that an affected individual must inherit BLM gene mutations from each parent.\(^1,4\)

Individuals who inherit only one mutation are called carriers. Carriers do not show symptoms of Bloom syndrome, but have a 50% chance of passing on the mutation to their children.

Two carriers of Bloom syndrome have a 1 in 4 (25%) chance for each pregnancy to be affected with Bloom syndrome.
Test information

Introduction

Testing for Bloom syndrome may include sister chromatid exchange, known familial mutation analysis, targeted mutation analysis, sequence analysis, or deletion/duplication analysis.

Sister Chromatid Exchange

SCE is the standard analysis for diagnosis of Bloom syndrome. The method involves exposing an individual's cells to bromodeoxyuridine (BrdU), a compound that helps identify which cells contain chromosomes with unusually large numbers of rearrangements, or “exchanges.” Individuals with Bloom syndrome will have a substantially higher number of these exchanges compared with unaffected individuals. This test can be used for prenatal diagnosis of at-risk pregnancies on chorionic villi or amniocytes.

BLM Known Familial Mutation Analysis

Once a deleterious mutation has been identified in an affected person, relatives and at-risk pregnancies can be tested.

BLM Targeted Mutation Analysis

This test looks for the BLM gene mutation most often found in Ashkenazi Jewish individuals, called blmAsh. The detection rate of this mutation in Ashkenazi Jewish individuals is greater than 97%.

BLM Sequencing

This test looks for mutations across the entire gene, and can identify at least 87% of disease-causing mutations in individuals with non-Jewish Ancestry and greater than 99% of disease-causing mutations in Ashkenazi Jewish individuals. It is typically used only for diagnosis of an affected individual or carrier testing of a non-Ashkenazi Jewish individual when the partner is a known carrier.

BLM Deletion/Duplication Analysis

This test looks for deletions and duplication in the gene that would not be detected by sequencing analysis. It is typically performed in reflex to sequencing analysis when there is a high suspicion for disease.
Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to Bloom syndrome testing.

Diagnostic testing strategy

A 2016 expert-authored review suggests the following diagnostic testing strategy:\textsuperscript{4}

“The diagnosis of Bloom Syndrome (Bsyn) is established in a proband with identification of biallelic pathogenic variants in \textit{BLM} on molecular genetic testing or, if molecular genetic testing is inconclusive, with identification of increased frequency of sister-chromatid exchanges (SCEs) on specialized cytogenetic studies.”

Carrier testing strategy

The American College of Medical Genetics (ACMG, 2008)\textsuperscript{6} and the American College of Obstetrics and Gynecologists (ACOG, 2009)\textsuperscript{7} support offering carrier testing for Bloom syndrome to individuals of Ashkenazi Jewish descent for the common \textit{blmAsh} mutation.

\begin{itemize}
  \item Guidelines support the testing of individuals of Ashkenazi Jewish descent, even when their partner is non-Ashkenazi Jewish. In this situation, testing would start with the individual who is Jewish and if \textit{blmAsh} mutation is detected, sequencing of \textit{BLM} in the non-Ashkenazi Jewish partner would follow.\textsuperscript{6} If the woman is pregnant, testing may need to be conducted on both partners simultaneously in order to receive results in a timely fashion.\textsuperscript{7}

  \item If one or both partners are found to be carriers of Bloom syndrome, genetic counseling should be provided and prenatal testing offered, if appropriate.
\end{itemize}

Prenatal testing strategy

A 2016 expert-authored review states:\textsuperscript{4}

\begin{itemize}
  \item “Prenatal diagnosis for pregnancies at increased risk is possible by sister-chromatid exchange (SCE) analysis of fetal cells obtained by amniocentesis usually performed at about 15 to 18 weeks’ gestation or chorionic villus sampling (CVS) at approximately ten to 12 weeks’ gestation.”

  \item “If the \textit{BLM} pathogenic variants have been identified in an affected family member, prenatal testing for pregnancies at increased risk may be available from a clinical laboratory that offers either testing of this gene or custom prenatal testing.”
\end{itemize}
• “Preimplantation genetic diagnosis (PGD) has been successfully utilized for one couple [Bloom's Syndrome Registry, unpublished data], and may be an option for some families in which the BLM pathogenic variants have been identified.”

Criteria

Introduction

Requests for Bloom syndrome testing are reviewed using these criteria.

Sister Chromatid Exchange (Chromosome Analysis for Breakage Syndromes)

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous sister chromatid exchange analysis performed, and
  o No previous BLM full sequencing, or BLM sequencing performed and only one mutation identified, and
  o No known BLM mutation in biologic relative, and
  o If Ashkenazi Jewish, targeted mutation analysis performed and no mutation detected or one mutation detected, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Unexplained severe intrauterine growth retardation that persists throughout infancy and childhood (less than 5th percentile), or
  o An unusually small individual (less than 5th percentile) who develops erythematous skin lesions in the “butterfly area” of the face after sun exposure, or
  o An unusually small individual (less than 5th percentile) who develops a malignancy OR

• Prenatal Testing for At-Risk Pregnancies:
  o Known increased risk due to affected first-degree relative, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

BLM Known Familial Mutation Analysis

• Genetic Counseling:
• Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing
  o No previous genetic testing of BLM, AND

• Carrier Screening:
  o Known family mutation in BLM identified in 1st, 2nd, or 3rd degree biologic relative(s), OR

• Prenatal Testing for At-Risk Pregnancies:
  o BLM mutation identified in both biologic parents, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**BLM Targeted Mutation Analysis**

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous BLM genetic testing, including Ashkenazi Jewish screening panels containing targeted mutation analysis for blmAsh, AND

• Carrier Screening:
  o Ashkenazi Jewish descent, and
  o Have the potential and intention to reproduce, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**BLM Sequencing**

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous BLM full sequencing, and
  o No known BLM mutation in biologic relative, and
• If Ashkenazi Jewish, targeted mutation analysis performed and no mutation detected or one mutation detected, AND

• Diagnostic Testing for Symptomatic Individuals:
  • Unexplained severe intrauterine growth retardation that persists throughout infancy and childhood (less than 5th percentile), or
  • An unusually small individual (less than 5th percentile) who develops erythematous skin lesions in the “butterfly area” of the face after sun exposure, or
  • An unusually small individual (less than 5th percentile) who develops a malignancy, OR

• Testing for Individuals with Family History or Partners of Carriers:
  • 1st, 2nd, or 3rd degree biologic relative with Bloom syndrome clinical diagnosis, family mutation unknown, and testing unavailable, or
  • Partner is monoallelic or biallelic for BLM mutation, and
  • Have the potential and intention to reproduce, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

BLM Deletion/Duplication Analysis

• Genetic Counseling:
  • Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  • Previous BLM full sequencing, and no mutations or only one mutation detected, AND

• Diagnostic Testing for Symptomatic Individuals:
  • Unexplained severe intrauterine growth retardation that persists throughout infancy and childhood (less than 5th percentile), or
  • An unusually small individual (less than 5th percentile) who develops erythematous skin lesions in the “butterfly area” of the face after sun exposure, or
  • An unusually small individual (less than 5th percentile) who develops a malignancy, OR

• Testing for Individuals with Family History or Partners of Carriers:
  • 1st, 2nd, or 3rd degree biologic relative with Bloom syndrome clinical diagnosis, family mutation unknown, and testing unavailable, or
- Partner is monoallelic or biallelic for BLM mutation, and
- Have the potential and intention to reproduce, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

References

Introduction

These references are cited in this guideline.


BRAF Testing for Colorectal Cancer

Introduction

BRAF testing for colorectal cancer is addressed by this guideline.

Procedures addressed

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What is BRAF

Definition

BRAF is a gene that forms a part of a cell-signaling pathway to help control cell growth. Changes or mutations in the BRAF gene can cause out of control cell growth, which may lead to cancer. The most common BRAF mutation is called V600E which was previously known as V599E.

Prevalence

About 5-9% of colorectal cancer tumors have a V600E BRAF mutation.

Prognosis

Patients with a V600E BRAF mutation appear to have a poorer prognosis. Tumors with BRAF mutations may have less response to anti-EGFR therapies like cetuximab (Erbitux®) and panitumumab (Vectibix®).

Test information

Introduction

Testing for a BRAF mutation may include targeted mutation analysis or sequencing.
Available tests

The following BRAF tests are available to identify mutations.

**Note**  BRAF mutation analysis has several other test applications with different criteria (such as melanoma therapeutic response, Lynch syndrome tumor screening, or Noonan syndrome diagnosis). Ensure you are reviewing the correct use of the test.

**Targeted mutation analysis**

Laboratories most commonly test for the BRAF V600E mutation, which accounts for about 90% of activating BRAF mutations.\(^2\) Mutation analysis requires relatively little tumor material for testing and has high sensitivity. It is also relatively inexpensive.\(^3,4\)

BRAF mutation analysis is done on fresh, frozen, or paraffin-embedded tissue from either a primary tumor or metastasis.\(^1,3,4\) Some molecular diagnostic laboratories perform BRAF mutation analysis by laboratory-developed methods, while others use FDA-approved test kits. Laboratory-developed tests may vary in the specimen type required, methodology used, mutations tested, sensitivity, and other test-specific data.

**Sequencing**

Some laboratories sequence all or part of the BRAF gene, which will find a broader spectrum of mutations than targeted mutation analysis. Laboratories that offer sequencing generally do so for a subset of exons where most BRAF activating mutations have been identified.

Sequence analysis requires more and higher quality tumor material for testing than targeted mutations. This method is typically less efficient and more expensive than targeted mutation analysis.\(^3-5\)

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to BRAF testing for colorectal cancer.

**National Comprehensive Cancer Network**

The National Comprehensive Cancer Network (NCCN, 2017) states the following.\(^1\)

“Limited data from unplanned retrospective subset analyses of patients with metastatic colorectal cancer treated in the first-line setting suggest that although BRAF V600E mutation confers a poor prognosis regardless of treatment, patients with disease characterized by this mutation may receive some benefit from the addition of cetuximab to front-line therapy.”
“Overall, the panel believes that evidence increasingly suggests that BRAF V600E mutation makes response to panitumumab or cetuximab, as single agents or in combination with cytotoxic chemotherapy, highly unlikely. The panel recommends BRAF genotyping of tumor tissue (either primary tumor or metastasis) at diagnosis of stage V1 disease.”

“Despite uncertainty over its role as a predictive marker, it is clear that mutations in BRAF are a strong prognostic marker.”

“Although BRAF genotyping can be considered for patients with tumors characterized by the wild-type KRAS/NRAS genes, this testing is currently optional and not a necessary part of deciding whether to use anti-EGFR agents.”

“The panel strongly recommends genotyping of tumor tissue (either primary tumor or metastasis) in all patients with metastatic colorectal cancer for RAS (KRAS exon 2 or non-exon 2; NRAS) and BRAF at diagnosis of stage IV disease.”

“Testing for KRAS, NRAS, and BRAF mutations should be performed only in laboratories that are certified under the clinical laboratory improvement amendments of 1988 (CLIA-88) as qualifies to perform high complexity clinical laboratory (molecular pathology) testing. No specific methodology is recommended (eg, sequencing, hybridization).”

Criteria

Testing may be considered in individuals who meet the following criteria:

• Individual has been diagnosed with stage IV, metastatic colorectal cancer, AND

• BRAF mutation testing is needed for prognostic purposes.

The following BRAF mutation testing application is considered investigational and experimental:

• BRAF mutation testing for the purpose of decision making regarding the use of anti-EGFR agents.

References

Introduction

These references are cited in this guideline.


BRAF Testing for Melanoma Kinase Inhibitor Response

Introduction

BRAF testing for melanoma kinase inhibitor response is addressed by this guideline.

Procedures addressed

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What is BRAF

Definition

BRAF is a gene that forms a part of a cell-signaling pathway to help control cell growth. Changes or mutations in the BRAF gene can cause out of control cell growth, which may lead to cancer.\(^1,2\)

**BRAF V600E**

The most common BRAF mutation is called V600E which was previously known as V599E. This mutation accounts for about 70-90% of mutations in this gene.\(^1,3\)

About 40-60% of cutaneous melanomas have a V600E BRAF mutation.\(^1\)

Treatment options

Vemurafenib (Zelboraf\(^\text{®}\)), dabrafenib (Tafinlar\(^\text{®}\)), and trametinib (Mekinist\(^\text{®}\)) are orally-administered kinase inhibitors that are able to block the function of the mutated BRAF protein.\(^1,2\) They are specifically indicated for the treatment of patients with metastatic or unresectable melanoma whose tumors have a BRAF V600E mutation.\(^1,2\) They are not recommended for use in patients with wild type BRAF melanoma.\(^2,4-6\)

Test information

Introduction

Laboratories most commonly perform targeted mutation analysis for the BRAF V600E.
mutation, which accounts for about 90% of activating BRAF mutations.\[^4\] Mutation analysis requires relatively little tumor material for testing and has high sensitivity. It is also relatively inexpensive.\[^2,3\]

**Targeted mutation analysis**

BRAF mutation analysis is done on fresh, frozen, or paraffin-embedded tissue from either a primary tumor or metastasis.\[^1-3\] Some molecular diagnostic laboratories perform BRAF mutation analysis by laboratory-developed methods, while others use FDA-approved test kits. Laboratory-developed tests may vary in the specimen type required, methodology used, mutations tested, sensitivity, and other test-specific data.

**Vemurafenib**

Vemurafenib was approved in 2011 for use along with an FDA approved companion diagnostic developed by Roche Molecular Diagnostics called the cobas® 4800 BRAF V600 Mutation Test.

The cobas 4800 BRAF V600 mutation test was clinically validated in the trials conducted for approval of vemurafenib. This testing specifically checks for the V600E mutation in formalin-fixed, paraffin-embedded melanoma tumor tissue.\[^2\]

**Dabrafenib and trametinib**

In 2013, dabrafenib and trametinib were approved for use along with an FDA approved companion diagnostic developed by Roche Molecular Diagnostics called the THxID™ BRAF test. The THxID BRAF test was clinically validated in the clinical studies supporting the approval of dabrafenib and trametinib.

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to BRAF testing for melanoma kinase inhibitor response.

**National Comprehensive Cancer Network**

The National Comprehensive Cancer Network (NCCN, 2016) includes vemurafenib, dabrafenib, dabrafenib in combination with trametinib, and vemurafenib in combination with cobimetinib as options for the treatment of advanced or metastatic melanoma which require companion diagnostic testing.

The guidelines state that “For patients with BRAF-mutant metastatic disease, BRAF-targeted therapy first-line options include BRAF/MEK inhibitor combination therapy with dabrafenib/trametinib or vemurafenib/cobimetinib, or single-agent BRAF inhibitor therapy with vemurafenib or dabrafenib.”\[^4\]
FDA

The FDA approved each of the following drugs with a companion diagnostic:

- **Zelboraf (vemurafenib)** — “ZELBORAF® is a kinase inhibitor indicated for the treatment of patients with unresectable or metastatic melanoma with BRAF V600E mutation as detected by an FDA-approved test. Limitation of Use: ZELBORAF is not indicated for treatment of patients with wild-type BRAF melanoma.”

- **Tafinlar (dabrafenib)** — “TAFINLAR is a kinase inhibitor indicated as a single agent for the treatment of patients with unresectable or metastatic melanoma with BRAF V600E mutation as detected by an FDA-approved test. TAFINLAR in combination with trametinib is indicated for the treatment of patients with unresectable or metastatic melanoma with BRAF V600E or V600K mutations as detected by an FDA-approved test. The use in combination is based on the demonstration of durable response rate. Improvement in disease-related symptoms or overall survival has not been demonstrated for TAFINLAR in combination with trametinib… Limitation of Use: TAFINLAR is not indicated for treatment of patients with wild-type BRAF melanoma.”

- **Mekinist (trametinib)** — “MEKINIST is a kinase inhibitor indicated as a single agent for the treatment of patients with unresectable or metastatic melanoma with BRAF V600E or V600K mutations as detected by an FDA-approved test. MEKINIST is indicated, in combination with dabrafenib, for the treatment of patients with unresectable or metastatic melanoma with BRAF V600E or V600K mutations as detected by an FDA-approved test. Limitation of use: MEKINIST is not indicated for treatment of patients who have progressed on prior BRAF-inhibitor therapy.”

- **Cotellic (cobimetinib)** — “COTELLIC is a kinase inhibitor indicated for the treatment of patients with unresectable or metastatic melanoma with a BRAF V600E or V600K mutation, in combination with vemurafenib. Limitation of use: COTELLIC is not indicated for treatment of patients with wild-type BRAF melanoma.”

**Criteria**

Testing may be considered in individuals who meet the following criteria:

- Individual has been diagnosed with metastatic or unresectable melanoma, AND
- At least one of the following treatments is being considered: Zelboraf® (vemurafenib), Tafinlar® (dabrafenib), Mekinist® (trametinib) in combination with Tafinlar® (dabrafenib), or Zelboraf® (vemurafenib) in combination with Cotellic®(cobimetinib), AND
• BRAF V600 testing has not been performed previously

Exclusions

BRAF V600E tumor marker testing is not currently indicated as a companion diagnostic or for therapy selection for any other tumor types and is therefore considered investigational and experimental.

References

Introduction

These references are cited in this guideline.


Introduction

BRCA analysis is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
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</tr>
</thead>
<tbody>
<tr>
<td>BRCA1 Known Familial Mutation Analysis</td>
<td>81215</td>
</tr>
<tr>
<td>BRCA2 Known Familial Mutation Analysis</td>
<td>81217</td>
</tr>
<tr>
<td>BRCA1/2 Sequencing</td>
<td>81211</td>
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<tr>
<td>BRCA1 Sequencing</td>
<td>81214</td>
</tr>
<tr>
<td>BRCA2 Sequencing</td>
<td>81216</td>
</tr>
<tr>
<td>BRCA1/2 Full Sequencing and Deletion/Duplication Analysis (Combined)</td>
<td>81162</td>
</tr>
<tr>
<td>BRCA1/2 Deletion/Duplication Analysis</td>
<td>81213</td>
</tr>
</tbody>
</table>

What is hereditary breast and ovarian cancer

Definition

Hereditary breast and ovarian cancer (HBOC) is an inherited form of cancer.

Characteristics of HBOC

HBOC is characterized by any of the following:\(^1,^2\)

- personal history of
  - breast cancer at a young age, typically under age 50
  - two primary breast cancers
  - both breast and ovarian cancer
  - triple negative breast cancer (ER-, PR-, HER2-)
  - ovarian, fallopian tube, or primary peritoneal cancer, or
- metastatic prostate cancer
- multiple cases of breast or ovarian cancer in a family
- personal or family history of
  - male breast cancer
  - pancreatic cancer with breast or ovarian cancer, or
  - prostate cancer with a Gleason score of at least 7 and a family history of ovarian, breast, prostate, or pancreatic cancer
- previously identified BRCA1 or BRCA2 mutation in the family, or
- any of the above with Ashkenazi Jewish ancestry.

**Inheritance**

Up to 10% of all breast cancer and 15% of all ovarian cancer is associated with an inherited gene mutation, with BRCA1 and BRCA2 accounting for about 20-25% of all hereditary cases.\(^1\)\(^,\)\(^3\)\(^-\)\(^5\)

BRCA mutations are inherited in an autosomal dominant manner. When a parent has a BRCA mutation, each offspring have a 50% risk of inheriting the mutation.\(^1\)

**Prevalence**

About 1 in 400 people in the general population has a BRCA1 or BRCA2 mutation. The prevalence of mutations is higher in people of Norwegian, Dutch, or Icelandic ethnicity.\(^1\)\(^,\)\(^3\)

The prevalence of BRCA mutations varies among African Americans, Hispanics, Asian Americans, and non-Hispanic whites.\(^3\)

**Ashkenazi Jewish ancestry**

About 1 in 40 people of Ashkenazi Jewish ancestry has a BRCA1 or BRCA2 mutation. The majority of the risk in the Ashkenazi Jewish population is associated with three common founder mutations, two of which are in the BRCA1 gene and one in the BRCA2 gene.\(^1\)\(^,\)\(^6\)\(^,\)\(^7\) These three mutations account for 99% of identified mutations in the Ashkenazi Jewish population.\(^1\)

**Cancer risks**

People with a BRCA mutation have an increased risk of various types of cancer.\(^1\)

<table>
<thead>
<tr>
<th>Type of cancer</th>
<th>Risk</th>
</tr>
</thead>
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<tr>
<td>Breast cancer</td>
<td>38-87%</td>
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<tr>
<td>Ovarian cancer</td>
<td>16.5-63%</td>
</tr>
<tr>
<td>Type of cancer</td>
<td>Risk</td>
</tr>
<tr>
<td>------------------------</td>
<td>-------------------</td>
</tr>
<tr>
<td>Male breast cancer</td>
<td>1-9%</td>
</tr>
<tr>
<td>Prostate cancer</td>
<td>up to 20%</td>
</tr>
<tr>
<td>Pancreatic cancer</td>
<td>1-7%</td>
</tr>
<tr>
<td>Melanoma</td>
<td>Increased risk with BRCA2</td>
</tr>
</tbody>
</table>

**Note**  The risk for breast and ovarian cancer varies among family members and between families.

**Screening and prevention**

Screening and prevention options are available to specifically address the increased risk of these cancers in a person with a BRCA mutation.\(^1\)

**Breast cancer risk and other genes**

Other inherited cancer syndromes that can include breast cancer are Li-Fraumeni syndrome (TP53 gene), Cowden syndrome (PTEN), Hereditary Diffuse Gastric Cancer syndrome (CDH1), and Peutz Jeghers syndrome (STK11). Additionally, other genes that can increase the risk for breast cancer are ATM, CHEK2, and PALB2.\(^1,6,8,9\)

**Test information**

**Introduction**

BRCA testing may include full gene sequencing, deletion/duplication analysis, known familial mutation analysis, or multigene panel testing.

**Sequence analysis**

Full sequence analysis of BRCA1/2 genes looks at all of the coding regions of the BRCA1/2 genes, and often includes analysis of five common BRCA1/2 gene duplications and deletions.

Full sequence testing is typically appropriate as an initial test for people who meet criteria and do NOT have Ashkenazi Jewish ancestry.\(^1,6\)

**Deletion/duplication analysis**

Deletion/duplication analysis looks for large rearrangements, duplications, and deletions in the BRCA1/2 genes.
**Known familial mutation testing**

Known familial mutation testing looks for a specific mutation in either the BRCA1/2 gene previously identified in a family member.

This test is appropriate for those who have a known BRCA mutation in the family and are not Ashkenazi Jewish.

---

**Note** Founder mutation testing may be appropriate for those with Ashkenazi Jewish ancestry, even with a known familial mutation, since these mutations are common enough that multiple mutations can be found in the same Ashkenazi Jewish individual or family. If the familial mutation is not one of the three Ashkenazi Jewish mutations, then known familial mutation analysis for that mutation should be performed in addition to the founder mutation panel.\(^1\,^6\)

---

**Ashkenazi Jewish founder mutation testing**

Ashkenazi Jewish founder mutation testing includes the three mutations most commonly found in the Ashkenazi Jewish population: 187delAG and 5385insC in BRCA1 and 6174delT in BRCA2.\(^1\)

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**Cancer multigene panels**

BRCA1/2 gene testing is also available in the form of multigene panels for individuals with a personal or family history of cancer suggestive of more than one hereditary cancer syndrome.

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**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to BRCA analysis.

**National Comprehensive Cancer Network**

The National Comprehensive Cancer Network (NCCN, 2017)\(^6\) evidence and consensus-based guidelines address test indications for those with a personal history of HBOC-related cancers, those with a known mutation in the family, and unaffected individuals with a family history of HBOC-related cancer.

These recommendations are Category 2A, defined as “lower-level evidence with uniform NCCN consensus.”

Based on these guidelines, and the recommendations of the National Society of Genetic Counselors (2013),\(^6\) BRCA sequence analysis is appropriate in individuals with a personal and/or family history of cancer when any of the following criteria are met:
• personal history of breast cancer plus one or more of the following in non-Ashkenazi Jewish individuals:
  o diagnosed at age 45 years or younger
  o diagnosed at age 50 or younger with at least one close blood relative with breast cancer diagnosed at any age
  o diagnosed at age 60 years or younger with a triple negative (ER-, PR-, HER2-) breast cancer
  o two breast primaries when the first breast cancer diagnosis occurred at age 50 or younger
  o diagnosed at any age with at least one close blood relative with breast cancer diagnosed at age 50 years or younger and/or at least one close blood relative diagnosed with epithelial ovarian, fallopian tube or primary peritoneal cancer at any age
  o diagnosed at any age with two or more close blood relatives with breast cancer, pancreatic cancer, or prostate cancer (Gleason score at least 7) at any age
  o diagnosed at age 50 years or younger with at least one close blood relative with pancreatic cancer or prostate cancer (Gleason score at least 7)
  o diagnosed at age 50 years or younger with an unknown or limited family history, or
  o diagnosed at any age with a close male relative with breast cancer, or

• personal history of epithelial ovarian/fallopian tube/primary peritoneal cancer (with or without a history of breast cancer)

• personal history of male breast cancer

• personal history of pancreatic cancer or prostate cancer (Gleason score at least 7) at any age with at least one close blood relative with ovarian cancer at any age or breast cancer diagnosed at age 50 or younger or two close blood relatives with breast, pancreatic, or prostate cancer (Gleason score at least 7) at any age

• personal history of pancreatic cancer and Ashkenazi Jewish ancestry

• personal history of metastatic prostate cancer (radiographic evidence of or biopsy-proven disease)

• family history only, no personal diagnosis of cancer plus one of the following:
  o first- or second-degree blood relative meeting any of the above criteria, or
  o third-degree blood relative with breast and/or ovarian cancer and 2 or more close blood relatives with breast cancer (at least one diagnosed at or before age 50) and/or ovarian, primary peritoneal, or fallopian tube cancer.

• Ashkenazi Jewish individuals who are negative for founder mutation testing, and have a high pre-test probability of carrying a BRCA mutation\textsuperscript{1,6}
BRCA1/2 mutation detected by tumor profiling in the absence of germline mutation analysis.\textsuperscript{6}

**Testing unaffected individuals**

NCCN states “Testing of unaffected individuals should only be considered when an appropriate affected family member is unavailable for testing.” They caution that the significant limitations in interpreting results from unaffected relatives must be discussed.

**Close blood relatives**

Close blood relatives include: first-degree relatives (parents, siblings, children); second-degree relatives (aunts, uncles, grandparents, grandchildren, nieces, nephews and half-siblings); and third-degree relatives (great-grandparents, great-aunts, great-uncles, and first cousins) on the same side of the family.

**Limited family history**

Limited family history is defined as “fewer than two first- or second-degree female relatives having lived beyond age 45 in either lineage.”\textsuperscript{6}

**Comprehensive genetic testing**

The National Comprehensive Cancer Network (2017)\textsuperscript{6} guidelines state that: “Unless the affected individual is a member of an ethnic group for which particular founder gene mutations are known, comprehensive genetic testing (i.e., full sequencing of the genes and detection of large gene rearrangements) should be performed.”

**National Society of Genetic Counselors**

The National Society of Genetic Counselors (2013)\textsuperscript{8} guidelines also state that: “[For patients with negative sequencing results], it may be appropriate to request additional analysis to detect large genomic rearrangements in both BRCA1 and BRCA2 genes.”

In non-Ashkenazi Jewish individuals: If no mutation or inconclusive results are reported after sequence analysis, testing for large deletions/duplications in BRCA1/2 should be considered. \textsuperscript{1,7,8}

**Stadler et al**

Frequency of gene rearrangements is reviewed in a 2010 study by Stadler et al:\textsuperscript{10}

“Genomic rearrangements in the BRCA1 gene are found in 1.3-5.1\% of families with histories highly suggestive of an inherited predisposition, accounting for 8-19\% of all BRCA1 mutations.”

“The prevalence of genomic rearrangements in the BRCA2 gene appears to be lower, with such alterations accounting for 0-11\% of all BRCA2 mutations.”

In their series of 108 patients with a qualifying history and negative results from BRCA1/2 sequencing, none had mutations found by rearrangement testing. The
authors conclude: “Major gene rearrangements involving the BRCA1/2 genes appear to contribute little to the burden of inherited predisposition to breast and ovarian cancer in the Ashkenazim.”

Jackson et al

Jackson et al 2014\(^\text{11}\) addresses the characteristics of individuals who are more likely to have a large rearrangements in BRCA1/2:

- Latin American/Caribbean ancestry
- number of first-degree relatives with breast cancer is 1 or more
- younger age at first breast cancer diagnosis, less than average age of 39.8 years
- more likely to have ER- and PR- breast cancers
- more likely to have more two breast cancers as well as ovarian cancer, or
- more likely to have infiltrating ductal carcinoma with ductal carcinoma in situ features.

U.S. Preventive Services Task Force

The U.S. Preventive Services Task Force (USPSTF, 2013) recommendations address women who do not have a personal history of breast and/or ovarian cancer, but rather have a family history of these cancer types.\(^\text{12}\) The USPSTF guideline recommends:

- “That primary care providers screen women who have family members with breast, ovarian, tubal (fallopian tube), or peritoneal cancer with one of several screening tools designed to identify a family history that may be associated with an increased risk for potentially harmful mutations in breast cancer susceptibility genes (BRCA1/2). Women with positive screening results should receive genetic counseling and, if indicated after counseling, BRCA testing.”

Grade B recommendation

The USPSTF considers this a Grade B recommendation: “The USPSTF found at least fair evidence that [the service] improves important health outcomes and concludes that benefits outweigh harms.”

Recommendations for genetic counseling

The USPSTF guidelines no longer make explicit recommendations as to who should have BRCA1/2 gene testing; they only make recommendations for genetic counseling. In general, women identified as high risk by these screening tools have one or more of the following characteristics:\(^\text{13}\)

- a first or second degree relative with breast cancer before 50 years old
- a first or second degree relative with ovarian cancer
- a first or second degree relative with bilateral/multifocal breast cancer
- a first or second degree male relative with breast cancer
- a first or second degree relative with both breast and ovarian cancers
- two or more relatives, first, second, third degree, with breast or ovarian cancer
- two or more relatives, first, second, third degree, with breast or prostate/pancreatic cancer, or
- presence of Ashkenazi Jewish ancestry with any of the above.

Criteria

Introduction

Requests for BRCA analysis are reviewed using these criteria.

Scope

Note  This guideline does not address BRCA analysis for individuals of Ashkenazi Jewish ancestry. For information on this testing, please see the guideline BRCA Ashkenazi Jewish Founder Mutation Testing. This guideline also does not address BRCA Analysis as part of multigene panels. For information on this testing, please see the guideline Hereditary Cancer Syndrome Multigene Panels.

Known Familial Mutation Analysis

- Genetic Counseling:
  - Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy),\(^6,8,12,14\) AND
- Previous Genetic Testing:
  - No previous full sequence testing or deletion/duplication analysis, and
  - Known family mutation in BRCA1/2 identified in 1st, 2nd, or 3rd degree relative(s), AND
- Age 18 years or older\(^15\), AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

Full Sequence Analysis

- Genetic Counseling:
- Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy)\textsuperscript{6,8,12,14}, AND

**Previous Genetic Testing:**
- No previous full sequencing of BRCA1/2, and
- No known mutation identified by previous BRCA analysis, AND

**Age 18 years or older\textsuperscript{15}, AND**

**Diagnostic Testing for Symptomatic Individuals:**\textsuperscript{6}
- Non-Ashkenazi Jewish descent, AND: \textsuperscript{1,6}
- Personal History:
  - Female with breast cancer diagnosis ≤45 years of age, and/or
  - Two breast primary tumors with first diagnosis ≤50 years of age and second diagnosis at any age (ipsilateral or bilateral), and/or
  - Diagnosed ≤60 years of age with estrogen receptor negative, progesterone receptor negative, and HER2 negative (triple negative) breast cancer and/or
  - Diagnosed ≤50 years of age with a limited family history (NCCN provides this guidance regarding limited family history: "individuals with limited family history, such as fewer than two first- or second- degree female relatives having lived beyond 45 in either lineage, may have an underestimated probability of a familial mutation"), and/or
  - Male with breast cancer at any age, and/or
  - Epithelial ovarian, fallopian tube, or primary peritoneal cancer diagnosis at any age, and/or
  - BRCA1/2 mutation detected by tumor profiling in the absence of germline mutation analysis, and/or
  - Metastatic prostate cancer (radiographic evidence of or biopsy-proven disease), OR

- Personal & Family History Combination:\textsuperscript{6}
  - Diagnosed ≤50 years of age with at least one close blood relative with breast cancer diagnosed at any age, and/or
  - Diagnosed ≤50 years of age with at least one close blood relative with pancreatic cancer or prostate cancer (Gleason score at least 7), diagnosed at any age, and/or
  - Initial breast cancer diagnosis at any age and one or more of the following:
    - Breast cancer in at least 1 close blood relative (first-, second-, or third-degree) ≤50 years of age, and/or

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400 Buckwalter Place Boulevard, Bluffton, SC 29910 (800) 918-8924
www.eviCore.com
• Epithelial ovarian, fallopian tube, or primary peritoneal cancer in at least 1 close blood relative (first-, second-, or third- degree) at any age, and/or
• At least 2 close blood relatives (first-, second-, or third- degree on same side of family) with breast cancer, pancreatic cancer, or prostate cancer (Gleason score at least 7) at any age, and/or
• Male close blood relative (first-, second-, or third- degree) with breast cancer, and/or
  ▪ Personal history of pancreatic cancer or prostate cancer (Gleason score at least 7) at any age with ≥1 close blood relatives (on the same side of the family) with ovarian cancer at any age or breast cancer ≤50 years or two close blood relatives with breast and/or pancreatic and/or prostate cancer (Gleason score at least 7) at any age, OR

• Predisposition Testing for Presymptomatic/Asymptomatic Individuals
  o Non-Ashkenazi Jewish descent, and one or more of the following: 1,6
    ▪ A first or second degree relative with breast cancer at age 45 or younger, or
    ▪ A first or second degree relative with two primary breast cancers, with the first diagnosis occurring at age 50 or younger, or
    ▪ A first or second degree relative with a triple negative breast cancer (ER-, PR-, Her2-) occurring at age 60 or younger, or
    ▪ A first or second degree relative with ovarian/fallopian tube/primary peritoneal cancer at any age, or
    ▪ A first or second degree relative with male breast cancer at any age, or
    ▪ A combination of two or more first or second degree relatives on the same side of the family with breast cancer, one of whom was diagnosed at age 50 or younger, or
    ▪ A combination of three or more first or second degree relatives on the same side of the family with breast cancer regardless of age at diagnosis, or
    ▪ A combination of both breast and ovarian/fallopian tube/primary peritoneal cancer among two or more first or second degree relatives on the same side of the family, or
    ▪ A first or second degree relative with both breast and ovarian/fallopian tube/primary peritoneal cancer at any age, or
    ▪ A combination of three or more first or second degree relatives on the same side of the family with breast or ovarian/fallopian tube/primary peritoneal cancer AND pancreatic or prostate (Gleason score at least 7) cancer at any age, OR
  o Ashkenazi Jewish individual who is negative for founder mutation testing, and has a high pre-test probability of carrying a BRCA mutation 1,6,8 AND
• Unaffected member is the most informative person to test. All affected family members are deceased, or all affected family members have been contacted and are unwilling to be tested, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**First-degree relatives (parents, siblings, children); second-degree relatives (aunts, uncles, grandparents, grandchildren, nieces, nephews and half-siblings); and third-degree relatives (great-grandparents, great-aunts, great-uncles, and first cousins) on the same side of the family.

**Billing and reimbursement considerations**

  o These criteria may only be applied to a single BRCA sequencing CPT code as defined in the table at the beginning of this guideline.

  o If BRCA gene testing will be performed as part of an expanded hereditary cancer syndrome panel, please also see that guideline for guidance.

**Deletion/Duplication Analysis**

• Genetic Counseling:

  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:

  o No previous BRCA deletion/duplication analysis, and

  o Meets criteria for full sequence analysis of BRCA1/2, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Billing and reimbursement considerations**

If BRCA1/2 deletion/duplication analysis will be performed concurrently with BRCA1/2 gene sequencing, CPT code 81162 is likely most appropriate.

If BRCA gene testing will be performed as part of an expanded hereditary cancer syndrome panel, please also see that guideline for guidance.

**References**

**Introduction**

These references are cited in this guideline.


3. NCI Fact Sheet for BRCA1 and BRCA2: Cancer Risk and Genetic Testing (Reviewed 04/01/2015) Available at: http://www.cancer.gov/about-cancer/causes-prevention/genetics/brcapfact-sheet#r1


Introduction

BRCA Ashkenazi Jewish founder mutation testing is addressed by this guideline.

Procedures addressed

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<td>BRCA 1 and BRCA 2 Ashkenazi Jewish Founder Mutations</td>
<td>81212</td>
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</table>

What is hereditary breast and ovarian cancer

Definition

Hereditary breast and ovarian cancer (HBOC) is an inherited form of cancer.

Characteristics of HBOC

HBOC is characterized by any of the following:\(^1\)\(^2\)

- personal history of
  - breast cancer at a young age, typically under age 50
  - two primary breast cancers
  - both breast and ovarian cancer
  - triple negative breast cancer (ER-, PR-, HER2-)
  - ovarian, fallopian tube, or primary peritoneal cancer, or
  - metastatic prostate cancer
- multiple cases of breast or ovarian cancer in a family
- personal or family history of
  - male breast cancer
  - pancreatic cancer with breast or ovarian cancer, or
prostate cancer with a Gleason score of at least 7 and a family history of ovarian, breast, prostate, or pancreatic cancer

- previously identified BRCA1 or BRCA2 mutation in the family, or
- any of the above with Ashkenazi Jewish ancestry.

### Inheritance

Up to 10% of all breast cancer and 15% of all ovarian cancer is associated with an inherited gene mutation, with BRCA1 and BRCA2 accounting for about 20-25% of all hereditary cases.\(^1,3\)

BRCA mutations are inherited in an autosomal dominant manner. When a parent has a BRCA mutation, each offspring have a 50% risk of inheriting the mutation.\(^1\)

### Prevalence

About 1 in 400 people in the general population has a BRCA1 or BRCA2 mutation. The prevalence of mutations is higher in people of Norwegian, Dutch, or Icelandic ethnicity.\(^1,3\)

The prevalence of BRCA mutations varies among African Americans, Hispanics, Asian Americans, and non-Hispanic whites.\(^3\)

**Ashkenazi Jewish ancestry**

About 1 in 40 people of Ashkenazi Jewish ancestry has a BRCA1 or BRCA2 mutation. The majority of the risk in the Ashkenazi Jewish population is associated with three common founder mutations, two of which are in the BRCA1 gene and one in the BRCA2 gene.\(^1,6,7\) These three mutations account for 99% of identified mutations in the Ashkenazi Jewish population.\(^1\)

### Cancer risks

People with a BRCA mutation have an increased risk of various types of cancer.\(^1\)

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</tr>
</tbody>
</table>

**Note** The risk for breast and ovarian cancer varies among family members and between families.
Screening and prevention

Screening and prevention options are available to specifically address the increased risk of these cancers in a person with a BRCA mutation.\(^1\)

Breast cancer risk and other genes

Other inherited cancer syndromes that can include breast cancer are Li-Fraumeni syndrome (TP53 gene), Cowden syndrome (PTEN), Hereditary Diffuse Gastric Cancer syndrome (CDH1), and Peutz Jeghers syndrome (STK11). Additionally, other genes that can increase the risk for breast cancer are ATM, CHEK2, and PALB2.\(^1,6,8,9\)

Test information

Introduction

BRCA testing may include Ashkenazi Jewish founder mutation testing, full gene sequencing, deletion/duplication analysis, known familial mutation analysis, or multigene panel testing.

Ashkenazi Jewish founder mutation testing

This test is appropriate for those who meet criteria and have Ashkenazi Jewish ancestry.\(^6-8\)

Ashkenazi Jewish founder mutation testing includes the three mutations most commonly found in the Ashkenazi Jewish population:

- \(187\text{delAG}\) and \(5385\text{insC}\) in BRCA1, and
- \(6174\text{delT}\) in BRCA2.\(^1\)

Testing for these three most common mutations detects about 98% of mutations in those with Ashkenazi Jewish ancestry.\(^1,6\)

Other testing options

See the BRCA Analysis guideline for other testing options:

- full sequence testing
- deletion/duplication analysis, or
Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to BRCA Ashkenazi Jewish founder mutation testing.

National Comprehensive Cancer Network

The National Comprehensive Cancer Network (NCCN, 2017)\textsuperscript{7} evidence and consensus-based guidelines include unaffected women with a family history of cancer, those with a known mutation in the family, those with a personal history of breast cancer and/or ovarian cancer, those with a personal history of pancreatic and/or prostate (Gleason score at least 7) cancer, and men with breast cancer.

Based on these guidelines, and the recommendations of the National Society of Genetic Counselors (2013)\textsuperscript{8} the founder mutation analysis is appropriate for any individual with Ashkenazi Jewish ancestry with a personal history of breast, epithelial ovarian, fallopian tube, primary peritoneal, or pancreatic cancer. When there is a personal history of prostate cancer (Gleason score at least 7), additional family history of hereditary breast ovarian cancer syndrome related cancers is required.

These recommendations are Category 2A, defined as "lower-level evidence with uniform NCCN consensus."

U.S. Preventive Services Task Force

The U.S. Preventive Services Task Force (USPSTF, 2013) recommendations address women who do not have a personal history of breast and/or ovarian cancer, but rather have a family history of these cancer types.\textsuperscript{10, 11}

The USPSTF guideline recommends that primary care providers identify women who have a family history of breast, ovarian, fallopian tube, or peritoneal cancer with one of several screening tools. These tools are designed to identify women who may be at an increased risk to carry a BRCA mutation. Women identified as high risk should then be referred for genetic counseling and, if indicated after counseling, BRCA testing.

Women identified as high risk by these screening tools typically have one or more of the following characteristics:\textsuperscript{10-11}

- a first or second degree relative with
  - breast cancer before 50 years old
  - ovarian cancer
  - bilateral/multifocal breast cancer, or

- known familial mutation.
• both breast and ovarian cancers
  • a first or second degree male relative with breast cancer
  • two or more relatives (first, second, third degree) with
    o breast or ovarian cancer or both
    o breast, prostate, or pancreatic cancer, or
  • presence of Ashkenazi Jewish ancestry with any of the above

**Grade B recommendation**

The USPSTF considers this a Grade B recommendation: “The USPSTF found at least fair evidence that [the service] improves important health outcomes and concludes that benefits outweigh harms.”

**Criteria**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy),7,8,10-12 AND
- Previous Genetic Testing:
  - No previous full sequence testing, and
  - No previous deletion/duplication analysis, and
  - No previous Ashkenazi Jewish founder mutation testing, AND
- Age 18 years or older13, AND
- Diagnostic Testing for Symptomatic Individuals:7
  - Ashkenazi Jewish descent, and
    - Epithelial ovarian, fallopian tube, or primary peritoneal cancer diagnosis at any age, and/or
    - Male or female breast cancer diagnosis at any age, or
    - Personal history of pancreatic cancer, or
    - Personal history of prostate cancer (Gleason score at least 7) at any age with at least one close blood relative** with breast cancer (at 50 or younger) and/or ovarian at any age and/or two relatives with breast, pancreatic or prostate cancer (Gleason score at least 7) at any age, or
    - Personal history of metastatic prostate cancer (radiographic evidence of or biopsy-proven disease), OR
- Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
Ashkenazi Jewish descent and a first or second degree relative meeting the following:

- Epithelial ovarian, fallopian tube, or primary peritoneal cancer diagnosis at any age, and/or
- Male or female breast cancer diagnosis at any age, or
- Personal history of pancreatic cancer, or
- Personal history of prostate cancer (Gleason score at least 7) at any age with at least one close blood relative** with breast cancer (at 50 or younger) and/or ovarian at any age and/or two relatives with breast, pancreatic or prostate cancer (Gleason score at least 7) at any age, and
- The affected relative is deceased, unable, or unwilling to be tested†, or
- Close blood relative (1st, 2nd, or 3rd degree) with a known founder mutation in a BRCA1/2 gene, AND

Rendering laboratory is a qualified provider of service per the Health Plan policy.

**First-degree relatives (parents, siblings, children); second-degree relatives (aunts, uncles, grandparents, grandchildren, nieces, nephews and half-siblings); and third-degree relatives (great-grandparents, great-aunts, great-uncles, and first cousins) on the same side of the family.

†Testing of unaffected individuals should only be considered when an affected family member is unavailable for testing due to the significant limitations in interpreting a negative result.

Note  Full gene sequencing of BRCA1/2 may be indicated if no founder mutations are detected by 81212 and the individual meets the criteria above. See BRCA Analysis guideline for criteria.

References

Introduction

These references are cited in this guideline.


2. Hampel H et al. A practice guideline from the American College of Medical Genetics and Genomics and the National Society of Genetic Counselors: referral indications


Introduction

BRCA sequencing for drug treatment response in ovarian cancer is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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What is BRCA sequencing for drug treatment response in ovarian cancer

Definition

BRCA genes are tumor suppressor genes, which means that they help prevent cells from growing and dividing too rapidly or in an uncontrolled way. They encode proteins involved in repairing damaged DNA, thereby helping to maintain the stability of the genetic information contained in the cells. If both copies of one, or both, of these genes are mutated, the DNA repair process does not occur properly and the damaged DNA can allow cells to grow and divide uncontrollably, leading to the development of a tumor.1
Ovarian cancer risk

A female who has an inherited (germline) mutation in a BRCA gene has approximately an 8.46 to 30-fold increased risk, which is 11-39%, to develop ovarian cancer before the age of 70 years. The baseline risk for ovarian cancer in the general population is 1.3%.²

Prevalence

The National Cancer Institute estimates approximately 22,280 women will be diagnosed with ovarian cancer and 14,280 are expected to die from this disease in 2016.³

About 1 in 400 people in the general population has a BRCA mutation.⁴ Approximately 15% of all ovarian cancer is associated with hereditary BRCA mutations.²

Survival

The 5-year survival rate for ovarian cancer is 46.2%.³

PARP

Poly (ADP-ribose) polymerase (PARP) is a family of proteins involved in a number of cellular processes including DNA repair and programmed cell death.

In the case of DNA repair, a PARP inhibitor blocks the PARP from performing its normal function of repairing damaged single-stranded DNA. If the damaged single-stranded DNA persists through cell replication and cell division, double stranded breaks in the DNA can occur.⁵

Available drugs

Olaparib (Lynparza™) and rucaparib (Rubraca™) are oral PARP inhibitors manufactured by AstraZeneca and Clovis Oncology, respectively. These drugs are thought to act by prohibiting PARP from starting the DNA repair process.⁵,⁶,⁷

When PARP is inhibited in individuals who have BRCA mutations, the proteins encoded for by these genes will not be able to repair the double-stranded breaks, so the cell will be overcome with damaged DNA and die. The cells that do not contain a BRCA mutation will still be able to effectively repair any DNA damage using other DNA repair methods and persevere even when PARP is inhibited.⁵,⁶,⁷

Test information

Introduction

Two companion diagnostics are available to assist with drug selection for patients who have been previously treated with chemotherapy for ovarian cancer.
BRACAnalysis

BRACAnalysis CDx™, developed by Myriad Genetic Laboratories was approved by the FDA in 2014 as an in vitro diagnostic device: “BRACAnalysis CDx is an in vitro diagnostic device intended for the qualitative detection and classification of variants in the protein coding regions and intron/exon boundaries of the BRCA1 and BRCA2 genes using genomic DNA obtained from whole blood specimens collected in EDTA. Single nucleotide variants and small insertions and deletions (indels) are identified by polymerase chain reaction (PCR) and Sanger sequencing. Large deletions and duplications in BRCA1 and BRCA2 are detected using multiplex PCR. Results of the test are used as an aid in identifying ovarian cancer patients with deleterious or suspected deleterious germline BRCA variants eligible for treatment with Lynparza (olaparib).”

FoundationFocus

FoundationFocus™ CDx<sub>BRCA</sub>, developed by Foundation Medicine, was approved by the FDA in 2016 as an in vitro diagnostic device: “The FoundationFocus CDx BRCA is a next generation sequencing based in vitro diagnostic device for qualitative detection of BRCA1 and BRCA2 alterations in formalin-fixed paraffin-embedded (FFPE) ovarian tumor tissue. The FoundationFocus CDx<sub>BRCA</sub> assay detects sequence alterations in BRCA1 and BRCA2 (BRCA1/2) genes. Results of the assay are used as an aid in identifying ovarian cancer patients for whom treatment with Rubraca (rucaparib) is being considered. If a patient is positive for any of the deleterious alterations specified in the BRCA1/2 classification, the patient may be eligible for treatment with Rubraca.”

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to BRCA sequencing for drug treatment response in ovarian cancer.

U.S. Food and Drug Administration (FDA)

The FDA approved olaparib (Lynparza) in December 2014 and rucaparib (Rubraca) in December 2016 when prescribed based on results from an FDA-approved companion diagnostic.9,10

- “Lynparza is a poly (ADP-ribose) polymerase (PARP) inhibitor indicated as monotherapy in patients with deleterious or suspected deleterious germline BRCA mutated (as detected by an FDA-approved test) advanced ovarian cancer who have been treated with three or more prior lines of chemotherapy.”9
• “Rubraca is a poly (ADP-ribose) polymerase (PARP) inhibitor indicated as monotherapy for the treatment of patients with deleterious BRCA mutation (germline and/or somatic) associated advanced ovarian cancer who have been treated with two or more chemotherapies. Select patients for therapy based on an FDA-approved companion diagnostic…”10

National Comprehensive Cancer Network

The National Comprehensive Cancer Network (NCCN, 2017) guidelines for ovarian cancer provide the following direction in regards to treatment selection:

• Olaparib: “For patients with deleterious germline BRCA-mutated (as detected by an FDA-approved test or other validated test performed in a CLIA-approved facility) advanced ovarian cancer who have been treated with three or more lines of chemotherapy.”11

• Rucaparib: “For patients with deleterious germline and/or somatic BRCA mutated (as detected by an FDA-approved test or other validated test performed in a CLIA-approved facility) advanced ovarian cancer who have been treated with two or more lines of chemotherapy.”11

Criteria

FoundationFocus CDxBRCA testing may be considered in individuals who meet ALL of the following criteria:

• Full gene sequencing of BRCA1 and BRCA2 has not been previously performed, AND
• Member does not have a known BRCA mutation in the family,** AND
• Member has advanced ovarian cancer, AND
• Treatment with rucaparib is being considered, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy

BRACAnalysis CDx testing may be considered in individuals who meet ALL of the following criteria:

• Full gene sequencing of BRCA1 and BRCA2 has not been previously performed, AND
• Member does not have a known BRCA mutation in the family,** AND
• Member has advanced ovarian cancer, AND
• Treatment with olaparib is being considered, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Please see guideline named BRCA Analysis for testing algorithm in cases where there is a known BRCA1/2 mutation in the family.
References

Introduction

These references are cited in this guideline.


10. Rubraca (rucaparib) prescribing information. Available at: http://www.accessdata.fda.gov/drugsatfda_docs/label/2016/209115s000lbl.pdf

Breast Cancer Index for Breast Cancer Prognosis

Introduction

Breast Cancer Index for breast cancer prognosis is addressed by this guideline.

Procedures addressed

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What is Breast Cancer Index for breast cancer prognosis

Definition

Breast Cancer Index® (BCI) is a commercial multigene expression profiling assay designed to assess prognosis in early-stage breast cancer patients.¹

Breast cancer recurrence

A large percentage of breast cancer patients (ER+/LN-) treated with endocrine therapy alone are free of disease 10+ years after initial diagnosis, and could forgo chemotherapy and its toxic side effects. Furthermore, a meta-analysis (n=~35,000 patients) reported a rate of recurrence of ~2% per year for breast cancer patients (ER+/LN-) receiving only tamoxifen.² Consequently, accurate prediction of the risk of breast cancer recurrence is important for establishing the most optimal course of treatment with endocrine therapy, adjuvant chemotherapy, or both for women with early-stage breast cancer.

Risk assessment

Conventional methods of risk assessment including using the following clinicopathologic factors

- tumor size
- involvement of regional lymph nodes
- histologic grade
- expression of hormone receptors (estrogen and progesterone), and
• human epidermal growth factor receptor 2 (HER2) amplification.

These may not be sufficiently accurate to identify those subgroups of patients who are at low risk of recurrence and who are unlikely to benefit from extended endocrine therapy or adjuvant chemotherapy.\(^5\)

As a result, alternative biomarker prognostic tests have been developed to more accurately predict individual risk of cancer recurrence and to better inform clinicians making treatment decisions for patients with early-stage breast cancer, including

• determining appropriate chemotherapy regimens
• decreasing treatment-associated complications, and
• avoiding unnecessary treatment.\(^4\)

**Intended use**

According to the manufacturer, "The Breast Cancer Index (BCI) Risk of Recurrence & Extended Endocrine Benefit Test is intended for use in patients diagnosed with estrogen receptor-positive (ER+), lymph node-negative (LN-) or lymph node positive (LN+; with 1-3 positive nodes) early-stage, invasive breast cancer, who are distant recurrence-free. BCI provides:

• A quantitative assessment of the likelihood of both late (post-5 years) and overall (0-10 year) distant recurrence following an initial 5 years of endocrine therapy (LN- patients) or 5 years of endocrine therapy plus adjuvant chemotherapy (LN+ patients), and
• Prediction of likelihood of benefit from extended (>5 year) endocrine therapy. BCI results are adjunctive to the ordering physician’s workup; treatment decisions require correlation with all other clinical findings."\(^1\)

**Test information**

**Introduction**

The test is intended to provide risk information beyond standard predictive and prognostic factors and identify those patients unlikely to benefit from extended endocrine therapy or adjuvant chemotherapy.\(^1\)

**Breast Cancer Index**

The Breast Cancer Index assay is an algorithmic gene expression-based signature, which combines 2 independent biomarkers (HOXB13:IL17BR [H:I or H/I] and the 5-gene molecular grade index (MGI) to evaluate estrogen-mediated signaling and tumor grade.\(^2\)
As a risk stratification tool, BCI attempts to stratify patients with early-stage estrogen-receptor positive (ER+), lymph-node negative (LN-) patients into three different risk groups, as well offer a continuous evaluation of an individual patient’s risk of distant recurrence.²

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to Breast Cancer Index testing.

**American Society of Clinical Oncology**

The American Society of Clinical Oncology (ASCO, 2016) published a clinical practice guideline regarding the use of biomarkers to guide clinical decision-making on adjuvant systemic therapy among women with early-stage invasive breast cancer. Based on a review of the peer-reviewed scientific evidence, the following recommendations were published:⁵

- “If a patient has ER/PgR-positive, HER2-negative (node-negative) breast cancer, the clinician may use the Breast Cancer Index to guide decisions on adjuvant systemic therapy. Type: evidence based. Evidence quality: intermediate. Strength of recommendation: moderate.”
- “If a patient has ER/PgR-positive, HER2-negative (node-positive) breast cancer, the clinician should not use the Breast Cancer Index to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: insufficient. Strength of recommendation: strong.”
- “If a patient has HER2-positive breast cancer or TN breast cancer, the clinician should not use the Breast Cancer Index to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: insufficient. Strength of recommendation: strong.”

**Relevant studies**

Several retrospective and prospective-retrospective studies, published by the manufacturer, have assessed the clinical validity of the BCI test for women with early stage breast cancer (ER+/LN-) to guide clinical decision making regarding adjuvant therapy (prognostic) or regarding treatment response (predictive).²⁶⁻⁹

**Clinical validity**

Results of clinical validity are generally consistent across these studies, reporting that women classified by the BCI test into higher risk categories tend to have worse rates of distant recurrence, and women in lower risk categories have better rates of distant recurrence. However, studies are hampered by several limitations which
weaken the overall quality of the evidence base, including small patient populations, lack of generalizability to broader real-world patient populations in clinical practice, retrospective study designs, lack of blinding, and heterogeneity in patient clinical characteristics across comparator groups. In addition, studies regarding clinical validity alone are insufficient to provide support for a recommendation that the BCI test can be used to guide clinician decisions in early-stage breast cancer, with the ability to augment or replace traditional prognostic and predictive factors currently used in clinical practice.

Clinical utility

The evidence is insufficient to determine if the BCI biomarker test demonstrates clinical utility with a reasonable balance of benefits and harms compared with treatment under standard assessments without the test. Currently available studies do not widely report clinically meaningful benefits measured by patient-relevant outcomes, including overall survival (OS), disease-free survival (DFS), or progression-free survival (PFS), beyond that being provided by conventional clinicopathologic factors.

Additional studies

Additional studies definitively demonstrating clinical utility in well-designed clinical trials with proper patient-relevant outcomes are necessary to confirm that BCI may help clinicians to make better informed treatment decisions and spare toxic chemotherapy for a large group of early-stage breast cancer patients.

Criteria

Introduction

Requests for Breast Cancer Index testing are reviewed using these criteria.

Criteria

• Previous Testing:
  o No repeat Breast Cancer Index testing on the same sample when a result was successfully obtained, and
  o No previous gene expression assay (e.g. OncotypeDx Breast) performed on the same sample when a result was successfully obtained, AND

• Required Clinical Characteristics:
  o Primary invasive breast cancer meeting all of the following criteria:
    o Unilateral tumor
      ▪ Tumor size >0.5cm (5mm) in greatest dimension (T1b-T3), and
• Hormone receptor positive (ER+ or PR+), and
• HER2 negative, and
  o Patient has no regional lymph node metastasis (pN0) or only micrometastases (pN1mi, malignant cells in regional lymph node(s) not greater than 2.0mm), and
  o Adjuvant endocrine systemic chemotherapy is a planned treatment option for the patient or results from this Breast Cancer Index test will be used in making adjuvant chemotherapy treatment decisions, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Other Considerations

Testing Multiple Samples:

• When more than one ipsilateral breast cancer primary is diagnosed, testing should be performed on the tumor with the most aggressive histologic characteristics. If an exception is requested, the following criteria will apply:
  o There should be reasonable evidence that the tumors are distinct (e.g., different quadrants, different histopathologic features, etc.), AND
  o There should be no evidence from either tumor that chemotherapy is indicated with or without knowledge of the Breast Cancer Index test result (e.g., histopathologic features or previous Breast Cancer Index result of one tumor suggest chemotherapy is indicated), AND
  o If both tumors are to be tested, both tumors must independently meet the required clinical characteristics

References

Introduction

These references are cited in this guideline.

1. Biothernostics. The Breast Cancer Index (BCI). Available at: https://www.breastcancerindex.com/


Introduction

Brugada syndrome genetic testing is addressed by this guideline.

Procedures addressed

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What is Brugada syndrome

Definition

Brugada syndrome (BrS) is an inherited channelopathy characterized by right precordial ST elevation. This can result in cardiac conduction delays at different levels, syncope, or a lethal arrhythmia resulting in sudden cardiac death.

Onset

Although the typical presentation of BrS is sudden death in a male in his 40s with a previous history of syncope, BrS has been seen in individuals between the ages of 2 days and 85 years,¹ as well as females.²

Diagnosis

The diagnosis of BrS is based on ECG results, clinical presentation and family history. A diagnosis of either type 1, 2, or 3 ECG results with a personal history of fainting spells, ventricular fibrillation, self-terminating polymorphic ventricular tachycardia, or
electrophysiologic inducibility can help identify those at risk for BrS. A family history of syncope, coved-type ECGs, or sudden cardiac death, especially in an autosomal dominant inheritance pattern, can help aid in the diagnosis.\textsuperscript{3,4}

\textbf{Cause}

BrS has been associated with at least 16 different genes and >400 mutations\textsuperscript{3,5-7}, and is estimated to be seen in about 1 in 2000 individuals. Approximately 65-75\% of families with a clinical diagnosis of BrS do not test positive for a mutation in one of the known genes, suggesting that there are other genes that have not been identified.\textsuperscript{3,5}

- SCN5A is responsible for the majority of BrS cases (15-30\%).
- There are reports that CACNA1C and CACNB2B may account for up to 11\% of cases of BrS.\textsuperscript{6,8}
- Each of the other genes comprise less than 5\% of mutations in each case.

BrS has variable expression and incomplete penetrance. Approximately 25\% of gene positive individuals have an ECG diagnostic of BrS.\textsuperscript{3,5} Additionally, 80\% individuals with a disease-causing mutation only present with symptoms when challenged with a sodium channel blocker.\textsuperscript{2,9}

\textbf{Prevalence}

BrS is found worldwide, but seems to have a higher incidence in Southeast Asia. In countries such as Japan, the Philippines, Laos, and Thailand, a condition called Sudden Unexplained Nocturnal Death syndrome (SUNDS) has been associated with mutations in the SCN5A, suggesting that this condition is actually Brugada Syndrome.\textsuperscript{10,11} In these countries, SUNDS is the second most common cause of death of men under age 40 years.\textsuperscript{3}

\textbf{Inheritance}

BrS is inherited in an autosomal dominant inheritance pattern, with the exception of KCNE5-related Brugada syndrome, which is inherited in an X-linked manner.\textsuperscript{3} This means that an individual has a 50\% chance of passing on a mutation to their children. Additionally, parents and siblings of known carriers have a 50\% chance of being carriers of the same mutation.

When a mutation in a child is not found in the parents, it is assumed that there is a de novo mutation in the child. De novo mutations are estimated to occur in approximately 1\% of cases.\textsuperscript{3} Siblings would still need to be tested to rule out germline mutations.

A DNA test for BrS should be offered to the person who has the most obvious disease, as that individual will more likely test positive than someone without disease. At this time, population wide carrier screening for BrS is not recommended.\textsuperscript{5}
**Test information**

**Introduction**

Testing for Brugada syndrome may include full sequence analysis, deletion/duplication testing, known familial mutation analysis, or multigene panels.

**Full sequence analysis**

Full sequence analysis of the SCN5A gene is available through a number of commercial laboratories.

**Deletion/duplication testing**

Deletion/duplication testing for SCN5A is available and is typically done in reflex to a negative result from full sequence analysis.

**Known familial mutation analysis**

Known familial mutation analysis can be considered for individuals with a known mutation in the family. Once a deleterious mutation is identified in a family member, at-risk relatives can be tested for only that specific mutation. Testing by single site analysis is greater than 99% accurate.³

**Multigene panels**

Multigene panels can be considered but this test is typically not recommended.

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to Brugada syndrome testing.

**Heart Rhythm Society, European Heart Rhythm Association, and Asia Pacific Heart Rhythm Society**

A 2013 expert consensus statement from the Heart Rhythm Society (HRS), the European Heart Rhythm Association (EHRA), and the Asia Pacific Heart Rhythm Society is silent on the role of genetic testing in diagnosis and management.¹²

**Heart Rhythm Society and European Heart Rhythm Association**

A 2011 expert consensus statement from the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA) recommends.⁵
• “Comprehensive or BrS1 (SCN5A) targeted BrS genetic testing can be useful for any patient in whom a cardiologist has established a clinical index of suspicion for BrS based on examination of the patient's clinical history, family history, and expressed electrophysiologic (resting 12-lead ECGs and/or provocative drug challenge testing) phenotype.” (Class IIa)

• “Genetic testing is not indicated in the setting of an isolated type 2 or type 3 Brugada ECG pattern.”

• “Mutation-specific genetic testing is recommended for family members and appropriate relatives following the identification of the BrS-causative mutation in an index case.”

Multigene panels

The clinical utility of Brugada multigene panels has not been well established. Mutations in SCN5A are responsible for 15-30% of cases of Brugada Syndrome, making it the most common known genetic cause of BrS. There are other genes associated with BrS, but mutations in each gene account for <5% of cases of BrS, therefore incremental mutation yield on a multi-gene panel is expected to be very low.\(^5\)

Criteria

Introduction

Requests for Brugada syndrome testing are reviewed using these criteria.

Brugada Syndrome Known Familial Mutation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous genetic testing for Brugada Syndrome, AND

• Diagnostic and Predisposition Testing:
  o Brugada Syndrome familial mutation identified in biologic relative(s), OR

• Prenatal Testing:
  o Brugada syndrome mutation identified in one biologic parent or 1\(^{st}\) degree relative, AND
• Rendering laboratory is a qualified provider of service per the Health plan policy.

Brugada Syndrome Full Sequence Analysis of SCN5A

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous genetic testing for Brugada Syndrome, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Type 1, 2, or 3 ECG results, and
  o Documented ventricular fibrillation, or
  o Self-terminating polymorphic ventricular tachycardia, or
  o A family history of sudden cardiac death, or
  o Coved-type ECGs in family members, or
  o Electrophysiologic inducibility, or
  o Syncope, or
  o Nocturnal agonal respiration (breaths that persist after cessation of heartbeat), OR

• Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  o Biologic relative(s) (1\textsuperscript{st}, 2\textsuperscript{nd}, or 3\textsuperscript{rd} degree) diagnosed with BrS clinically, and no familial mutation identified, or
  o Sudden death in biologic relative(1\textsuperscript{st}, 2\textsuperscript{nd}, or 3\textsuperscript{rd} degree), and
  o Type 1 ECG changes, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Brugada Deletion/Duplication Analysis of SCN5A

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:(a)
  o No mutation identified with Brugada Syndrome sequence analysis of SCN5A, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Brugada Syndrome Multigene Panels

• Brugada syndrome multigene panels are considered investigational and/or experimental.
  
  o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.

  o In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References

Introduction

These references are cited in this guideline.


5. Ackerman MJ, Priori SG, Willems S, et al. HRS/EHRA expert consensus statement on the state of genetic testing for the channelopathies and cardiomyopathies: this document was developed as a partnership between the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA). Europace. 2011;13(8):1077-1109.


CADASIL Testing

Introduction

CADASIL testing is addressed by this guideline.

Procedures addressed

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What is CADASIL

Definition

CADASIL (Cerebral Autosomal Dominant Arteriopathy with Subcortical Infarcts and Leukoencephalopathy) is an adult-onset form of cerebrovascular disease. There are no generally accepted clinical diagnostic criteria for CADASIL and symptoms vary among affected individuals.

Signs and symptoms

Typical signs and symptoms include¹ ²

- stroke-like episodes before age 60 years
- cognitive disturbance
- psychiatric or behavioral abnormalities
- migraine with aura, and
- recurrent seizures with onset in middle age.

Diagnosis

Brain Magnetic Resonance Imaging (MRI) findings include T2-signal-abnormalities in the white matter of the temporal pole and T2-signal-abnormalities in the external capsule.¹ ²
CADASIL is suspected in an individual with the clinical signs and MRI findings. A positive family history for stroke or dementia is also indicative of disease in symptomatic individuals. However, a negative family history should not exclude the diagnosis, as de novo mutations may occur, and affected family members are frequently misdiagnosed.1,3

For a firm diagnosis of CADASIL, at least one of the following is required:

- Documentation of characteristic GOM deposits within small blood vessels by skin biopsy.1-3
- Documentation of a typical NOTCH3 mutation by genetic gene sequencing.1-3

Prevalence

CADASIL is a rare disease. Cases have been reported worldwide with a prevalence of 1 in 50,000 to 1 in 121,000 individuals, although this may be an underestimate.1-3

CADASIL is thought to be the most common form of hereditary stroke and vascular dementia in adults.

Cause

CADASIL is an autosomal dominant disease caused by mutations in the NOTCH3 gene. Each offspring of an individual with CADASIL has a 50% chance of inheriting the disease-causing mutation.

To date, NOTCH3 is the only gene in which mutations are known to cause CADASIL. NOTCH3 encodes a transmembrane receptor that is primarily expressed in vascular smooth-muscle cells, preferentially in small arteries. Mutations in NOTCH3 generally increase or decrease the number of cysteine residues in the extracellular domain of the protein, which then accumulate in small arteries of affected individuals.1 These accumulations are seen as granular osmophilic material (GOM) deposits in the walls of affected vessels seen on biopsy and are a pathologic hallmark of CADASIL.1

No clear genotype-phenotype correlations exist for individuals with CADASIL and symptoms can vary considerably even within families.3,4

Management and treatment

A correct diagnosis of CADASIL is important because the clinical course of disease is different from individuals with other types of cerebral small-vessel disease and proven therapies for stroke have not been validated in individuals with CADASIL.3 However, no specific treatments for CADASIL exist.1-3 Management and treatment of individuals is generally symptomatic and supportive.1-3
Test information

Introduction

Testing for CADASIL may include a skin biopsy, sequence analysis, or known familial mutation analysis.

Skin biopsy

Specificity of skin biopsy findings is high as the characteristic deposits have not been documented in any other disorder. Specificity has been reported to range from 45%-100%. Sensitivity and specificity can be maximized by to >90% by immunostaining for NOTCH3 protein.

Sequence analysis

NOTCH3 mutation detection may reach >95% in individuals with strong clinical suspicion of CADASIL. To date, all mutations in NOTCH3 causing CADASIL have been in exons 2-24, including intron-exon boundaries, with the majority of pathogenic mutations occurring in exons 2-6. Some laboratories outside of the US offer tiered testing beginning with sequence analysis of select exons followed by sequence analysis of the remaining exons if a mutation is not identified. Other laboratories offer only sequence analysis of the entire coding region. In the United States, a limited number of laboratories offer CADASIL testing and all perform full gene sequencing at the time of this review.

There is evidence of founder mutation in individuals from the islands of Taiwan and Jeju as well as Finland and middle Italy.

Known familial mutation analysis

Once a mutation in an affected individual has been identified, testing at risk individuals in the family is possible.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to CADASIL testing. No evidence-based U.S. testing guidelines have been identified.

Pescini et al (2012)

Pescini et al (2012) published a scale to help guide clinicians in selecting patients for NOTCH3 genetic analysis due to a high probability of a CADASIL genetic diagnosis. This scale assigns weighted scores to common features of CADASIL. The authors state that their scale is “accurate with optimal sensitivity and specificity values (96.7%
and 74.2%, respectively); however, our results need to be confirmed and further validated."\(^7\)

**Choi et al (2010)**

A two-center cohort study found that blood pressure and hemoglobin A1c levels were associated with cerebral mini bleeds in CADASIL patients.\(^3\) Therefore, controlling blood pressure and glucose levels may improve the clinical course of the disease. It is also reasonable to control for high cholesterol and high blood pressure given the high rate of ischemic stroke seen in CADASIL.\(^3\)

**Tikka et al (2009)**

Evidence from one 2009 retrospective cohort study suggests that an adequate skin biopsy for analysis of granular osmophilic material is a cost effective way to determine a diagnosis of CADASIL in symptomatic individuals.\(^5\)

The authors suggest that biopsy results can be used to guide the decision for who should have genetic testing, particularly in individuals with no known familial mutation or from ethnic populations with no evidence of founder mutations.\(^5\)

**Clinical utility**

Patients with CADASIL should avoid anticoagulants, angiography, and smoking to avoid disease-related complications, so clinical utility is represented.\(^1,3\) Because of the risk for intracerebral hemorrhage, use of antiplatelets rather than anticoagulants is considered for prevention of ischemic attacks. Evidence against the use of intravenous tissue plasminogen activator (IV tPA) has been suggested due to the possibility of hemorrhage; however, this is not conclusive.\(^8\) Statins are used for treatment of hypercholesterolemia and antihypertensive drugs are used for hypertension.\(^6\)

**Criteria**

**Introduction**

Requests for CADASIL testing are reviewed using these criteria.

**Known Familial Mutation Testing**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous genetic testing for NOTCH3 mutations, AND
- Predictive or Diagnostic Testing:
Member has a first-degree relative (i.e. parent, sibling, child) with an identified NOTCH3 gene mutation, OR

- Diagnostic Testing:
  - Genetic testing may be performed after member has had a skin biopsy with the following results:
    - Ambiguous or indeterminate results from electron microscopy, and
    - Ambiguous or indeterminate results from immunohistochemistry analysis, AND
  - A high index of suspicion remains for CADASIL diagnosis based on clinical findings, AND
  - Rendering laboratory is a qualified provider of service per the Health Plan policy.

**NOTCH3 Targeted Sequencing**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous genetic testing for NOTCH3 mutations, AND
- Diagnostic Testing:
  - Genetic testing may be performed after member has had a skin biopsy with the following results:
    - Ambiguous or indeterminate results from electron microscopy, and
    - Ambiguous or indeterminate results from immunohistochemistry analysis, AND
  - A high index of suspicion remains for CADASIL diagnosis based on clinical findings, AND
  - Rendering laboratory is a qualified provider of service per the Health Plan policy.

**References**

**Introduction**

These references are cited in this guideline.


Canavan Disease Testing

Introduction

Canavan disease testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
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<tbody>
<tr>
<td>ASPA Known Familial Mutation Analysis</td>
<td>81403</td>
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<tr>
<td>ASPA Targeted Mutation Analysis</td>
<td>81200</td>
</tr>
<tr>
<td>ASPA Sequencing</td>
<td>81479</td>
</tr>
<tr>
<td>ASPA Deletion/Duplication Analysis</td>
<td>81479</td>
</tr>
</tbody>
</table>

What is Canavan disease

Definition

Canavan disease is a genetic disorder leading to progressive damage to the brain’s nerve cells.\(^1\)\(^2\)

Prevalence

Canavan disease is most often found in Ashkenazi Jewish populations.\(^1\)\(^2\)

- About 1 in 40 people of Ashkenazi Jewish descent are carriers for Canavan disease.\(^2\) Because of this relatively high carrier rate, population based screening in the Ashkenazi Jewish population is available. (See Ashkenazi Jewish Carrier Screening).
- Between 1 in 6,400 and 1 in 13,500 Ashkenazi Jews have the disease.\(^1\)

The prevalence among the general population is significantly lower.\(^2\)

Symptoms

Signs and symptoms of Canavan disease usually begin in infancy and include:\(^1\)

- developmental delays including motor skills, learning disabilities, or problems sleeping
• weak muscle tone (hypotonia)
• large head size (macrocephaly)
• abnormal posture, and
• seizures.

Cause

Canavan disease is caused by changes, or mutations, to the ASPA gene. ASPA helps make an enzyme called aspartoacylase. ASPA helps make an enzyme called aspartoacylase. This enzyme is essential to maintain the health of myelin, the nerve cells’ protective covering, by breaking down harmful compounds that would otherwise degrade myelin. The most significant of these compounds that break down myelin is called N-acetylaspartic acid (NAA).

In the absence of aspartoacylase, the myelin protective covering of the nerve is eventually destroyed. Without this protective covering, nerve cells malfunction and die.

Inheritance

Canavan disease is an autosomal recessive disorder, meaning that an affected individual must inherit two ASPA gene mutations - one from each parent.

Individuals with only one mutation are called carriers. Carriers do not show symptoms of Canavan disease, but have a 50% chance of passing on the mutation to their children who will also be carriers.

If two unaffected carriers have children, each of their pregnancies has a 1 in 4 (25%) chance of being affected with Canavan disease.

Diagnosis

Canavan disease is suspected when a patient presents with classic signs and symptoms. Diagnosis is confirmed by biochemical or genetic testing or both. Biochemical tests analyze either NAA levels or aspartoacylase enzyme activity in someone with suspected Canavan disease.

• Affected individuals cannot break down NAA, so it accumulates in the blood or urine.
• Affected individuals will have severely reduced or nonexistent aspartoacylase enzyme activity.

Molecular genetic testing can be used for confirmation of the diagnosis and to help family planning by identifying individuals at risk of being carriers.

Survival

Canavan disease does not usually allow survival beyond childhood.
Test information

Introduction

Testing for Canavan disease may include targeted mutation analysis, sequence analysis, deletion/duplication analysis, or known familial mutation analysis.

Targeted mutation analysis

Targeted mutation analysis is the most common genetic test for Canavan disease. The panel looks for up to four of the most common mutations in the ASPA gene linked to Canavan disease, including the Glu285Ala and Tyr231X mutations, which account for 98% of all Ashkenazi Jewish cases.\(^2,3\) The panel also includes the p.Ala305Glu mutation, which accounts for between 30% and 60% of all non-Ashkenazi Jewish cases.\(^2,3\)

Sequence analysis

Sequence analysis looks for mutations across the entire coding region of the ASPA gene. In addition to the more common mutations found in the Ashkenazi Jewish population, sequencing is also able to find less common mutations found in non-Ashkenazi Jews.\(^2,3\) Sequence analysis has a detection rate of 87% in all populations.\(^2\)

Deletion/duplication analysis

Deletion/duplication analysis will find gene rearrangements that are too large to be detected by sequencing. Large deletions in the ASPA gene have been reported but are believed to be uncommon.\(^2\) Therefore, deletion/duplication analysis is unlikely indicated in most cases.

Known familial mutation analysis

Once mutations have been identified in a symptomatic individual, carrier testing can be performed on at-risk relatives using this same targeted mutation panel or perhaps known familial mutation analysis for the specific mutation identified in the affected individual.

If both members of a couple are carriers with identified mutations, prenatal diagnosis of an at-risk pregnancy is possible using this same targeted mutation panel or known familial mutation analysis for the specific mutations identified in the parents.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to Canavan disease testing.
2010 expert-authored review

A 2010 expert-authored review states the following regarding molecular genetic testing for diagnostic purposes:²

- The targeted mutation panel may be used to confirm a clinical diagnosis, biochemical diagnosis, or both, especially if the patient has Ashkenazi Jewish ancestry.
- “Sequence analysis of the ASPA coding region is available on a clinical basis for individuals in whom mutations were not identified by targeted mutation analysis.”
- “Deletion/duplication analysis. Exonic or whole-gene deletions are rare in individuals with Canavan disease. The authors encountered two individuals with complete deletion of the ASPA gene and two with partial deletions. Deleted segments of various sizes of cDNA have been reported.”

American College of Medical Genetics

The American College of Medical Genetics (ACMG, 2008) supports offering carrier testing for Canavan disease to individuals of Ashkenazi Jewish descent for the two common mutations. It is anticipated that the detection rate will be ~97%. This test should be offered to individuals of reproductive age, preferentially prior to pregnancy, with genetic counseling performed by a geneticist or genetic counselor. ACMG supports the testing of individuals of Ashkenazi Jewish descent, even when their partner is non-Ashkenazi Jewish. In this situation, testing would start with the individual who is Ashkenazi and reflex back to the partner if necessary.⁴

American College of Obstetrics and Gynecologists

The American College of Obstetrics and Gynecologists (ACOG, 2009) recommends that individuals who are considering a pregnancy or are pregnant should consider testing if at least one member of the couple is Ashkenazi Jewish or has a relative with Canavan disease. If the woman is pregnant, testing may need to be conducted on both partners simultaneously in order to receive results in a timely fashion. If one or both partners are found to be carriers of Canavan disease, genetic counseling should be provided, and prenatal testing offered, if appropriate.⁵

Criteria

Introduction

Requests for Canavan Disease testing are reviewed using these criteria.

ASPA Known Familial Mutation Analysis

- Genetic Counseling:
- Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous genetic testing of ASPA, AND

- Carrier Screening for Asymptomatic Individuals:
  - Known family mutation in ASPA in 1st, 2nd, or 3rd degree biologic relative, OR
  - ASPA mutations identified in both biologic parents

**ASPA Targeted Mutation Analysis for Common Mutations**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous ASPA genetic testing, including Ashkenazi Jewish screening panels containing targeted mutation analysis for Canavan disease, AND

- Diagnostic Testing or Carrier Screening:
  - Ashkenazi Jewish descent, regardless of disease status and N-acetylaspartic acid (NAA) levels, OR

- Prenatal Testing for At-Risk Pregnancies:
  - ASPA Ashkenazi mutations identified in both biologic parents.

**ASPA Sequencing**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous ASPA gene sequencing, and
  - No known ASPA mutation in family, and
  - No mutations or one mutation detected by common mutation panel, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Increased levels of N-acetylaspartic acid (NAA) in urine, and
An individual age three to five months of age with a triad of hypotonia, macrocephaly and head lag, or

Failure to attain independent sitting, walking or speech, OR

• Testing for Individuals with Family History or Partners of Carriers:
  
  o 1st, 2nd, or 3rd degree biologic relative with Canavan disease clinical diagnosis, family mutation unknown, and testing unavailable, or
  
  o Partner is monoallelic or biallelic for ASPA mutation, and

  ▪ Have the potential and intention to reproduce

References

Introduction

These references are cited in this guideline.


Introduction

Celiac disease testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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<tr>
<td>HLA Typing</td>
<td>81383</td>
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</tbody>
</table>

What is Celiac disease

Definition

Celiac disease is an immune-mediated disorder that mainly affects the digestive tract.\(^1-4\)

Symptoms

Symptoms include diarrhea, constipation, vomiting, abdominal pain and bloating, growth problems, iron deficiency anemia, osteoporosis, and other complications of malabsorption.\(^1-4\)

Prevalence

Celiac disease affects infants, children, and adults and can present at any age. It affects about 1 in every 100 people in the U.S.\(^2,3\)

Cause

Celiac is caused by exposure to dietary gluten, which is a protein molecule found in wheat, barley and rye, in people who are predisposed based on their genetic makeup.\(^1-4\)
Diagnosis

An initial diagnosis of celiac disease is highly suspected based on serologic testing and is confirmed by finding characteristic changes on intestinal biopsy. Intestinal biopsy remains the gold standard for making a diagnosis of celiac disease.\textsuperscript{1-4}

Increased risk

Patients with certain medical conditions and relatives of people with celiac disease are known to have an increased risk of developing the condition.\textsuperscript{2,3}

Test information

Introduction

Two genetic markers are associated with celiac disease: HLA-DQ2 and HLA-DQ8. These variants are present in about 30-40\% of the general population, but more than 99\% of patients with celiac disease have one or more of these variants\textsuperscript{1}. If a person suspected of having celiac disease is found not to have one of these markers, the diagnosis can be essentially excluded.\textsuperscript{2-4}

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to Celiac disease testing.

Guidelines

Consensus-based guidelines from the American Gastroenterological Association (2006), the National Institutes of Health (2005), North American Society for Pediatric Gastroenterology, Hepatology and Nutrition (2005) and the 2013 American College of Gastroenterology Practice Guidelines state that HLA typing for celiac disease should be used as outlined in this table.\textsuperscript{2-5}

<table>
<thead>
<tr>
<th>Test type</th>
<th>Use</th>
<th>Level of recommendation</th>
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<tbody>
<tr>
<td>Human leukocyte antigen DQ2/DQ8 testing</td>
<td>Do not use routinely in the initial diagnosis of Celiac disease</td>
<td>Strong recommendation, moderate level of evidence</td>
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<tr>
<td>Human leukocyte antigen DQ2/DQ8 genotyping testing</td>
<td>Use to effectively rule out the disease in selected clinical situations</td>
<td></td>
</tr>
</tbody>
</table>
Criteria

Introduction

Requests for Celiac disease testing are reviewed using these criteria.

Criteria

Testing may be considered in individuals who meet the following criterion:

• Celiac disease is in the differential diagnosis, but the individual has had ambiguous or indeterminate results from serology and biopsy.

References

Introduction

These references are cited in this guideline.


Charcot-Marie-Tooth Neuropathy Testing

Introduction

Testing for Charcot-Marie-Tooth disease is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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Procedures addressed by this guideline | Procedure codes
---|---
GARS Sequencing | 81406
LMNA Sequencing | 81406
FIG4 Sequencing | 81406
SH3TC2 Sequencing | 81406
GJB1 Deletion/Duplication Analysis | 81479
Hereditary peripheral neuropathies (eg, Charcot-Marie-Tooth, spastic paraplegia), genomic sequence analysis panel, must include sequencing of at least 5 peripheral neuropathy-related genes (eg, BSCL2, GJB1, MFN2, MPZ, REEP1, SPAST, SPG11, SPTLC1) | 81448

What is Charcot-Marie-Tooth neuropathy

Definition

Charcot-Marie-Tooth neuropathy (CMT) is a group of inherited genetic conditions characterized by chronic motor and sensory polyneuropathy.¹ Onset is typically before 30 years of age.¹ The key finding in CMT is symmetric, slowly progressive distal motor neuropathy in feet, hands, or both. This is expressed as distal muscle weakness or wasting and atrophy with sensory loss, depressed reflexes, and foot deformities including pes cavus and hammer toes.¹

Diagnosis

The diagnosis of CMT is suspected based on clinical and family history, neurological exam, nerve conduction and EMG studies.¹ Acquired causes of neuropathy including alcoholism, vitamin B12 deficiency, thyroid disease, diabetes mellitus, HIV infection, and others, should be ruled out.¹

Molecular genetic testing can be used to establish a specific diagnosis, which aids in understanding the prognosis and risk assessment for family members.¹

Prevalence

CMT is the most common inherited neurological disorder. The prevalence of all CMT types is 1 in 3,300.¹

Types and subtypes

CMT is divided into five types based on EMG findings and mode of inheritance:
• CMT1
• CMT2
• Dominant Intermediate CMT
• CMT4, and
• CMT X.¹

Within each type, there are a number of subtypes, distinguished by causative gene. Mutations in over 40 genes have been linked to CMT.¹ Within a CMT type, specific subtypes are often distinguishable only by genetic testing. For most types, there is one gene that accounts for a large proportion of affected individuals.¹

Identifying CMT subtype in a sporadic patient, who has no known family history, can be difficult due to high new mutation rates.

Inheritance

CMT can be inherited in an autosomal dominant, autosomal recessive, or an X-linked manner.¹

Test information

Introduction

Testing for CMT may include gene sequencing, deletion/duplication analysis, or panel testing.

Genetic testing

There are various methods used to test for mutations in genes which can cause CMT neuropathy

• Single gene analysis
• Deletion/duplication analysis, particularly for the 1.5-Mb duplication at 17p11.2 that includes PMP22
• Panel testing using next generation sequencing

CMT panel testing

CMT multi-gene panels include a wide variety of genes associated with CMT neuropathy. Multi-gene panels may also include genes believed to be associated with CMT neuropathy but with a lower impact on risk than recognized syndromes. Results for such genes are of less clear value because there often are not clear management recommendations for mutation-positive individuals.
Under certain circumstances, technologies used in multi-gene testing may fail to identify mutations that might be identifiable through single-gene testing. If high clinical suspicion remains for a particular syndrome after negative multi-gene test results, consultation with the testing lab and/or additional targeted genetic testing may be warranted.

Multi-gene tests vary in technical specifications (e.g., depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis). Since genes can be easily added or removed from multi-gene tests over time by a given lab, medical records must document which genes were included in the specific multi-gene test used from each patient, and in which labs they were performed.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to CMT testing.

American Academy of Neurology

Evidence-based guidelines from the American Academy of Neurology (2009; reaffirmed in 2013) recommend testing for CMT, but with a tiered approach:

- “Genetic testing should be conducted for the accurate diagnosis and classification of hereditary neuropathies.”
  - This is considered a level A recommendation which is defined as “established as effective, ineffective or harmful (or established as useful/predictive or not useful/predictive) for the given condition in the specified population.”

- “Genetic testing may be considered in patients with cryptogenic polyneuropathy who exhibit a hereditary neuropathy phenotype. Initial genetic testing should be guided by the clinical phenotype, inheritance pattern, and electrodiagnostic features and should focus on the most common abnormalities which are CMT1A duplication/HNPP deletion, Cx32 (GJB1), and MFN2 mutation screening.”
  - This is considered a level C recommendation which is defined as “possibly effective, ineffective or harmful (or possibly useful/predictive or not useful/predictive) for the given condition in the specified population.”

- “There is insufficient evidence to determine the usefulness of routine genetic testing in patients with cryptogenic polyneuropathy who do not exhibit a hereditary neuropathy phenotype.”
  - This is considered a level U recommendation which is defined as “data inadequate or conflicting; given current knowledge, treatment (test, predictor) is unproven.”
DiVincenzo et al.

DiVincenzo et al. [2014] described their experience testing more than 17,000 patients for CMT using a commercially available comprehensive panel of 14 genes. Overall, they identified a mutation in 18.5% of patients. Notably they state that “Among patients with a positive genetic finding in a CMT-related gene, 94.9% were positive in one of four genes (PMP22, GJB1, MPZ, or MFN2). The results of our study in a population in over 17,000 individuals support the initial genetic testing of four genes (PMP22, GJB1, MPZ, and MFN2) followed by an evaluation of rarer genetic causes in the diagnostic evaluation of CMT.”

Expert-authored review

In an expert-authored review, the following genetic testing strategies are recommended:

1. “In families with at least two-generation involvement, known male-to-male transmission, and very slow NCVs (<15 m/sec).”
   - “First, obtain testing for the PMP22 duplication (CMT1A).”
   - “If normal, follow with testing of MPZ (CMT1B) and single-nucleotide variants in PMP22 (CMT1E).”
   - “If both are normal, test for LITAF (CMT1C) and EGR2 (CMT1D).”

2. “In families with at least two-generation involvement and slow NCVs (15-35 m/sec), but without male-to-male transmission, molecular genetic testing of PMP22 (CMT1A), GJB1 (CMTX), MPZ (CMT1B), and LITAF (CMT1C) should be performed sequentially.”

3. “In families with probable X-linked inheritance of the CMT phenotype, molecular genetic testing of GJB1 (CMTX) is appropriate to confirm the diagnosis.”

4. “In individuals with the CMT2 phenotype, MFN2 (CMT2A2) can be tested first followed by testing of MPZ (CMT2I/2J) and GJB1 (CMTX1). Other rarer causes include GDAP1 (CMT2H/2K), NEFL (CMT2E/1F), and EGR2 (CMT1D).”

Regarding cases of early childhood onset: “[…] pathogenic variants in MPZ, PMP22, and EGR2 should be evaluated first as they were most frequent in those individuals with hypotonia and breathing difficulties. Testing of FGD4, PRX, MTMR2, SBF2, SH3TC2, and GDAP1 is indicated in individuals with early foot deformities and delay in motor milestones after an uneventful neonatal period.”

5. For individuals with a negative family history (e.g., a simplex case):
   - “Molecular genetic testing of PMP22dup (CMT1A), MPZ (CMT1B), and GJB1 (CMTX) should be performed on males and females who have no family history of neuropathy because de novo duplications of the 17p11 region (causing CMT1A) are common and because females who have a GJB1 pathogenic variant (causing CMTX1) may be asymptomatic.”
“If no pathogenic variant is identified in any of these three genes testing for rarer subtypes can be considered.”

“Early-onset severe CMT may be caused by mutation of PMP22 (CMT1A or 1E), GDAP1 (CMT4A), EGR2 (CMT4E), PRX (CMT4F), SH3TC2 (CMT4C), FDG4 (CMT4H), or MTMR2 (CMT4B1).”

“An alternative genetic testing strategy is use of a multi-gene panel that includes genes associated with CMT and other genes of interest... A reasonable testing approach is to first test for pathogenic variant(s) in the single most likely gene that best fits the phenotype (e.g., CMT1A or CMT2A). If such testing does not identify the pathogenic variant(s), then proceed to a genetic neuropathy panel followed by exome sequencing if appropriate.”

“Testing of asymptomatic at-risk children is discouraged.”

**Comprehensive CMT panels**

Comprehensive CMT panels test most known genes related to CMT simultaneously, but this is not usually necessary or cost-effective, and therefore not recommended.¹ ⁴

**Criteria**

**Introduction**

Requests for CMT testing are reviewed using these criteria.

**Known Familial Mutation Analysis**

- **Previous Genetic Testing:**
  - No previous genetic testing for the familial mutation, and
  - Pathogenic CMT-related mutation in a 1st or 2nd degree biologic relative, AND

- **Diagnostic Testing for Symptomatic Individuals:**
  - Distal muscle weakness and atrophy, or
  - Weak ankle dorsiflexion (e.g. foot drop), or
  - Distal sensory loss, or
  - Depressed or absent tendon reflexes, or
  - Foot deformity (e.g. high arches, hammer toes, pes cavus), or
Electrodiagnostic studies consistent with a peripheral neuropathy, OR

Predisposition Testing for Presymptomatic/Asymptomatic Individuals:

- Age 18 years or older

**PMP22 Deletion/Duplication Analysis**

- Previous Genetic Testing:
  - No previous PMP22 deletion/duplication analysis, and
  - No known CMT-related mutation in the member’s family, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Distal muscle weakness and atrophy, or
  - Weak ankle dorsiflexion (e.g. foot drop), or
  - Distal sensory loss, or
  - Depressed or absent tendon reflexes, or
  - Foot deformity (e.g. high arches, hammer toes, pes cavus), AND

- The member does not have a known underlying cause for their neuropathy (e.g. diabetic neuropathy, vitamin B12 deficiency, chronic inflammatory demyelinating polyneuropathy, known mutation), AND

- Member’s electrodiagnostic studies are consistent with a primary demyelinating neuropathy

**CMT Neuropathy Multigene Panel**

When a multi-gene panel is being requested and will be billed with the appropriate CPT panel code, 81448, the panel will be considered medically necessary when the following criteria are met:

- Previous Genetic Testing:
  - No previous CMT neuropathy multi-gene panel testing, and
  - No known CMT-related mutation in the member’s family, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Distal muscle weakness and atrophy, or
  - Weak ankle dorsiflexion (e.g. foot drop), or
  - Distal sensory loss, or
  - Depressed or absent tendon reflexes, or
Foot deformity (e.g. high arches, hammer toes, pes cavus), AND

- The member does not have a known underlying cause for their neuropathy (e.g. diabetic neuropathy, vitamin B12 deficiency, chronic inflammatory demyelinating polyneuropathy, known mutation), AND
- The panel includes the genes with the highest diagnostic yield for the member’s suspected CMT neuropathy subtype, AND
- Member’s electrodiagnostic studies are consistent with an axonal neuropathy or combined axonal and demyelinating neuropathy (e.g., CMT1 is NOT the most likely diagnosis), OR
- Member’s electrodiagnostic studies are consistent with a primary demyelinating neuropathy (e.g., CMT1 is the most likely diagnosis) and PMP22 deletion/duplication analysis was previously performed and was negative

Billing and reimbursement considerations

- When separate procedure codes will be billed for individual CMT-related genes (e.g., Tier 1 MoPath codes 81200-81355 or Tier 2 MoPath codes 81400-81408), the entire panel will be approved if the above criteria are met. However, the laboratory will be redirected to the use of an appropriate panel CPT code, 81448, for billing purposes.
- The billed amount should not exceed the list price of the test.
- Broad CMT neuropathy panels may not be medically necessary when a narrower panel is available and more appropriate based on the clinical findings.
- Genetic testing is only necessary once per lifetime. Therefore, a single gene included in a panel or a multi-gene panel may not be reimbursed if testing has been performed previously. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.
- If a panel was previously performed and an updated, larger panel is being requested, only testing for the medically necessary, previously untested genes will be reimbursable. Therefore, only the most appropriate procedure codes for those additional genes will be considered for reimbursement.
- If the laboratory will not accept redirection to 81448 due to their panel not sequencing at least 5 genes, the medical necessity of each billed component procedure will be assessed independently.

  - In general, only a limited number of panel components that are most likely to explain the member’s presentation will be reimbursable. The remaining individual components will not be reimbursable.
When the test is billed with multiple stacked codes, only sequencing of the following genes may be considered for reimbursement, based on electrodiagnostic findings and the family history:

- Primary demyelinating neuropathy with negative PMP22 deletion/duplication analysis (CMT1 suspected): MPZ, PMP22, LITAF (SIMPLE) and EGR2.
- Primary axonal neuropathy (CMT2 suspected): MFN2, MPZ and HSPB1 (HSP27). If there is no evidence of male-to-male transmission in the family, GJB1 (for CMTX) is also reimbursable.
- Combined axonal and demyelinating neuropathy (intermediate CMT suspected): DNM2, YARS, MPZ, and GNB4.

References

Introduction

These references are cited in this guideline.


Chromosomal Microarray for Prenatal Diagnosis

Introduction

Chromosomal microarray testing for prenatal diagnosis is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
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<tr>
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<td>81228</td>
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<tr>
<td>Chromosomal Microarray [SNP], Constitutional</td>
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What are copy number variants in developmental disorders

Introduction

Copy number variants (CNVs) are small deletions and duplications of genetic material and account for a significant proportion of developmental disorders without a clear etiology based on clinical findings.\(^1\) CNVs are detected using chromosomal microarray (CMA) testing. CMA is known by several names including array-comparative genomic hybridization (aCGH) and single-nucleotide polymorphism arrays (SNP-array).\(^1\)

Prevalence

Intellectual disability (ID) and congenital birth defects affect approximately 3-4% of the general population.\(^1\) Major structural birth defects can often be identified prenatally by ultrasound evaluation, while some minor anomalies and ID cannot.\(^2\)

Cause

The etiology of congenital anomalies is complex. Some developmental problems may be caused by environmental factors, such as injury and infection. However, genetic causes also play a significant role.\(^1,2\)
First-line test

Routine chromosome analysis (karyotyping) by chorionic villus sampling (CVS) or amniocentesis has historically been a first-line test in the evaluation of a pregnancy identified with congenital birth defects.\(^2\)

Chromosomal microarray on chorionic villi or amniocytes may be indicated in a pregnancy identified with one or more major structural abnormalities. Identifying an underlying genetic cause in these patients may\(^2\)

- provide diagnostic and prognostic information
- guide prenatal management and decision-making, and
- allow for testing of family members and accurate recurrence risk counseling.

CNV detected in fetus

If a unique CNV is detected in a fetus, it is usually necessary to test both parents to determine whether the CNV is inherited or a new (de novo) genetic change. This information along with parental findings can be used to weigh the possibilities of a benign vs. pathogenic variant. However, even with parental studies, the clinical outcome may remain unclear.\(^5\) A de novo variant is more likely to represent a pathologic abnormality.\(^5, 6\)

Test information

Introduction

Prenatal diagnosis may include chromosomal microarray testing.

Chromosomal microarray

Chromosomal microarray (CMA) testing generally works by fluorescently tagging DNA from a patient test sample with one color and combining it with a control sample tagged in a different color. The two samples are mixed and then added to the array chip, where they compete to hybridize with the DNA fragments on the chip. By comparing the test sample versus the control, computer analysis can determine where genetic material has been deleted or duplicated in the individual.

Coverage and resolution

There are a growing number of CMA testing platforms, including non-chip based applications, which differ in approach and resolution. Testing guidelines do not endorse one CMA over another. However, international consensus guidelines do suggest that CMAs should have coverage better than that offered by a standard karyotype (~5 Mb), and resolution of greater than or equal to 400 kb throughout the genome.\(^4\)
Subtelomeric and disease-specific FISH tests not needed

CMAs include the subtelomeric regions and all known chromosome microdeletion syndrome regions, such as those for 22q11.2 (DiGeorge) syndrome, Williams syndrome (7p11.2), and Smith-Magenis syndrome (17p11.2). Therefore, subtelomeric and disease-specific FISH tests are not needed in parallel with CMA, or as follow-up to normal CMA results.

Cell division in culture not required

In contrast to typical chromosome analysis, CMA testing does not require dividing cells in culture. This makes testing possible in samples that may be difficult to culture, such as those from perinatal losses.5,6

Limitations of CMA

While there are significant advantages of CMA over conventional karyotyping with regard to resolution and yield, there are disadvantages as well. Limitations of CMA include

- the inability to detect
  - balanced translocations or inversions
  - certain forms of polyploidy
  - low level mosaicism, and
  - some marker chromosomes, and
- the detection of CNVs of uncertain clinical significance
- the inability to differentiate free trisomies from unbalanced Robertsonian translocations, and
- the high cost of testing as compared to traditional karyotyping.2

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to chromosomal microarray testing for prenatal diagnosis.

American College of Obstetricians and Gynecologists Committee on Genetics and the Society for Maternal-Fetal Medicine

The American College of Obstetricians and Gynecologists Committee on Genetics and the Society for Maternal-Fetal Medicine (2013) published a joint committee opinion regarding the application of chromosomal microarray in the prenatal setting. This opinion recommended that CMA replaces fetal karyotyping for “patients with a fetus with
one or more major structural anomalies identified on ultrasonographic examination and who are undergoing invasive prenatal diagnosis”.5

**Diagnostic yield of CMA**

Diagnostic yield of CMA testing differs based on clinical presentation. The results of one recent multicenter trial of CMA in the prenatal setting were published in 2012. This study reported that CMA identified a clinically relevant deletion or duplication in 6% of prenatal cases with a structural anomaly and normal karyotype. In addition, 1.7% of prenatal cases with an indication of advanced maternal age or positive screening results and normal karyotype had a clinically relevant deletion or duplication identified by CMA.3

In a large series of fetuses with ultrasound anomalies and normal conventional karyotype, CMA detected chromosome abnormalities in 5% of fetuses and up to 10% in those with 3 or more anatomic abnormalities.2

**Criteria**

- **Genetic Counseling:**
  - Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- **Previous Genetic Testing:**
  - No previous chromosomal microarray testing in the same pregnancy, AND

- **Diagnostic Prenatal Testing:**‡
  - The member has sufficient risk of fetal CNV to justify invasive prenatal diagnosis. [It is important to note that invasive diagnostic procedures such as chorionic villus sampling and amniocentesis are associated with risks; the provider and patient must have determined that the associated benefits outweigh the risks.]

‡Microarray may also be used in association with in utero fetal demise, stillbirth, or neonatal death. If microarray will be performed on fetal tissue after delivery, reference the Chromosomal Microarray Testing for Developmental Disorders guideline.

**Exclusions and other considerations**

- If routine karyotype and CMA are ordered simultaneously, only the most appropriate test based on clinical history will be considered for coverage.
- If CMA has been performed, the following tests are often excessive and are not considered medically necessary. Each test may require medical necessity review.
  - Routine karyotype: Full karyotype in addition to CMA is typically considered excessive. However, a limited 5 cell analysis may be approved in addition to
CMA if criteria for CMA are met. This approval may be subject to claims review to ensure that the appropriate procedure code for a limited 5 cell analysis is billed (CPT 88261 x1).

- FISH analysis
- Telomere analysis
- More than one type of microarray analysis (i.e. if 81228 performed, 81229 is not medically necessary)

Billing and reimbursement considerations

- FISH or other procedure codes that do not accurately describe the test methodology performed (e.g. 88271) are not eligible for reimbursement of CMA.

References

Introduction

These references are cited in this guideline.


Chromosomal Microarray Testing For Developmental Disorders

Introduction

Chromosomal microarray testing for developmental disorders is addressed by this guideline.

Procedures addressed

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What are copy number variants in developmental disorders

Introduction

Copy number variants (CNVs) are small deletions and duplications of genetic material and account for a significant proportion of developmental disorders without a clear etiology based on clinical findings. CNVs are detected using chromosomal microarray (CMA) testing. CMA is known by several names including array-comparative genomic hybridization (aCGH) and single-nucleotide polymorphism arrays (SNP-array).

Prevalence

Intellectual disability (ID) and congenital birth defects affect approximately 3-4% of the general population.\(^1\) Autism spectrum disorders (ASD), including pervasive developmental delay (PDD), are also of increasing concern, with recent CDC incidence figures estimating 1 in 155 affected children.\(^2\)
Cause

The etiology of developmental disorders is complex. Some developmental problems may be caused by environmental factors, such as injury and infection. However, genetic causes also play a significant role.\textsuperscript{1,2}

A causative explanation can be determined in about 40-60\% of patients with ID\textsuperscript{3} and in over 30\% of patients with ASD.\textsuperscript{2} Identifying an underlying genetic cause in these patients may:\textsuperscript{2,3}

\begin{itemize}
  \item provide diagnostic and prognostic information
  \item improve health screening and prevention for some conditions
  \item allow for testing of family members and accurate recurrence risk counseling, and
  \item empower the patient and family to acquire needed services and support.
\end{itemize}

Diagnostic yield

Diagnostic yield differs based on clinical presentation:

\begin{itemize}
  \item Approximately 10-19\% of people with unexplained ID or developmental delay (DD) will have CNVs.\textsuperscript{4-7}
  \item A similar diagnostic yield for ASD is estimated at 7-10\%.\textsuperscript{2}
  \item About 13\% of spontaneous pregnancy losses had CNVs identified in one small prospective study.\textsuperscript{8}
  \item Chromosomal microarray may also be useful in the workup of non-immune fetal hydrops.\textsuperscript{9}
\end{itemize}

Parental testing

If a CNV is detected in a child, it is usually necessary to test both parents to determine whether the CNV is inherited or a new (de novo) genetic change. This information along with parental findings can be used to weigh the possibilities of a benign vs. pathogenic variant. However, even with parental studies, the clinical outcome may remain unclear.\textsuperscript{5} A de novo variant is more likely to represent a pathologic abnormality.\textsuperscript{5,6}

Test information

Introduction

Testing for developmental disorders may include chromosomal microarray testing.
Chromosomal microarray (CMA) testing generally works by fluorescently tagging DNA from a patient test sample with one color and combining it with a control sample tagged in a different color. The two samples are mixed and then added to the array chip, where they compete to hybridize with the DNA fragments on the chip. By comparing the test sample versus the control, computer analysis can determine where genetic material has been deleted or duplicated in the individual.

Coverage and resolution
There are a growing number of CMA testing platforms, including non-chip based applications, which differ in approach and resolution. Testing guidelines do not endorse one CMA over another. However, international consensus guidelines do suggest that CMAs should have coverage better than that offered by a standard karyotype (~5 Mb), and resolution of greater than or equal to 400 kb throughout the genome.4

Subtelomeric and disease-specific FISH tests not needed
CMAs include the subtelomeric regions and all known chromosome microdeletion syndrome regions, such as those for 22q11.2 (DiGeorge) syndrome, Williams syndrome (7p11.2), and Smith-Magenis syndrome (17p11.2). Therefore, subtelomeric and disease-specific FISH tests are not needed in parallel with CMA, or as follow-up to normal CMA results.

Cell division in culture not required
In contrast to typical chromosome analysis, CMA testing does not require dividing cells in culture. This makes testing possible in samples that may be difficult to culture, such as those from perinatal losses.5,6

Limitations of CMA
While there are significant advantages of CMA over conventional karyotyping with regard to resolution and yield, there are disadvantages as well. Limitations of CMA include

- the inability to detect
  - balanced translocations or inversions
  - certain forms of polyploidy
  - low level mosaicism, and
  - some marker chromosomes, and
- the detection of CNVs of uncertain clinical significance
- the inability to differentiate free trisomies from unbalanced Robertsonian translocations, and
Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to chromosomal microarray testing for developmental disorders.

American College of Medical Genetics

The American College of Medical Genetics (ACMG, 2010) Professional Practice and Guidelines Committee recommends CMA as a first-tier test for the evaluation of “multiple anomalies not specific to a well-defined genetic syndrome, apparently non-syndromic developmental delay/intellectual disability, and autism spectrum disorders.”

International Standard Cytogenomic Array Consortium

The International Standard Cytogenomic Array Consortium (ISCA, 2010) recommends offering CMA as a first-tier genetic test, in place of karyotype, for patients with unexplained developmental delay/intellectual disability, autism spectrum disorders, or birth defects.

American College of Obstetricians and Gynecologists and Society for Maternal Fetal Medicine

The American College of Obstetricians and Gynecologists (ACOG, 2013) and Society for Maternal Fetal Medicine (SMFM, 2013) joint committee opinion on chromosomal microarray states that:

- “In cases of intrauterine fetal demise or stillbirth when further cytogenetic analysis is desired, chromosomal microarray analysis on fetal tissue (i.e. amniotic fluid, placenta, or products of conception) is recommended because of the increased likelihood of obtaining results and improved detection of causative abnormalities.”
- “Limited data are available on the clinical utility of chromosomal microarray analysis to evaluate first-trimester and second-trimester pregnancy losses; therefore, this is not recommended at this time.”

Criteria

Introduction

Requests for chromosomal microarray testing for developmental disorders are reviewed using these criteria.
Criteria

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:**
  o No previous chromosomal microarray (CMA) testing, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Testing performed on living child or adult, and
  o Diagnosis cannot be made on clinical evaluation alone, and
  o Common aneuploidy (trisomy 13, 18, 21, or sex chromosome) is not a suspected diagnosis, and
  o One of the following presentations:
    ▪ Apparently nonsyndromic DD/ID, or
    ▪ Autism spectrum disorder, or
    ▪ Multiple congenital anomalies† not specific to a well-delineated genetic syndrome, OR

• Diagnostic Testing for Intrauterine Fetal Demise or Stillbirth:
  o Common aneuploidy (trisomy 13, 18, 21, or sex chromosome) is not a suspected diagnosis, and
  o Multiple congenital anomalies† not specific to a well-delineated genetic syndrome, or
  o Fetal demise or stillbirth occurred in third trimester of pregnancy

†Multiple congenital anomalies defined as 1) two or more major anomalies affecting different organ systems or 2) one major and two or more minor anomalies affecting different organ systems. [Major structural abnormalities are generally serious enough as to require medical treatment on their own (such as surgery) and are not minor developmental variations that may or may not suggest an underlying disorder.]

**Microarray is considered a first tier test in the evaluation of postnatal developmental disorders. Therefore, it often is not necessary to do chromosome analysis or FISH in conjunction with microarray. Microarray requests following such testing will require review.

Exclusions and other considerations

• CMA is not considered medically necessary in cases of family history of chromosome rearrangement in phenotypically normal individuals
• CMA is not considered medically necessary in individuals experiencing infertility, first or second trimester miscarriages, or recurrent pregnancy loss.\textsuperscript{10}

• If routine karyotype and CMA are ordered simultaneously, only the most appropriate test based on clinical history will be considered for coverage.

• If CMA has been performed, the following tests are often excessive and are not considered medically necessary. Each test may require medical necessity review:
  
  o Routine karyotype: Full karyotype in addition to CMA is typically considered excessive. However, a limited 5 cell analysis may be approved in addition to CMA if criteria for CMA are met. This approval may be subject to claims review to ensure that the appropriate procedure code for a limited 5 cell analysis is billed (CPT 88261 x1).
  
  o FISH analysis
  
  o Telomere analysis
  
  o More than one type of microarray analysis (i.e. if 81228 performed, 81229 is not medically necessary)

\textbf{Billing and reimbursement considerations}

FISH or other procedure codes that do not accurately describe the test methodology performed (e.g. 88271) are not eligible for reimbursement of CMA.

\textbf{References}

\textbf{Introduction}

These references are cited in this guideline.


**Introduction**

Chromosome analysis for blood, bone marrow, and solid tumor cancers is addressed by this guideline.

**Procedures addressed**

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<tr>
<td>Chromosome Analysis, Solid Tumor</td>
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**What are chromosome abnormalities in cancer**

**Introduction**

A chromosome abnormality is any difference in the structure, arrangement, or amount of genetic material packaged into the chromosomes. Chromosome abnormalities have been identified in many types of cancer, including leukemias, lymphomas, and solid tumors.¹

**Chromosome abnormalities**

Chromosome abnormalities can include

- deletions
- duplications
- balanced or unbalanced rearrangements, and
- gain or loss of whole or partial chromosomes.

Some chromosome abnormalities are characteristic of certain types of malignancy, and can be used to classify a type or subtype of cancer. For example, the “Philadelphia chromosome” is defined by a common translocation between chromosomes 9 and 22, and indicates chronic myelogenous leukemia in most cases.¹
Disease monitoring and treatment response

These abnormalities can play a key role in the development, diagnosis, and monitoring of cancer.\(^1\)

The cytogenetics of a cancer can also change over time or in response to treatment. Therefore, chromosome analysis can be used to monitor disease progression and treatment response.\(^1\)

Test information

Introduction

Chromosome analysis is routinely performed on bone marrow biopsy for the diagnosis and monitoring of leukemia, lymphoma, and other hematological disorders.

Chromosome analysis

Chromosome analysis (karyotyping) requires stimulating cells to divide, arresting cell division at metaphase when the chromosomes can be seen microscopically, and staining to visualize the banding patterns.\(^2\)

Chromosome analysis identifies any differences from normal that can be seen under the microscope. This includes all of the following:

- entire missing or extra chromosomes
- deletions or duplications within a chromosome that are large enough to be seen by microscope, and
- rearrangements including translocations and inversions.

Chromosome microarray

Smaller copy number changes can be identified using chromosome microarray, although that testing is not routine for cancer.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to chromosome analysis for blood, bone marrow, and solid tumor cancers.

National Comprehensive Cancer Network

The National Comprehensive Cancer Network (NCCN) considers chromosome analysis of a bone marrow biopsy to be routine standard of care in the evaluation of
acute myeloid leukemia (AML), chronic myelogenous leukemia (CML), multiple myeloma (MM), myelodysplastic syndromes (MDS), and Burkitt’s lymphoma (BL).³

**American College of Medical Genetics**

The American College of Medical Genetics (ACMG, 2010) provides technical laboratory guidelines for chromosome studies for acquired abnormalities:⁴

- “A patient with an acquired clonal chromosomal abnormality or one who is at high risk for developing such an abnormality may have multiple cytogenetic studies during the course of his or her disease.”
- “Bone marrow/blood: In most cases, bone marrow is the tissue of choice for analysis of suspected premalignant or malignant hematologic disorders.”
- “Lymph nodes: Common diagnoses include Hodgkin and non-Hodgkin lymphomas, including follicular, diffuse large B-cell, marginal zone, mantle cell, T-cell, and anaplastic large cell lymphoma.”
- “Solid tumors: Cytogenetic analysis of tumor tissue is performed to detect and characterize chromosomal abnormalities for purposes of diagnosis, prognosis, and patient management.”

**Criteria**

Chromosome analysis on a bone marrow biopsy meets criteria without further review when performed in the evaluation of leukemia, lymphoma, and other hematological disorders.

**References**

Introduction

These references are cited in this guideline.


5. American College of Medical Genetics. Section E6.1-6.4 of the ACMG technical standards and guidelines: chromosome studies of neoplastic blood and bone marrow – acquired chromosomal abnormalities. 2016. Available at https://pdfs.semanticscholar.org/34bc/c7ce2ff76bf7637f8c5b9892f823cbd0f68e.pdf
ConfirmMDx for Prostate Cancer Risk Assessment

Procedures addressed

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<td>ConfirmMDx for Prostate Cancer</td>
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What is ConfirmMDx testing for prostate cancer

Definition

The ConfirmMDx™ test (MDx Health) is a proprietary epigenetic assay that measures gene methylation associated with the presence of cancer. Results are intended to assist in determining which patients likely have a true negative biopsy, and which patients are at increased risk for occult cancer. Results may prevent unnecessary repeat biopsies in unaffected men, and triage higher risk patients for repeat biopsies and treatment, as needed.¹

- Prostate cancer is the most common cancer among men, with over 150,000 new cases identified each year in the United States.²³ The median age at diagnosis is 66 years.⁴ Older men are more likely to be affected than younger men, and African American men have higher rates compared to men of other ethnic backgrounds.⁴
- Screening programs for prostate cancer allow for its early detection. Screening is typically performed by prostate-specific antigen (PSA) test and digital rectal examination (DRE).³
- Diagnosis is confirmed by prostate biopsy.⁵⁻⁷ Biopsy is typically performed by collecting approximately 12 needle biopsy cores.⁷
- Initial biopsies only detect 65-77% of prostate cancers, and repeat biopsies are frequently performed.⁸⁻⁹ The false negative rate of biopsy may be as high as 25%.¹⁰

Test information

- ConfirmMDx™ measures the methylation levels (using quantitative methylation PCR) of 3 genes (GSTP1, APC and RASSF1) associated with prostate cancer. The test is performed on formalin-fixed, paraffin-embedded prostate specimens from a 12-core biopsy.
• Results are reported with methylation positive/negative for each biopsy core, along with a map of the regions where methylation is distributed.¹
• Negative predictive value of the test is approximately 90%, based on results of a large, blinded clinical evaluation study.¹¹

Guidelines and evidence

• The National Comprehensive Cancer Network (NCCN) 2016 Clinical Practice Guidelines for Prostate Cancer Early Detection state the following:⁷
  o “Those patients with negative prostate biopsies should be followed with DRE and PSA. Tests that improve specificity in the post-biopsy state-including 4Kscore, PHI, percent free PSA, PCA3, and ConfirmMDx—should be considered in patients thought to be higher risk despite a negative prostate biopsy.”
  o “Biomarkers that improve the specificity of detection are not recommended as firstline screening tests. However, there may be some patients who meet PSA standard for consideration of prostate biopsy, but for whom the patient and/or physician wish to further define the probability of high-grade cancer. A percent free PSA <10%, PHI >35 or 4K score (which provides an estimate of the probability of high-grade prostate cancer) are potentially informative in patients who have never undergone biopsy or after a negative biopsy; a PCA3 score >35 is potentially informative after a negative biopsy.”

Criteria

Coverage for ConfirmMDx will be granted when the following criteria are met:

• No previous ConfirmMDx testing on the same sample when a result was successfully obtained, AND
• No previous 4Kscore testing performed after the most recent negative biopsy when a result was successfully obtained, AND
• Member is not under active surveillance for low stage prostate cancer, AND
• Negative prostate biopsy within the past 24 months, AND
• Member is considered at higher risk for prostate cancer by one or more of the following:
  o Family history of 1st degree relative with prostate cancer diagnosed younger than age 65 years,⁷,¹²,¹³,¹⁴ and/or
  o African American race,⁷,¹²,¹³,¹⁴ and/or
- Known mutation in a gene associated with increased risk of prostate cancer (e.g., BRCA1/2, MLH1, MSH2, MSH6, PMS2, EPCAM)^7,12

References


Corus CAD for Obstructive Coronary Artery Disease

Procedures addressed

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What is the Corus CAD test for obstructive CAD

Definition

Corus CAD is a blood-based test designed to exclude the presence of obstructive CAD in symptomatic patient.

- Heart disease is the leading cause of death for both men and women, accounting for 1 in 6 US deaths. Coronary heart disease is the most common type of heart disease.

- Patients with signs and symptoms of obstructive CAD, the result of a chronic inflammatory process that ultimately results in progressive luminal narrowing and acute coronary syndromes, may be evaluated with a variety of tests according to risk. Coronary angiography is the gold standard for diagnosing obstructive CAD, but it is invasive and associated with a low but finite risk of harm. Thus, coronary angiography is recommended solely for patients at high risk of CAD.

- For patients initially assessed to be at low-to-intermediate risk, observation and noninvasive diagnostic methods, which may include imaging methods such as coronary computed tomography angiography (CCTA) or Myocardial Perfusion Imaging (MPI), may be recommended.

- Even noninvasive imaging methods, however, have potential risks of exposure to radiation and contrast material. Despite efforts to risk stratify patients with noninvasive testing, the subsequent yield of coronary angiography remains low. In one study of nearly 400,000 patients without known CAD undergoing elective coronary angiography, only approximately 38% were found to have obstructive CAD (if the definition of obstructive CAD was broadened to include stenosis of 50% or more in any coronary vessel, the prevalence increased to 41%).
• If symptoms are atypical, then they should be concurrent with at least one risk factor such as high cholesterol, hypertension, family history, smoking, post-menopausal state, morbid obesity, and known non-cardiac vascular disease.  
  
  o It is suggested as a first-line diagnostic modality in the ambulatory care setting ahead of noninvasive imaging to rule out obstructive CAD as the cause of a patient’s symptoms.  
  
  o Corus CAD is intended for use in adult patients with stable, non-acute presentation of symptoms suggestive of obstructive CAD who:  
    ▪ are not diabetic  
    ▪ have not been diagnosed with prior myocardial infarction (MI) nor have had a previous revascularization procedure  
    ▪ are not currently taking steroids, immunosuppressive agents or chemotherapeutic agents  
    ▪ have a known history of obstructive CAD

Test information

• Corus CAD is a gene expression test that integrates the mRNA activity of 23 genes known to be involved in the development of and/or response to atherosclerosis into a single score, which can identify patients without obstructive CAD.  
  
  o Obstructive CAD is defined as:  
    ▪ >50% stenosis in at least one coronary artery by Quantitative Coronary Angiography (QCA) core lab.  
    ▪ >50% QCA stenosis corresponds to 65 – 75% stenosis on clinical angiography.  

• Some of these genes are sex-specific, accounting for key biological differences between men and women in the development of CAD.  

• A proprietary algorithm converts gene expression changes to a score that ranges from 1 to 40. The specific numeric value is translated into a percentage likelihood of the patient having obstructive CAD.  
  
  o Patients with scores less than or equal to 15 (“low score”) have a low likelihood (<8%) of having obstructive CAD.  

• The test potentially eliminates 46% of patients (those with scores less than or equal to 15) from further cardiac workup due to the low likelihood of their symptoms being caused by obstructive CAD.
Test performance in the intended use population (disease prevalence of about 15\%):\textsuperscript{8}

- Sensitivity = 89\%
- Specificity = 52\%
- Negative predictive value (NPV) = 96\%

Guidelines and evidence

- Corus CAD is not mentioned in any of the current applicable American College of Cardiology (ACC) or American Heart Association (AHA) guidelines, policy statements or scientific statements.\textsuperscript{9,10,11,12}

- Clinical validity studies:
  - PREDICT\textsuperscript{7}
    - Prospective, multi-center, blinded study in 39 U.S. sites.
    - 1569 non-diabetic patients undergoing cardiac catheterization.
    - The predictive accuracy of the Corus CAD score was good, with AUC = 0.70± 0.02.
    - Corus CAD significantly improved the ability to detect underlying obstructive CAD compared with clinical assessment (based on the Diamond-Forrester [D-F] clinical risk score).
    - Test significantly improved MPI accuracy in identifying underlying obstructive CAD.
  - COMPASS\textsuperscript{8}
    - Prospective, multi-center study in 19 U.S. sites.
    - 431 non-diabetic symptomatic patients scheduled for MPI.
    - Primary end point: Receiver-operating characteristics (ROC) analysis to discriminate less than or equal to 50\% stenosis by QCA.
    - Corus CAD significantly improved the ability to detect underlying obstructive CAD compared to MPI.
    - Corus CAD outperformed clinical factors as assessed by D-F criteria and Morise score.
    - Six-month follow-up on 97\% of patients showed that 27 of 28 patients with major adverse cardiovascular events (MACE) or revascularization had scores >15.

- Clinical utility studies:
o **IMPACT-CARD**

- Prospective, single-center study at Vanderbilt University Medical Center.
- 83 prospective non-diabetic symptomatic patients presenting to the cardiologist's office with 83 matched historical controls.
- A change in the diagnostic testing pattern pre/post Corus CAD testing was noted in 48/83 patients (58% observed vs. 10% expected change, p<0.001).
  - Low Score (less than or equal to 15): 56% decreased intensity of testing; 44% had no change.
  - High Score (>15): 52% increased intensity of testing; 39% had no change.
- 71% reduced testing rate in prospective group vs. historical cohort (p<0.001).
- Follow-up (chart review/phone call) in 180 d to ensure plan was followed & get MACE.
  - 0 patients of 161 (0.0%; 97% follow-up) had MACE.

o **IMPACT-PCP**

- Prospective, multi-center study of 4 practice sites.
- 251 non-diabetic symptomatic patients presenting to the primary care physician’s (PCP) office.
- 51% of patients had a low score (less than or equal to 15).
- A change in the diagnostic testing pattern pre/post Corus CAD testing was noted in 145/251 patients (58% observed vs. 10% expected change, p<0.001).
  - Low Score (less than or equal to 15): 60% decreased intensity of testing; 38% had no change.
  - High Score (>15): 40% increased intensity of testing; 47% had no change.
- Follow-up (chart review/phone call) in 30 days to ensure plan was followed & record MACE.
  - 1 patient of 247 (0.4%) had “MACE” (hemorrhagic CVA 5 days after testing, later determined not to meet criteria for MACE).

o **REGISTRY-1**

- Prospective, multi-center chart review of non-diabetic patients with typical and/or atypical symptoms suggestive of obstructive CAD at 7 sites.
342 patients presenting to PCP office.

- Study designed for 670 patients with an interim look at 335.
- Study stopped early due to meeting primary endpoint.

49% of patients had a low score (less than or equal to 15).

Patients with low Corus CAD score (less than or equal to 15) had 94% decreased odds of referral versus patients with high score (> 15) (p < 0.0001).

- For every 10 point decrease in score, had 14x decreased likelihood of referral to cardiology or advanced cardiac testing (p < 0.0001).

Referral rate: 6% for low scores, 70% for high scores.

Followed for minimum of 180 days (Avg. F/U = 267 days).

- 21 cardiac caths, 2 from patients with low scores; 19 from patients with high scores.
- MACE rate = 1.5% (5/342); 1 in low score group (percutaneous coronary intervention [PCI]), 3 in high score group (PCI x 2 and myocardial infarction [MI]) plus another not judged to be related to CV disease.

Recently completed clinical trials

- The PRESET Registry: A Registry to Evaluate Patterns of Care Associated With the Use of Corus® CAD in Real World Clinical Care Settings. ClinicalTrials.gov Identifier: NCT01677156.16
  - Primary outcome measures: “To describe referral patterns for cardiac care and testing within 1 month after gene expression testing.”16

- Effect of Exercise Stress Testing on Peripheral Gene Expression Using CORUS™ CAD Diagnostic Test. ClinicalTrials.gov Identifier: NCT01486030.17
  - Primary outcome measures: “Gene expression score difference between peak exercise and baseline.”17

- PROspective Multicenter Imaging Study for Evaluation of Chest Pain - The PROMISE Trial. ClinicalTrials.gov Identifier: NCT01174550.18
  - Primary outcomes measures: “Time to primary endpoint as defined as a composite of death, myocardial infarction (MI), major complications from cardiovascular (CV) procedures or testing, and unstable angina hospitalization. The Kaplan-Meier events rates (cumulative percentage of participants with an event) were estimated for the anatomic and functional diagnostic test groups.”18

While clinical utility studies have demonstrated that Corus CAD results can influence clinical decision making, there is insufficient data to demonstrate that
these decisions improve health outcomes as measured by the presence of major adverse cardiovascular events (MACE).

- The relatively small number of patients in the clinical utility trials (total n = 676) and the distribution of these patients (across less than a dozen practice sites) also raises questions about whether these results are generalizable to the entire US.

Criteria

- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  - In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

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<tr>
<td>Cxbladder Detect</td>
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<tr>
<td>Cxbladder Monitor</td>
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What is Cxbladder

Definition

Cxbladder is a family of non-invasive urinary biomarker tests manufactured by Pacific Edge Diagnostics. Cxbladder was developed as an alternative or adjunct to conventional tests for the initial diagnosis of bladder cancer or for later disease recurrence.\(^1\)\(^2\)

- Bladder cancer is typically diagnosed using a combination of cytologic evaluation of urine, imaging tests, and cystoscopy. However, patients have reported that cystoscopy is uncomfortable and expensive, and as a result, investigators are exploring alternative methods of to detect bladder cancer.
- The following tests are included in the Cxbladder family:\(^2\)
  - Cxbladder Triage
    - Used to rule out bladder cancer at an early stage.\(^2\)
  - Cxbladder Detect
    - Used to assess the probability of bladder cancer.\(^2\)
  - Cxbladder Monitor
    - Used to assess the probability of disease recurrence.\(^2\)
  - Cxbladder Resolve
    - Used to identify patients with high grade or late stage bladder cancer.\(^2\)
Test information

- According to the manufacturer, levels of messenger RNA (mRNA) of five biomarker genes, including MDK, HOXA13, CDC2, IGFBP5, CXCRI, are believed to be in higher concentrations in urine samples of patients with bladder cancer.

- The Cxbladder test involves the extraction, purification, and quantification of mRNA of the 5 biomarkers by reverse transcription (RT) quantification polymerase chain reaction (RT-qPCR).²

  - Cxbladder Triage
    - Combines bladder cancer risk factors as well as urinary biomarkers to rule out the presence of bladder cancer.²

  - Cxbladder Detect
    - Analyzes five urinary biomarkers to identify bladder cancer.²

  - Cxbladder Monitor
    - Combines clinical information and urinary biomarkers to assess the chance that bladder cancer has recurred.²

  - Cxbladder Resolve
    - Used to identify high grade or late stage bladder cancer in patients with haematuria.²

Guidelines and evidence

- The National Comprehensive Cancer Network (NCCN, 2017) Clinical Practice Guidelines do not specifically comment on the Cxbladder tests. However, the guidelines states the following regarding the use of other available urinary biomarkers:³

  - “Management of bladder cancer is based on the pathologic findings of the biopsy specimen, with attention to histology, grade, and depth of invasion. These factors are used to estimate the probability of recurrence and progression to a more advanced stage. Consideration may be given to FDA-approved urinary biomarker testing by fluorescence in situ hybridization or nuclear matrix protein 22 in monitoring for recurrence.”

- In 2011, the U.S. Preventive Services Task Force https://www.uspreventiveservicestaskforce.org/Page/Document/evidence-summary/14/bladder-cancer-in-adults-screening updated its 2004 evidence review with regard to bladder cancer screening, and reported the following:⁴

  - “no study evaluated the sensitivity or specificity of tests for hematuria, urinary cytology, or other urinary biomarkers for bladder cancer in asymptomatic persons without a history of bladder cancer. The positive predictive value of
screening is less than 10% in asymptomatic persons, including higher-risk populations. No study evaluated harms associated with treatment of screen-detected bladder cancer compared with no treatment.”

- “screening tests that might be feasible for primary care include tests for hematuria, urinary cytology, and other urinary biomarkers. The U.S. Preventive Services Task Force (USPSTF) last reviewed the evidence on bladder cancer screening in 2004 but found insufficient evidence to guide a recommendation.”

- The diagnostic accuracy of Cxbladder has been compared with standard diagnostic methods in individuals suspected of urothelial cancer due to gross hematuria in the urine.\textsuperscript{1,5-8}

- According to Chou et al. (2015), the diagnostic accuracy study carries a medium risk of bias and reported sensitivity of the Cxbladder test of 82% (95% CI, 70% to 90%) and specificity of 85% (95% CI, 81% to 88%).

- There are available studies that evaluated the effects of on patient-relevant outcomes (survival, quality of life) of using the Cxbladder test (clinical utility).

- Additional research is needed to assess how Cxbladder testing will be used in the disease management of patients with cancer. Questions persist regarding if Cxbladder has sufficient clinical utility to replace invasive cystoscopy or if Cxbladder has the potential to augment or clarify uncertain results obtained using conventional diagnostic methods.

- Ongoing clinical trials

- The Cxbladder Monitoring Study: A Clinical, Non-Intervention Study of the Cxbladder Urine Test for the Detection of Recurrent Urothelial Carcinoma (UC).\textsuperscript{9}

  - Primary Outcome Measures:
    - “Proportion of participants with bladder cancer who are correctly identified as having cancer (true positives) and no cancer (true negatives) by the Cxbladder test.”
    - “The Cxbladder test results will be compared to that of cystoscopy, which is the gold standard method for diagnosing urothelial cancer; the true positive and true negative rates will be measured, along with the false positive and false negative rates of the test. The results will be reported as sensitivity and specificity of the Cxbladder test for detecting urothelial cancer in patients with recurrent disease.”
    - “Probability that patients identified as having cancer and no cancer by the Cxbladder test truly have cancer (positive predictive value; PPV), and truly have no cancer (negative predictive value; NPV) respectively.”
Criteria

- These tests are considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
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References


CYP2C19 Variant Analysis for Clopidogrel Response

Procedures addressed

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<tr>
<td>CYP2C19 Genotyping</td>
<td>81225</td>
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What is CYP2C19 testing for clopidogrel response

Definition

Clopidogrel (Plavix®) is a prodrug that must be converted by CYP2C19 to an active form to inhibit clot formation. Variants in the CYP2C19 gene can result in reduced or enhanced enzyme function, which in turn affects clopidogrel activity.\(^1,2\) The CYP2C19*2 genetic variant alone accounts for about 12% of the variability in clopidogrel response.\(^3\)

- CYP2C19 variant testing can be used to predict response to clopidogrel and modify the therapeutic strategy when necessary.\(^1,2\) CYP2C19 variant testing determines if a person is a poor, intermediate, extensive, or ultrarapid metabolizer.
  - A person with two nonfunctional alleles (any combination of *2-*8) is classified as a poor metabolizer.\(^1,2\) About 2-3% of Caucasians and blacks and up to 20% East Asians are poor metabolizers.
  - People with one loss-of-function allele (*1 and any combination of *2-*8) are intermediate metabolizers and represent 30-50% and 40-45% of these populations, respectively.\(^4,5\)
  - The CYP2C19*17 variant is associated with increased enzyme function or gain of function carriers. Prevalence of the CYP2C19*17 allele is typically <5% in Asians and about four times higher in Caucasian and African populations.\(^6\)

- Several studies have demonstrated a reduced effectiveness of clopidogrel in people with reduced CYP2C19 metabolism. Poor metabolizers may be at increased risk of nonfatal stroke, MI, or death from any cause in patients with poor metabolism.\(^7-12\) In contrast, an analysis of the CURE trial and ACTIVE trial, involving 5059 genotyped patients with acute coronary syndromes, did not find an effect of CYP2C19 genotype on outcome in homozygous, heterozygous or in those who were not carriers of the loss of function alleles.\(^12\)
• CYP2C19 ultrarapid metabolizers (*17 carriers) may be at increased risk for clopidogrel-related bleeding. However, a recent study showed ultrarapid metabolizers had a greater benefit from clopidogrel therapy than non-carriers, without increased bleeding events.

Test information
• CYP2C19 testing identifies the most common gene variants and is performed on buccal or blood samples.
  o CYP2C19*1 is the normal functioning allele.
  o The most common loss of function alleles are *2 and *3.
  o CYP2C19*4, *5, *6, *7, and *8 alleles are much less common and are associated with absent or reduced CYP2C19 enzyme function.
  o CYP2C19*17 allele is associated with increased enzyme function or gain-of-function carriers.

Guidelines and evidence
• U.S. Food and Drug Administration (FDA) approved product labeling for clopidogrel (Plavix®) was updated in July 2017 to revise a boxed warning of the diminished effectiveness in individuals with poor CYP2C19 metabolism. The revised boxed warning provides more general guidance about the impact of reduced platelet activity:
  o “The effectiveness of Plavix® results from its antiplatelet activity, which is dependent on its conversion to an active metabolite by the cytochrome P450 (CYP) system, principally CYP2C19.”
  o “Plavix® at recommended doses forms less of the active metabolite and so has a reduced effect on platelet activity in patients who are homozygous for nonfunctional alleles of the CYP2C19 gene, (termed “CYP2C19 poor metabolizers”).”
  o “Tests are available to identify patients who are CYP2C19 poor metabolizers. Consider use of another platelet P2Y12 inhibitor in patients identified as CYP2C19 poor metabolizers.”
• In January 2015, the American Heart Association published a Scientific Statement on Basic Concepts and Potential Applications of Genetics and Genomics for Cardiovascular and Stroke Clinicians. They noted:
  o “No cardiovascular pharmacogenetic application has yet been fully validated or widely adopted.”
“In aggregate, the available data suggests that patients at highest risk for cardiovascular events (those who have undergone PCI [percutaneous coronary intervention] and are in the acute period after the procedure) may have worse outcomes on clopidogrel if they are reduced-function variant carriers.”

“...the magnitude of benefit of clopidogrel in a given patient population influences the risk associated with CYP2C19 loss-of-function variants. Specifically, if the magnitude of benefit is small, the impact of genotype on clopidogrel efficacy may also be small. Therefore, the risk of genotype appears to be greatest among patients for whom clopidogrel has the greatest efficacy (i.e., largest risk reduction), specifically those undergoing percutaneous coronary intervention with stenting. Meta-analyses suggest that this group may be at up to 3- to 4-fold increased risk for stent thrombosis among *2 variant carriers.”

In December 2013, the American Heart Association published a Scientific Statement on Genetics and Genomics in the Prevention and Treatment of Cardiovascular Disease. They surmised:

“...the magnitude of benefit of clopidogrel in a given patient population influences the risk associated with CYP2C19 loss-of-function variants. Specifically, if the magnitude of benefit is small, the impact of genotype on clopidogrel efficacy may also be small. Therefore, the risk of genotype appears to be greatest among patients for whom clopidogrel has the greatest efficacy (i.e., largest risk reduction), specifically those undergoing percutaneous coronary intervention with stenting. Meta-analyses suggest that this group may be at up to 3- to 4-fold increased risk for stent thrombosis among *2 variant carriers.”

In July 2013, the Clinical Pharmacogenetics Implementation Consortium published an update to their antiplatelet therapy recommendations for acute coronary syndrome (ACS) patients undergoing percutaneous coronary intervention (PCI) based on CYP2C19 status. They concluded:

Genotype-directed therapy could identify those with ACS/PCI who benefit most from alternative antiplatelet therapy. Current data do not support the use of CYP2C19 genotype data to guide treatment in other scenarios.

Standard dosing of clopidogrel, as recommended in the product label, is warranted among ACS/PCI patients with a predicted CYP2C19 extensive metabolizer or ultrarapid metabolizer phenotype (i.e., *1/*1, *1/*17, and *17/*17).

If genotyping identifies a patient as a CYP2C19 PM (i.e., any combination of *2 through *8), literature supports the use of an alternative antiplatelet agent (e.g., prasugrel (Effient®) or ticagrelor (Brilinta®) when not contraindicated.

Data support switching to an alternative antiplatelet agent for CYP2C19 IMs (e.g., *1/*2, *1/*3, and *2/*17) when not contraindicated. However, given the wide inter-individual variability in residual platelet activity observed among clopidogrel-treated IMs, other factors that may place an IM at increased risk of a CV event (or adverse bleeding event) must be considered to most effectively individualize therapy.
o It is currently premature to support an increased dosing strategy based on CYP2C19 genotype. Large clinical trials that evaluated higher-dose clopidogrel in ACS/PCI patients with high on-treatment platelet reactivity have concluded that adjusting clopidogrel dose on the basis of platelet function monitoring alone does not reduce the incidence of death from CV causes, nonfatal myocardial infarction, or stent thrombosis.

• In August 2012, the American College of Cardiology Foundation (ACCF)/American Heart Association (AHA) Task Force on Practice Guidelines, in collaboration with the American College of Emergency Physicians, the Society for Cardiovascular Angiography and Interventions, and the Society of Thoracic Surgeons, commented:
  o Genetic testing for CYP2C19 loss-of-function alleles may be considered on a case-by-case basis, especially for patients who experience recurrent ACS events despite ongoing therapy with clopidogrel.

• In May 2012, the American Heart Association published a Policy Statement on Genetics and Cardiovascular Disease. They concluded:
  o “...it is now unambiguously clear that the use of standard doses of clopidogrel in patients with CYP2C19 loss-of-function variants is associated with an increased frequency of major adverse cardiovascular events and, in particular, of in-stent thrombosis among patients receiving drug-eluting stents.”

• In July 2010, the American College of Cardiology Foundation (ACCF) and the American Heart Association (AHA) issued a Clopidogrel Clinical Alert for approaches to the FDA black box warning which include the following points:
  o An emphasis on adherence to the existing ACCF/AHA guidelines for the use of antiplatelet therapy.
  o Clinicians should be aware that genetic variability in CYP enzymes alter clopidogrel metabolism and that diminished responsiveness to clopidogrel has been associated with adverse patient outcomes in registry experiences and clinical trials.
  o The predictive value of pharmacogenomic testing is very limited at this time, but studies are ongoing.
  o Evidence is insufficient to recommend routine genetic testing or platelet function testing but may be considered for people at moderate to high risk for poor outcomes. If a person is tested and found to be a poor metabolizer, other therapies should be considered:
    ▪ For coronary patients - consider prasugrel (Effient®) (NOTE: Or ticagrelor (Brilinta®), now that it has been approved).
- For TIA/stroke patients - consider aspirin or aspirin plus extended release dipyridamole. Prasugrel is contraindicated in TIA/stroke (NOTE: ticagrelor (Brilinta®) should not be used in patients with active pathological bleeding or a history of intracranial hemorrhage).

  - For people who experience adverse reactions (i.e. adverse CV event or thrombosis, not bleeding) on clopidogrel several options exist:
    - Clopidogrel can be switched to prasugrel (NOTE: Or ticagrelor, now that it has been approved).
    - Clopidogrel dose can be increased (though little data exists).
    - Platelet function testing may be performed to determine if patients are clopidogrel non-responders.
    - For stroke patients, aspirin alone or combination of aspirin plus extended-release dipyridamole can be considered.

  - Higher loading doses and maintenance doses of clopidogrel have been found to improve platelet inhibition and might be considered alternatives for high-risk patients who respond poorly to clopidogrel. New antiplatelet drugs such as prasugrel and if approved, ticagrelor (NOTE: ticagrelor has been approved), are additional alternatives. Other possibilities are adding cilostazol (Pletal®) to standard doses of aspirin and clopidogrel, though data with this combination is still accruing. Follow up platelet function testing might be considered to ensure adequate platelet inhibition.

Criteria

- Previous Testing:
  - No previous genetic testing of CYP2C19, AND

- Personal History:
  - Currently on clopidogrel therapy, or
  - Use of clopidogrel therapy is being proposed for a patient at moderate to high risk for a poor outcome, such as:
    - Experiencing symptoms consistent with ACS when percutaneous coronary intervention is an option, and/or
• Considering a drug-eluting stent

Exclusions

- Current data do not support the use of CYP2C19 genotype data to guide treatment in other scenarios, including altering the dosing recommendation for clopidogrel based on the CYP2C19 genotype.

References

11. Aleil B, Jacquemin L, De Poli F, et al. Clopidogrel 150 mg/day to overcome low responsiveness in patients undergoing elective percutaneous coronary intervention:


CYP2C9 and VKORC1 Testing for Warfarin Response

Procedures addressed

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<td>VKORC1 Genotyping</td>
<td>81355</td>
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<td>Warfarin responsiveness testing by genetic technique using any method</td>
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What is Warfarin sensitivity testing?

Definition

Warfarin (Coumadin®) is a commonly prescribed anticoagulant with a narrow therapeutic range and a 20-fold inter-individual variation in dose requirements. Incorrect dosage, especially during the initial dosing phase, is associated with either severe bleeding or failure to prevent thromboembolism.

- Approximately 21% of patients who receive anticoagulant therapy will experience a major or minor bleeding event. Environmental and genetic factors combined influence 55% of warfarin dose variability and include: age, height, body mass index (BMI), gender, diet, genetic variations in CYP2C9 and VKORC1, use of concomitant medications and indication for warfarin.

- The activity of two genes [cytochrome P450 2C9 (CYP2C9) and vitamin K epoxide reductase complex subunit-1 (VKORC1)] impact the rate of warfarin metabolism and account for up to 40% of the inter-individual dose requirements for warfarin.

- CYP2C9 is a p450 enzyme that influences warfarin pharmacokinetics by impacting the rate of metabolism. Poor or intermediate metabolizing 2C9 variants are seen in between 2% to 20% of the population depending on ethnicity. Carriers of alleles *2 and *3 have decreased warfarin metabolism and may require lower warfarin doses.

- Vitamin K activity is important to the blood’s ability to clot. VKORC1 influences the pharmacodynamics and sensitivity of warfarin on the vitamin K cycle. Approximately 14% to 89% of the population display VKORC1 enzyme inhibition making them more sensitive to warfarin. Carriers of VKORC1 AA genotype (high warfarin
sensitivity) require a significantly lower warfarin dose compared to individuals with genotype GA or GG.\(^4\)

- Testing these two genes predicts variability in warfarin dosage requirements. The presence of gene variants in CYP2C9 and VKORC1 indicate that more careful dosing and monitoring is required to achieve therapeutic anticoagulation and to decrease risk of bleeding or clotting during warfarin dose titration.

**Test information**

- The CYP2C9 allele is thought to be the predominant cause of the variation of warfarin dosing.\(^5\)
- There are approximately 37 alleles reported in the CYP complex, however many do not have a functional impact.
  - Two alleles, *2 and *3 (CYP2C9*2 and CYP2C9*3) are linked to a slower metabolism of warfarin, thereby needing an increase in warfarin dose. These alleles are found in approximately 12.2% and 7.9%, respectively, of the European Caucasian population.\(^6\)
  - Other variants, *4, *5, and *6 are seen in the Asian and African American populations, but typically around a <1% incidence.\(^7\)
- Diagnosis of these alleles can occur through sequence analysis of the CYP2C9 and VKORC1 genes. Mutation analysis detects virtually 100% of alleles.\(^2\)

**Guidelines and evidence**

- There has been a mixed response to genotyping from professional associations, payors, and other organizations, largely because data supporting the utility of genetic testing to improve clinical endpoints is conflicting. For example, two recent meta-analyses came to opposite conclusions:
  - A genotype-guided dosing strategy did not result in a greater percentage of time that the INR was within the therapeutic range, fewer patients with an INR greater than 4, or a reduction in major bleeding or thromboembolic events compared with clinical dosing algorithms.\(^5\)
  - Genotype-guided initial dosing is able to reduce serious bleeding events by approximately 50% (RR = 0.47; 95% CI, 0.23-0.96; P = 0.040) compared with clinically-guided dosing approaches.\(^7\)
- The Clinical Pharmacogenetics Implementation Consortium (CPIC, 2017) guidelines state “This guideline recommends that pharmacogenetic warfarin dosing be accomplished through the use of one of the pharmacogenetic dosing algorithms...The two algorithms provide very similar dose recommendations...The warfarindosing.org website contains both algorithms, the Gage algorithm as the
primary algorithm and the IWPC [International Warfarin Pharmacogenetics Consortium] algorithm as the secondary algorithm..." It also notes "In patients of African ancestry, CYP2C9*5, *6, *8, *11 are important for warfarin dosing. If these genotypes are not available, warfarin should be dosed clinically without consideration for genotype."\(^8\)

- In January 2015, the American Heart Association published a Scientific Statement on Basic Concepts and Potential Applications of Genetics and Genomics for Cardiovascular and Stroke Clinicians.\(^9\) They noted:
  - "No cardiovascular pharmacogenetic application has yet been fully validated or widely adopted."
  - "Building on these early findings, additional clinical studies of warfarin pharmacogenetics are underway."

- The American College of Medical Genetics (ACMG, 2008) and the American College of Chest Physicians (ACCP, 2008) both suggest against routine genotyping to guide warfarin dosing until better evidence is available to support a policy decision, but the ACMG does say that testing might be useful to explain unexpected warfarin responses.\(^6, 10\)

- An FDA Advisory Committee convened in November of 2005 voted unanimously that "sufficient mechanistic and clinical evidence exists to support the recommendation to use lower doses of warfarin for individuals with genetic variations in CYP2C9 and VKORC1 that lead to reduced activities." Furthermore, their report states "genotyping people in the induction phase of warfarin therapy would reduce adverse events and improve achievement" of a stable dose for anticoagulation.\(^8\) Product labeling for Coumadin (warfarin) has been updated based on FDA recommendation to include a table recommending initial dosing ranges for patients with different combinations of CYP2C9 and VKORC1 genotypes. Labeling also includes the range of expected therapeutic warfarin doses based on CYP2C9 and VKORC1 genotypes.\(^4\)

- Publications based on the European Pharmacogenetics of Anticoagulant Therapy (EU-PACT) trial data have indicated that pharmacogenomics-driven dosing algorithms may need to be age- and ethnicity-specific. This revised approach to dosing algorithms will need additional research and validation.\(^11, 12\)

**Criteria**

This test is considered investigational and/or experimental.

- Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical
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- In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References


CYP2D6 Variant Analysis for Drug Response

Procedures addressed

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<tr>
<td>CYP2D6 Genotype Cascade Mayo Clinic</td>
<td>0028U</td>
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What is CYP2D6 testing

Definition

The cytochrome P450 2D6 (CYP2D6) enzyme is involved in metabolizing many medications, including tamoxifen, tetrabenazine, deutetraphenazine and eliglustat.

- Studies suggest that certain variations (polymorphisms) in the CYP2D6 gene result in reduced or absent enzyme function, which may lead to lower levels of active tamoxifen metabolites and reduced treatment efficacy.
- CYP2D6 testing has, therefore, been proposed to guide adjuvant therapy decisions in some circumstances.

  - Tamoxifen users:
    - Postmenopausal women considering tamoxifen have a choice between tamoxifen and aromatase inhibitors. Results of CYP2D6 testing could influence that decision, although data about the utility of testing has been mixed (see Guidelines/Evidence below for details).
    - Testing is not indicated for perimenopausal and premenopausal women with hormone-positive breast cancer. Tamoxifen is the current standard of care for these patients, and no alternative treatment plans have been approved.
    - Testing is not recommended for patients considering tamoxifen in the preventative setting.

  - Tetrabenazine (Xenazine) users:
    - Tetrabenazine is a “vesicular monoamine transporter 2 (VMAT) inhibitor indicated for the treatment of chorea associated with Huntington’s disease.”
CYP2D6 testing is used to help guide tetrabenazine dosage in patients that are being considered for a tetrabenazine dose greater than 50mg.\(^2\)

For extensive and intermediate metabolizers, “the maximum recommended daily dose is 100 mg and the maximum recommended single dose is 37.5 mg.”\(^2\)

“In poor metabolizers, the initial dose and titration is similar to extensive metabolizers except that the recommended maximum single dose is 25 mg, and the recommended daily dose should not exceed a maximum of 50 mg.”\(^2\)

- **Deutetrabenazine (Austedo) users:**
  - Deutetrabenazine is a “reversible depletor of monoamines (such as dopamine, serotonin, norepinephrine, and histamine) from nerve terminals”. Metabolites of deutetrabenazine, are reversible vesicular monoamine transporter 2 (VMAT) inhibitors.\(^4\)
  - Deutetrabenazine is indicated for the treatment of chorea associated with Huntington’s disease.\(^4\)
  - Maximum recommended daily dosage of deutetrabenazine is 48 mg. In patients who are poor CYP2D6 metabolizers, however, the total daily dosage should not exceed 36 mg (with a maximum single dose of 18 mg).\(^4\)

- **Eliglustat (Cerdelga) users:**
  - Eliglustat is a “glucosylceramide synthase inhibitor indicated for the long-term treatment of adult patients with Gaucher disease type 1 who are CYP2D6 extensive metabolizers (EMs), intermediate metabolizers (IM), or poor metabolizers (PM) as detected by an FDA-cleared test.”\(^3\)
  - CYP2D6 intermediate metabolizers and extensive metabolizers are recommended to take a dose of 84 mg twice daily. This dosage requirement is decreased to 84 mg once a day for CYP2D6 poor metabolizers.\(^3\)

### Test information

- CYP2D6 testing is usually performed on a buccal swab or blood sample using polymerase chain reaction (PCR) to look for certain common variants.
- Genotype results are generally assigned a metabolizer phenotype:

<table>
<thead>
<tr>
<th>Phenotype</th>
<th>Genotype(s)(^11)</th>
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<tr>
<td>Poor Metabolizer (PM)</td>
<td>Two CYP2D6 inactive variants</td>
</tr>
<tr>
<td>Intermediate Metabolizer (IM)</td>
<td>One normal and one inactive variant</td>
</tr>
</tbody>
</table>
Phenotype | Genotype(s)\textsuperscript{11}
--- | ---
One inactive and one reduced-activity variant | Two reduced-activity variants
Extensive Metabolizer (EM) | Two normal CYP2D6 alleles
Ultrarapid Metabolizer (UM) | More than two copies of the normal CYP2D6 allele

- The frequency of the CYP2D6 metabolizer phenotypes varies with ethnicity. About 5-10% of Caucasians are poor metabolizers, while the frequency is much lower in Africans and Asians.\textsuperscript{10}

**Guidelines and evidence**

- Tetrabenazine, deutetrabenazine, and eliglustat:
  - CYP2D6 is listed as an FDA-approved biomarker for both tetrabenazine and eliglustat.\textsuperscript{12}
  - Product labeling for tetrabenazine, deutetrabenazine, and eliglustat address CYP2D6 testing.\textsuperscript{2,3,4}

- Tamoxifen
  - Evidence-based guidelines from the National Comprehensive Cancer Network (NCCN, 2016) state: “At this time, based on current data the [NCCN Breast Cancer] panel recommends against CYP2D6 gene testing for women being considered for tamoxifen therapy.”\textsuperscript{8} (category 2A: The recommendation is based on lower level evidence and there is uniform NCCN consensus)
  - Practice guidelines from the American Society of Clinical Oncologists (ASCO, 2009) state: “Given the limited evidence, CYP2D6 testing is currently not recommended in the preventive setting.”\textsuperscript{9}
  - Two important large clinical trials have most directly addressed clinical utility of CYP2D6 testing for tamoxifen response.\textsuperscript{13,14} Both found that CYP2D6 genotype did not predict long-term outcome among tamoxifen users.
    - Regan et al. performed CYP2D6 variant testing on tumor tissue from 4393 patients enrolled in the BIG 1-98 trial and evaluated the association with breast cancer recurrence. BIG 1-98 was an international, randomized double-blind trial that compared tamoxifen monotherapy, letrozole (an aromatase inhibitor) monotherapy, and sequential therapy (2 years of one and 3 years of another). Patients were mostly Caucasian and all had postmenopausal, hormone receptor-positive, operable breast cancer. Results found a non-statistically significant association between metabolizer phenotype and recurrence (poor metabolizer vs. extensive metabolizer HR = ...
0.58, 95% CI = 0.28 to 1.21). The authors concluded “The results of this study do not support using the presence or absence of hot flushes or the pharmacogenetic testing of CYP2D6 to determine whether to treat postmenopausal breast cancer patients with tamoxifen.”

- Similarly, Rae et al. found no association between CYP2D6 genotype and breast cancer recurrence in people treated with tamoxifen from the randomized double-blind Arimidex, Tamoxifen, Alone or in Combination (ATAC) trial (n=1203; poor metabolizer vs. extensive metabolizer HR = 1.25, 95% CI = 0.55 to 3.15). The authors conclude “The results do not support the hypothesis that CYP2D6 genotype predicts clinical benefit of adjuvant tamoxifen treatment among postmenopausal breast cancer patients.”

Criteria

CYP2D6 testing will be granted when the following criteria are met:

Testing for Tetrabenazine Response

- No previous CYP2D6 testing performed, AND
- Member has a diagnosis of Huntington's disease, AND
- Treatment with tetrabenazine is being considered in a dosage greater than 50mg per day, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

Testing for Deutetrabenazine Response

- No previous CYP2D6 testing performed, AND
- Member has a diagnosis of Huntington's disease, AND
- Treatment with deutetrabenazine is being considered in a dosage greater than 36mg per day, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

Testing for Eliglustat Response

- No previous CYP2D6 testing performed, AND
- Member has a diagnosis of Gaucher disease, AND
- Treatment with eliglustat is being considered, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Exclusions and other considerations

• CYP2D6 testing for tamoxifen response is considered investigational/experimental and, therefore, not eligible for reimbursement.
• CYP2D6 testing for all other indications is considered investigational/experimental and, therefore, not eligible for reimbursement.
• CYP2D6 Genotype Cascade from Mayo Clinic (CPT 0028U) is considered investigational/experimental and, therefore, not eligible for reimbursement.

References


12. FDA. Table of valid genomic biomarkers in the context of approved drug labels. Available at [http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm](http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm).


Introduction

Cystic fibrosis testing is addressed by this guideline.

Procedures addressed

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What is cystic fibrosis

Definition

Classic cystic fibrosis (CF) is a genetic disorder that causes chronic lung disease, pancreatic insufficiency, and male infertility.\(^1,2\) It is caused by mutations in the CFTR gene.\(^1\)

Prevalence

CF affects approximately 1 in 3200 Caucasian newborns.\(^1\) While CF is most common in Caucasians, it can occur in any ethnic group.\(^2\)

Inheritance

CF is an autosomal recessive condition. Males and females are equally likely to be affected.\(^1\) If both parents are carriers of CF, the risk for a pregnancy to be affected is 1 in 4 (25%).\(^1\) If one partner is affected with CF and the other partner is a carrier, the risk for a pregnancy to be affected is 1 in 2 (50%). Preimplantation and prenatal diagnosis are available for couples known to be at-risk.
Prognosis

Patient registry data from 2010 indicate that the median lifespan for people with classic CF is about 38 years. Treatment advances continue to extend the life of patients with CF. Several therapies in development or currently available target specific CFTR gene mutations, such as the FDA-approved Kalydeco™ for people with the G551D mutation.

Detection

Most signs of CF cannot be identified on prenatal ultrasound examination. However, pregnancies in which fetal echogenic bowel is identified on ultrasound are at an increased risk to be affected with CF.

Prenatal diagnosis for CF can be performed on a sample from chorionic villus sampling (CVS) or amniocentesis:

- If both parents are known carriers, a mutation panel that includes both parental mutations is typically the test of choice.
- If only one parent is a carrier, or if testing is indicated because of echogenic bowel, testing with a large mutation panel or sequencing offers greater sensitivity.

Newborn screening (NBS) programs include screening for CF, though the screening protocol may vary by state.

CFTR-related disorders

Several other conditions share some clinical similarities to CF, are also caused by mutations in the CFTR gene, but do not meet the diagnostic criteria for classic CF. These are called “CFTR-related disorders” and include congenital bilateral absence of vas deferens (CBAVD/CAVD), acute recurrent or chronic pancreatitis, and some respiratory tract conditions such as bronchiectasis, sinusitis, and nasal polyps. Such conditions may also be called non-classic CF.

CBAVD is frequently identified after semen analysis shows absent sperm (azoospermia). CBAVD is often caused by one severe CFTR mutation and one mild mutation (including the 5T allele). At least one CFTR mutation can be found in up to 80% of men with CAVD. Because of this association, CFTR analysis is routinely performed for men with azoospermia.

Test information

Introduction

Testing for cystic fibrosis tests may include CFTR mutation panels, CFTR sequencing, CFTR deletion/duplication analysis, intron 8 poly-T analysis, or CFTR known-familial mutation analysis.
CFTR mutation panels

The American College of Medical Genetics has defined a panel of 23 common, pan-ethnic mutations that occur at a frequency of at least 0.1% in patients with cystic fibrosis. While this panel was created for carrier screening purposes, the CF diagnostic guidelines also endorse its use in that setting for most patients. Laboratories performing mutation panel testing routinely include all of these mutations. Many laboratories expand their panels with more mutations intended to increase the detection rate, particularly in non-Caucasian populations. Expanded mutation panels generally test for 70 or more CFTR mutations. The detection rates of expanded panels vary by laboratory and depend on the mutations included and the patient's ethnicity.

CFTR sequencing

CFTR sequencing detects more than 98% of mutations. Sequencing is generally performed in reflex to normal mutation panel results, and reserved for specific situations in which a mutation panel is insufficient.

CFTR deletion/duplication analysis

CFTR deletion/duplication analysis identifies mutations that sequencing would not find. This test is performed in reflex to normal sequencing results.

CFTR known familial mutation analysis

Once the mutations in affected or carrier family members have been identified, other relatives and at-risk pregnancies can be tested for those mutations. Mutation panels are often used in this situation, as long as they include the family mutation. If the family mutation is rare or unique, testing for just that mutation may be needed.

Intron 8 poly-T analysis

Intron 8 poly-T analysis identifies the number of thymidine bases in intron 8 of the CFTR gene. The three common variants are 5T, 7T, and 9T. The 5T variant is considered a mild mutation with reduced penetrance, while 7T and 9T are considered normal variants.

Testing is typically done in reflex to the identification of an R117H mutation by CFTR mutation panel testing. The 5T variant also modifies the effect of the R117H mutation if the two mutations are located on the same chromosome. R117H is a mild CFTR mutation included in the standard panel recommended by the American College of Medical Genetics. If R117H is identified by CF testing, reflex testing for the 5T variant is indicated to provide information relevant to genetic counseling.

5T variant analysis

5T variant analysis may also be performed alone or included in CFTR testing panels when the testing is done specifically to evaluate a man with CAVD. The 5T variant is more commonly found in men with CAVD in the absence of other
symptoms of CF. In one large study, 25% of men with CAVD who had CFTR mutations identified had at least one copy of the 5T variant identified.\textsuperscript{10}

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to cystic fibrosis testing.

American College of Obstetrics and Gynecology and American Society for Reproductive Medicine

Evidence-based guidelines from the American College of Obstetrics and Gynecology (ACOG) (2005\textsuperscript{9}, limited update 2011\textsuperscript{11}) and the American College of Medical Genetics and Genomics (ACMG) (2004)\textsuperscript{7} recommend that CF carrier screening using a mutation panel be offered to all couples who are pregnant or planning a pregnancy or those with a family history of CF.

- ACOG adds “It is becoming increasingly difficult to assign a single ethnicity to individuals. It is reasonable, therefore, to offer CF carrier screening to all patients. Screening is most efficacious in the non-Hispanic white and Ashkenazi Jewish populations.”\textsuperscript{9}
- These guidelines state that expanded mutation screening or sequencing may be beneficial in:
  - An individual with a family history of CF with an unknown mutation\textsuperscript{7,9}
  - An individual whose reproductive partner is a known CF carrier, has CF, or has CAVD\textsuperscript{7,9}

American Society for Reproductive Medicine in partnership with the Society for Male Reproduction and Urology

Consensus-based guidelines from the American Society for Reproductive Medicine in partnership with the Society for Male Reproduction and Urology (2008) recommend cystic fibrosis testing for men with CAVD and their partners, stating:\textsuperscript{12}

- “A man with CBAVD should be assumed to harbor a CFTR mutation. Therefore, before any treatments using his sperm, testing should be offered to the female partner to exclude the possibility (approximately 4%) that she too may be a carrier. All such couples should be offered genetic counseling.” These guidelines do not specify a preferred testing methodology.
Cystic Fibrosis Foundation

Consensus-based guidelines from the Cystic Fibrosis Foundation (2008) outlines the ways in which a CF diagnosis can be established (see below). Characteristic features of CF include chronic sinopulmonary disease (such as persistent infection with characteristic CF pathogens, chronic productive cough, bronchiectasis, airway obstruction, nasal polyps, and digital clubbing), gastrointestinal/nutritional abnormalities (including meconium ileus, pancreatic insufficiency, chronic pancreatitis, liver disease, and failure to thrive), salt loss syndromes, and obstructive azoospermia in males (due to CAVD).

These guidelines state that, “Individuals with sweat chloride values in the intermediate range (30 to 59 mmol/L for infants under age 6 months; 40 to 59 mmol/L for older individuals) should undergo extensive CFTR mutation analysis (that is, expanded panel of CFTR mutations, evaluation for deletions, or gene sequencing).”

When at least one characteristic feature is present, a diagnosis of CF can be confirmed by:

- Two abnormal sweat chloride values
- Identification of two CFTR gene mutations
- Characteristic transepithelial nasal potential difference (NPD)

In the absence of symptoms, a CF diagnosis can be established in:

- A newborn with two CFTR gene mutations identified via newborn screening
- A pregnancy found to have two CFTR mutations on prenatal testing

American College of Obstetrics and Gynecology

Evidence-based guidelines from the American College of Obstetrics and Gynecology (2011) recommend: “For couples in which both partners are carriers, genetic counseling is recommended to review prenatal testing and reproductive options.” In the discussion, ACOG adds that for “A woman [who] is a carrier of a CF mutation and her partner is unavailable for testing or paternity is unknown. Genetic counseling to review the risk of having an affected child and prenatal testing options and limitations may be helpful.”

Society of Obstetricians and Gynaecologists of Canada

No US evidence-based guidelines have been identified that specifically address CF prenatal diagnosis for echogenic bowel. However, it is standard practice and evidence-based guidelines from the Society of Obstetricians and Gynaecologists of Canada (SOGC, 2005) state: “Grade 2 and 3 echogenic bowel is associated with both chromosomal and nonchromosomal abnormalities. Expert review is recommended to initiate the following:...laboratory investigations that should be offered, including fetal...”
karyotype, maternal serum screening, DNA testing for cystic fibrosis (if appropriate), and testing for congenital infection (II-2 A).” [Evidence level II-2: “Evidence from well-designed cohort (prospective or retrospective) or case-control studies, preferably from more than one centre or research group.” Recommendation classification A: “There is good evidence to support the recommendation for use of a diagnostic test, treatment, or intervention.”]

Criteria

Introduction

Requests for cystic fibrosis testing are reviewed using these criteria.

CFTR Standard Panel Testing

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy)\textsuperscript{14}, AND

• Previous Genetic Testing:
  o No previous genetic testing for CFTR mutation(s), AND

• Diagnostic Testing for Symptomatic Individuals:
  o Individuals with an intermediate range/equivocal sweat chloride test (30-55mmol/L in infants, or 40-59mmol/L after 6 months of age), or
  o Individuals with a negative sweat chloride test when
  o Symptoms of CF are present, or
  o Idiopathic chronic (acute recurrent) pancreatitis present with non-focal workup, or
  o Infants with meconium ileus or other symptoms indicative of CF and are too young to produce adequate volumes of sweat for sweat chloride test, or
  o Infants with an elevated IRT value on newborn screening, or
  o Males with oligospermia/azoospermia/congenital absence of vas deferens (CAVD)\textsuperscript{8,15,16}, OR

• Carrier Screening:
  o Be of reproductive age, and
  o Have potential and intention to reproduce, or
  o Have reproductive partner with family history of CF, or
  o Have reproductive partner with CAVD, or
Currently pregnant, OR

- Prenatal Testing:
  - Either biological parent has a diagnosis of CF, or
  - Family history of CF is present, or
  - Both parents are carriers of CF mutations, or
  - Echogenic bowel has been identified on ultrasound in a fetus, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**CFTR Known Familial Mutation Analysis**

- Genetic Counseling:
  - Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy)\(^\text{14}\), AND

- Previous Testing:
  - No previous genetic testing for known CFTR family mutation(s), or
  - Previous CFTR panel testing was not inclusive of known family mutation, AND

- Carrier Screening:
  - Familial CFTR mutation(s) in known biologic relative, OR

- Prenatal Testing:
  - Either biological parent is a known carrier of a CFTR mutation, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**CFTR Sequencing**

- Genetic Counseling:
  - Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy)\(^\text{14}\), AND

- Previous Genetic Testing:
  - Previous CFTR Standard Panel was negative (no mutation found) or only one mutation was found, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Individuals with a negative or equivocal sweat chloride test, and
    - Unexplained COPD or bronchiectasis with unexplained chronic or recurrent sinusitis and abnormal pulmonary function tests (PFTs), or
- Idiopathic chronic (acute recurrent) pancreatitis is present, or
  - Infants with meconium ileus or other symptoms indicative of CF and are too young to produce adequate volumes of sweat for sweat chloride test, or
  - Infants with an elevated IRT value on newborn screening and a negative 23 mutation panel, OR

- Carrier Screening:
  - An individual with a family history of CF with an unknown mutation, or
  - An individual whose reproductive partner is a known CF carrier, has a diagnosis of CF, or has a diagnosis of CAVD, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**CFTR Deletion/Duplication Analysis**

- Genetic Counseling:
  - Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous CFTR deletion/duplication testing, and
  - Previous CFTR Gene Sequencing was negative (no mutation found) or only one mutation was found, and
  - No known familial mutation, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**CFTR Intron 8 Poly T Analysis**

- Genetic Counseling:
  - Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous CFTR intron 8 poly T testing, AND

- Diagnostic Testing:
  - Diagnosis of male infertility (congenital absence of vas deferens [CAVD], obstructive azoospermia), or
  - Diagnosis of non-classic CF, OR

- Carrier Testing:
- CFTR mutation analysis performed and R117H mutation detected, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


DecisionDx Uveal Melanoma

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<tr>
<td>DecisionDx-PRAME</td>
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What is DecisionDx Uveal Melanoma

Definition

Melanoma is the most deadly form of skin cancer. The median age at diagnosis is 59 years. According to NCCN, “The incidence of melanoma continues to increase dramatically, at an overall rate of 33% for men and 23% for women from 2002 to 2006.”

- Risk factors for melanoma include the following: “skin type, previous history of melanoma, multiple clinically atypical moles or dysplastic nevi, and a positive family history of melanoma.” Some individuals have a genetic mutation which increases their risk of melanoma.
- Screening for melanoma typically involves a visual skin examination. A skin biopsy of the lesion and pathological examination can diagnose melanoma.
- “Uveal Melanoma, commonly known as ocular or choroidal melanoma, is a rare cancer of the eye.” Although this type of melanoma is treatable in most cases, approximately 50% of individuals with uveal melanoma will develop metastasis.
- DecisionDx Uveal Melanoma (DecisionDx-UM) is a test designed to assess an individual’s risk of metastasis.

Test information

- DecisionDx-UM measures gene expression of 15 genes present in an ocular melanoma tumor. This test is designed to assess the risk of metastasis within 5 years.
- DecisionDx-UM test results are reported as follows:
  - Class 1A – very low risk (2%) of metastasis within 5 years
- Class 1B – moderate risk (21%) of metastasis within 5 years
- Class 2 – high risk (72%) of metastasis within 5 years

- DecisionDx-PRAME is a test that can be added on to the DecisionDx-UM assay. According to Castle Biosciences, “PRAME (preferentially expressed antigen in melanoma) is a cancer testis antigen gene that is not expressed at appreciable levels in normal adult tissues but its expression can become aberrantly increased in some types of cancer, including sarcoma, hematological malignancies, breast cancer, and melanoma.”

Guidelines and evidence

- The National Comprehensive Cancer Network (NCCN, 2017) states the following regarding gene expression tests for melanoma:
  - “While there is interest in newer prognostic molecular techniques such as gene expression profiling to differentiate benign from malignant neoplasms, or melanoma at low versus high risk for metastasis, routine (baseline) genetic testing of primary cutaneous melanomas (before or following SLNB) is not recommended outside of a clinical study (trial).”
  - “Gene expression profiling for melanoma could be an enormously valuable contribution to understanding the biology of the disease. However, the difficulty of embracing gene expression profiling as an independent predictor of outcome is illustrated by the inconsistency of results across studies aimed at defining the most predictive gene sets for melanoma.”

- Based on the review of the available peer-reviewed published literature, the DecisionDx-UM 15-gene assay has sufficient evidence for use as a prognostic test in patients diagnosed with primary, localized uveal melanoma to assist clinicians with predicting disease severity and improving disease management strategies.

- Given that effective treatments for advanced metastases are limited, detecting metastasis at an earlier disease stage may improve personalized treatment strategies and may lead to improved patient survival. As evidenced by analytic studies, the GEP assay tends to yield a classification result more often than cytological examination. Compared with cytological classification and chromosomal classification, clinical validity studies demonstrate that the GEP assay provides a more accurate risk assessment of disease progression. Some studies suggest that the prognostic value of the test may also be increased by obtaining multiple FNAB samples compared with a single sample. In summary, using this assay may assist clinicians in evaluating how frequently and how intensively patients diagnosed with localized uveal melanoma should undergo disease surveillance. Information provided by the test may help clinicians develop an effective management plan once a diagnosis of uveal melanoma is established.

- Evidence regarding clinical utility is still limited and indirect since no well-designed studies are available that directly compare patient health outcomes as a result of
patient management with and without DecisionDx-UM testing. Future well-designed studies evaluating clinical utility will further establish the ability of DecisionDx-UM to improve overall survival or progression-free survival among patients with localized uveal melanoma.

- According to Castle Biosciences, “Although the exact clinical implications of PRAME are still under investigation and will be prospectively evaluated in an upcoming multicenter trial led by Dr. Harbour, PRAME has the potential to be important for more precise risk assessment and therapeutic options.”

**Criteria**

- DecisionDx-UM testing is considered medically necessary when the following criteria are met:
  
  o No previous DecisionDx-UM testing performed after current diagnosis when a result was successfully obtained, AND
  
  o Member has primary, localized uveal melanoma, AND
  
  o No evidence of metastatic disease, AND
  
  o Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Exclusions and other considerations**

- DecisionDx-PRAME testing (CPT code: 81479) is considered experimental and/or investigational and will not be reimbursed.

**References**


Dentatorubral-Pallidoluysian Atrophy Testing

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What is Dentatorubral-Pallidoluysian Atrophy

Definition

Dentatorubral-pallidoluysian atrophy (DRPLA) is a progressive neurologic disorder.

- Age of onset ranges from one year of age to 72 years of age; the mean age of onset is 31.5 years of age.¹
  - In adults (over ~age 20), DRPLA presents as ataxia, choreoathetosis, and dementia or character changes.
  - In people under ~age 20, DRPLA typically manifests with progressive intellectual deterioration, behavior changes, ataxia, myoclonus, and seizures.
  - Neuropathology demonstrates degeneration of the dentatorubral and pallidoluysian systems.² In addition, white matter lesions have been described.¹
- DRPLA is also known as Myoclonic Epilepsy with Choreoathetosis; Naito-Oyanagi Disease; Haw River Syndrome; Ataxia, Chorea, Seizures, and Dementia.¹
- Although initially thought to be a disorder of the Japanese population, DRPLA has been diagnosed in people from a variety of other ethnic backgrounds. Its prevalence is estimated to be about 0.48 in 100,000 in the Japanese population based on a study conducted by Tsuji et al in 2008.³
- The diagnosis of DRPLA is based on presenting findings, family history, and the results of molecular genetic testing demonstrating an expansion of the CAG trinucleotide/polyglutamine tract in ATN1.¹
  - Normal alleles typically have a repeat length of 6 to 35.
  - Individuals with DRPLA have a full penetrance allele with repeat length > 48 repeats, usually 48-93.¹
So-called “mutable normal” alleles may exist, i.e., alleles with repeats between 36 and 47. Mutable normal alleles do not result in symptoms for the individual, but they are unstable and may increase in size when transmitted to offspring.\textsuperscript{1}

The age of onset and clinical presentation is indirectly correlated with the size of the expansion. On average, people with large expansions have earlier onset than those with a smaller number of repeats.\textsuperscript{1,2}

Although the size of the trinucleotide repeat is inversely correlated with the age of onset, the number of repeats cannot be used for specific prediction of symptoms or age of onset in an asymptomatic person. Repeat length is estimated to account for 50-68\% of the variability in age of onset, the other contributing factors are not known.\textsuperscript{4}

DRPLA is inherited in an autosomal dominant manner. Males and females are equally likely to be affected. A person with DRPLA has a 50\% chance of passing the ATN1 mutation to each of his/her children.

Most individuals with DRPLA have inherited the mutation from a parent. The parent may not have had signs of DRPLA because the number of repeats he or she had were below the “threshold” for manifesting symptoms (“mutable normal” or “intermediate” alleles) or the number of repeats was within the disease-causing range, but small in number thus the parent with the abnormal allele has not yet developed symptoms.

Unaffected persons with mutable normal or intermediate alleles may pass this allele to offspring and the allele may undergo intergenerational expansion to a disease-causing range. The amount that of expansion depends upon the size of the repeat and gender of the transmitting parent. When the expansion is inherited from the father, increase in size of the expansion tends to be larger than when the disease-causing allele is inherited from the mother.\textsuperscript{1} As a result, individuals who inherit the mutation from their father tend to have onset of disease 26-29 years earlier than their affected parent; when inheritance is from the mother, the onset of disease is about 14-15 years earlier.\textsuperscript{1}

Test information

DRPLA molecular genetic testing identifies the number of CAG trinucleotide/polyglutamine repeats in ATN1. A repeat length of >48 confirms the diagnosis of disease. Testing is \textgreater \textasciitilde99\% accurate. Once the diagnosis is confirmed in an affected relative, pre-symptomatic/predictive testing, prenatal diagnosis, and preimplantation genetic diagnosis are available to at-risk family members.

Guidelines and evidence

No U.S. guidelines exist for genetic testing for DRPLA.
• A 2010 expert-authored review states:\(^1\)
  o “The diagnosis of dentatorubral-pallidoluysian atrophy (DRPLA) is established in individuals with disease-causing CAG trinucleotide expansions in ATN1 (DRPLA) who are:”
    ▪ “Under age 20 years and have ataxia, myoclonus, seizures, and progressive intellectual deterioration;”
    ▪ “Over age 20 years and have ataxia, choreoathetosis, dementia, and psychiatric disturbance.”
  o “Most individuals diagnosed with DRPLA have an affected parent. It is appropriate to evaluate both parents of an affected individual with molecular genetic testing even if they are asymptomatic.”
  o “Testing of asymptomatic at-risk adults for DRPLA in the presence of nonspecific or equivocal symptoms is predictive testing, not diagnostic testing. When testing at-risk individuals for DRPLA, it is helpful to test for the CAG expansion in an affected family member to confirm the molecular diagnosis in the family.”
  o “At-risk asymptomatic adult family members may seek testing in order to make personal decisions regarding reproduction, financial matters, and career planning. Others may have different motivations including simply the “need to know.” Testing of asymptomatic at-risk adult family members usually involves pre-test interviews in which the motives for requesting the test, the individual's knowledge of DRPLA, the possible impact of positive and negative test results, and neurologic status are assessed.”
  o “Requests from parents for testing of asymptomatic at-risk individuals during childhood require sensitive and understanding counseling. Consensus holds that individuals under age 18 at risk for adult-onset disorders should not have testing in the absence of symptoms.”
  o “If the disease-causing mutation has been identified in the family, prenatal diagnosis for pregnancies at increased risk is possible by analysis of DNA extracted from fetal cells obtained by amniocentesis (usually performed at ~15-18 weeks’ gestation) or chorionic villus sampling (usually performed at ~10-12 weeks’ gestation).”

**Criteria**

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous ATN1 testing for DRPLA, AND
• Diagnostic Testing for Symptomatic Individuals:
  o less than 20 years of age and 2 or more of the following:
    ▪ Ataxia
    ▪ Myoclonus
    ▪ Seizures
    ▪ Progressive intellectual deterioration/behavior changes
    ▪ Affected 1st degree biologic relative or Japanese/Haw River descent, OR
  o 20 years of age or older and 2 or more of the following:
    ▪ Ataxia
    ▪ Choreoathetosis
    ▪ Affected 1st degree biologic relative or Japanese/Haw River descent, OR

• Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  o ATN1 CAG trinucleotide expansion detected in 1st degree biologic relative, or
  o Suspected DRPLA in a deceased 1st, 2nd or 3rd degree biologic relative who was not genetically diagnosed

References

DPYD Variant Analysis for 5-FU Toxicity

Procedures addressed

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<tbody>
<tr>
<td>DPYD Genotyping</td>
<td>81232</td>
</tr>
</tbody>
</table>

What is dihydropyrimidine dehydrogenase (DPYD) testing for 5-FU toxicity

Definition

5-fluorouracil (5-FU) is a common, broad-spectrum chemotherapeutic agent.1,2

- Dihydropyrimidine dehydrogenase (DPD) is the enzyme involved in the first step of the breakdown of 5-fluorouracil (5-FU), to 5-fluoro-5, 6-dihydro-fluorouracil (FUH2).3-5
- More than 80% of a dose of 5-fluorouracil is metabolized by DPD to FUH2. This metabolite has much lower toxicity than 5-FU.4
- A small percentage (≤10%) of 5-FU patients develop grade III-IV toxicity (neutropenia, nausea, vomiting, severe diarrhea, stomatitis, mucositis, and neuropathy),6,7 which can be life-threatening.
- One primary cause for toxicity is DPD deficiency.4,6,7 An estimated 0.1-3% of the population has DPD deficiency, caused by variants in the dihydropyrimidine dehydrogenase (DPYD) gene.5,8 In particular, about 1% of the population has the DPYD IVS14 +1G>A variant (also called DPYD*2A) that is found to be associated with a seven-fold increased risk for grade III/IV 5-FU toxicity.9-11
- Individuals found to have a DPYD genetic variant require lowered drug doses or alternative therapies.7,9
- Testing may also be used to investigate a possible cause of toxicity if a person experiences adverse effects while on a 5-FU based therapy.6

Test information

- Testing for the DPYD variants DPYD*2A (rs3918290), DPYD*13 (rs55886062), and rs67376798 A (on the positive chromosomal strand) should be considered prior to initiating treatment with 5-fluorouracil and capecitabine for most patients.
• Testing is widely available and highly accurate for the DPYD*2A (rs3918290) variant (>99% detection rate). Testing is also available for other variants.

Guidelines and evidence

• The FDA has acknowledged DPD deficiency as a risk factor for 5-FU related toxicity on multiple drug inserts. However, testing is not explicitly recommended or required prior to treatment initiation.
  o The 2016 updated prescribing information for Xeloda® lists DPD deficiency as a contraindication.12
    ▪ “…patients with certain homozygous or certain compound heterozygous mutations in the DPD gene that result in complete or near complete absence of DPD activity are at increased risk for acute early-onset of toxicity and severe, life-threatening, or fatal adverse reactions caused by XELODA…”
    ▪ “Patients with partial DPD activity may also have increased risk of severe, life-threatening, or fatal adverse reactions caused by XELODA.”
    ▪ “No XELODA dose has been proven safe for patients with complete absence of DPD activity. There is insufficient data to recommend a specific dose in patients with partial DPD activity as measured by any specific test.”

  o Similar warnings exist about use in patients with DPD enzyme deficiency in the Contraindications section of the prescribing information for Carac® Cream and Efudex® topical solutions and cream.13,14
  o DPYD variant testing is listed by the FDA as a valid biomarker in the context of approved drug labeling.15

• In October 2013, the Clinical Pharmacogenetics Implementation Consortium (CPIC) published a guideline on Dihydropyrimidine Dehydrogenase Genotype and Fluoropyrimidine Dosing.16 They recommended:
  o “…alternative drug for patients who are homozygous for DPYD non-functional variants—*2A (rs3918290), *13 (rs55886062), and rs67376798 A (on the positive chromosomal strand)—as these patients are typically DPD deficient.”
  o “Patients who are homozygous for DPYD*2A, *13, or rs67376798 may demonstrate complete DPD deficiency and the use of 5-fluorouracil or capecitabine is not recommended in these patients.”
  o “However, available evidence does not clearly indicate a degree of dose reduction needed to prevent fluoropyrimidine related toxicities…”
  o “Consider a 50% reduction in starting dose for heterozygous patients (intermediate activity).”
“…followed by an increase in dose in patients experiencing no or clinically tolerable toxicity to maintain efficacy, a decrease in dose in patients who do not tolerate the starting dose to minimize toxicities…”

- Though not specified in other professional guidelines, there is general consensus that given the large number of patients treated each year with 5-FU, and the human and economical cost of severe toxic side effects, pre-therapeutic detection of DPD deficiency should be considered.\(^7,17,18\)

**Criteria**

DPD testing for genetic variants DPYD*2A (rs3918290), DPYD*13 (rs55886062), and rs67376798 A (on the positive chromosomal strand) is indicated in individuals considering or currently on therapy with any 5-FU containing drug including, but not limited to:

- 5-fluorouracil (Fluorouracil®, Adrucil®)
- Capecitabine (Xeloda®)
- Fluorouracil topical formulations (Carac®, Efudex®, Fluoroplex®)

**References**

6. Halmos B, Krishnamurthi SS. Enterotoxicity of chemotherapeutic agents. UpToDate, database online v. 17.3.


15. FDA Table of valid genomic biomarkers in the context of approved drug labels. Available at: http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm.


Duchenne and Becker Muscular Dystrophy Testing

Procedures addressed

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<th>Procedures addressed by this guideline</th>
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<td>81403</td>
</tr>
<tr>
<td>DMD Deletion/Duplication Analysis</td>
<td>81161</td>
</tr>
<tr>
<td>DMD Sequencing</td>
<td>81408</td>
</tr>
</tbody>
</table>

What are Duchenne and Becker Muscular Dystrophy

Definition

Duchenne muscular dystrophy (DMD) is an X-linked inherited neuromuscular disorder affecting 1 in 3500 boys.\(^1\,2\) It is typically diagnosed by age 5.

- The main clinical findings of DMD include:\(^1\)
  - rapidly progressive skeletal muscle weakness and wasting that is more proximal than distal
  - a delay in motor milestones (such as walking at 18 months)
  - calf pseudohypertrophy
  - wheelchair dependency by 13 years
  - dilated cardiomyopathy
  - reduced life expectancy
  - greatly elevated serum creatine kinase (CK) concentration

- Genetic testing confirms a clinical diagnosis in affected males. Muscle biopsy may be used for diagnosis when molecular testing does not find a mutation.\(^2\)
- Although this is an X-linked disorder, some females may exhibit symptoms, and some carriers may develop related symptoms later in life, including muscle weakness and cardiomyopathy.\(^1\)
Becker muscular dystrophy (BMD) is a similar disorder, caused by mutations in the same gene, which has a later age of onset and is less common than Duchenne. It is typically diagnosed by age 10, and people with BMD are often still able to walk into their 20s. The typical features include:  

- progressive skeletal muscle weakness
- wheelchair dependence after age 16 years if at all
- flexion contractures of the elbows
- preservation of neck flexor muscle strength (differentiates BMD from DMD)
- dilated cardiomyopathy
- greatly elevated serum CK concentration

**Test information**

- DMD deletion/duplication testing is the best first test, which detects genetic changes in about 65-80% of males with DMD and up to 95% of males with BMD.  
DMD deletion/duplication testing can also be used to identify a mutation in a known or suspected carrier female if an affected male is not available for molecular analysis.  

- DMD sequence analysis will identify about 30-35% of DMD genetic changes.  
DMD sequencing analysis can also be used to identify a mutation in a known or suspected carrier female, if an affected male is not available for molecular analysis.  

- Once the familial mutation is identified, at-risk family members can have reliable and accurate testing for just that mutation.

**Guidelines and evidence**

- The Centers for Disease Control and Prevention (CDC) selected the Care Considerations Working Group (2010) to create guidelines for diagnosis and management of DMD:

  - “Testing for a DMD mutation in a blood sample is always necessary even if DMD is first confirmed by the absence of dystrophin protein expression on muscle biopsy. The results of genetic testing provide the clinical information required for genetic counseling, prenatal diagnosis, and consideration for future mutation-specific therapies.”

  - “If deletion/duplication testing is negative, then dystrophin gene sequencing should be done to look for point mutations or small deletions/insertions.”
“Full characterization of the mutation (deletion endpoints or exact position of any point mutation) is required to allow correlation of the predicted effect of the mutation on the reading frame of the gene, which is the major determinant of the phenotypic variability seen in dystrophinopathy, as well as to determine eligibility for the mutation-specific treatments currently in trials”.


- “Carriers of DMD or BMD should be made aware of the risk of developing cardiomyopathy and educated about the signs and symptoms of heart failure.”

- “Carriers of DMD or BMD should be referred for evaluation by a cardiac specialist with experience in the treatment of heart failure and/or neuromuscular disorders. Patients should undergo initial complete cardiac evaluation in late adolescence or early adulthood or at the onset of cardiac signs and symptoms, if these signs or symptoms appear earlier.”

- “Carriers should be screened with a complete cardiac evaluation at a minimum of every 5 years starting at 25 to 30 years of age.”

- “Treatment of cardiac disease is similar to that outlined for boys with DMD or BMD.”

### Criteria

#### DMD Known Familial Mutation Analysis

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous genetic testing of DMD, AND
- Diagnostic Testing for Symptomatic Individuals:
  - DMD mutation identified in 1st, 2nd, or 3rd degree biologic relative(s), OR
- Carrier Screening and Predictive Testing for Presymptomatic/Asymptomatic At-Risk Individuals:
  - DMD mutation identified in 1st, 2nd, or 3rd degree biologic relative(s), OR
- Prenatal Testing for At-Risk Pregnancies:
  - DMD mutation identified in mother or sibling
DMD Deletion/Duplication Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous DMD genetic testing, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Progressive symmetric muscle weakness (proximal greater than distal)—e.g., leg, pelvic and shoulder girdle muscles, and calf hypertrophy, and positive Gower maneuver, or
  o Elevated serum CK concentration, and
  o Progressive symmetric muscle weakness (proximal greater than distal)—e.g., leg, pelvic and shoulder girdle muscles, or
  o Calf hypertrophy, or
  o Positive Gower maneuver, or
  o Male gender, or
  o Onset of symptoms by early adulthood (usually by adolescence), or
  o Delayed motor milestones, or
  o Gait problems; waddling gait or
  o Learning difficulties, or
  o Quadriceps weakness; activity-induced cramping, or
  o Family history consistent with X-linked inheritance, OR

• Carrier Screening and Predictive Testing for Presymptomatic/Asymptomatic at Risk Individuals:
  o DMD or BMD diagnosed in 1st or 2nd degree family member and no known mutation at time of testing, AND
  o Family history consistent with X-linked inheritance

DMD Sequencing

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous testing:
  o No mutations detected by deletion/duplication analysis in DMD, and
  o No previous full sequencing analysis of DMD

References


Early Onset Familial Alzheimer Disease

Procedures addressed

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<td>81405</td>
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<tr>
<td>PSEN1 Deletion/Duplication</td>
<td>81479</td>
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<tr>
<td>PSEN1 Known Familial Mutation</td>
<td>81403</td>
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<tr>
<td>APP Sequencing</td>
<td>81406</td>
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<tr>
<td>APP Deletion/Duplication</td>
<td>81479</td>
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<tr>
<td>APP Known Familial Mutation</td>
<td>81403</td>
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<tr>
<td>PSEN2 Sequencing</td>
<td>81406</td>
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<tr>
<td>PSEN2 Known Familial Mutation</td>
<td>81403</td>
</tr>
</tbody>
</table>

What is early onset Alzheimer disease

Definition

Alzheimer disease (AD) is characterized by adult onset, progressive dementia with cerebral cortical atrophy and beta amyloid plaque formation.\(^1\) Common findings include memory loss, confusion, speech issues, hallucinations, and personality and behavioral changes such as poor judgment, agitation, and withdrawal.\(^2,3\) Symptoms of Alzheimer disease usually start after 60-65 years old.\(^2\)

- Of all people with Alzheimer disease, about 25% have at least two affected relatives (called "familial Alzheimer disease"). Most familial Alzheimer disease is late-onset, but in about 5% of cases symptoms start at an unusually young age (called "early onset familial Alzheimer disease" or EOFAD).\(^2\)
- EOFAD is suspected when:\(^1\)
  - More than one family member has Alzheimer disease.
Symptoms consistently start before 65 and often before 55.

Genetics

- Table 1 below summarizes three subtypes of EOFAD. While not clinically distinguishable, the underlying genetic cause differs. Among families with EOFAD, 40-80% will have a detectable mutation in the APP, PSEN1, or PSEN2 gene. Therefore, some families with EOFAD will not have an identifiable mutation by current testing. There may be other disease causing genes that have not been identified to date.
- EOFAD is inherited in an autosomal dominant fashion.
- A person who is found to have a mutation in one of the genes known to cause EOFAD has a 50% chance to pass the mutation to his/her children.
- Most people with EOFAD have an affected parent. In cases where there appears to be no parent affected, most people have a second degree relative with the condition. De novo (new) mutations are possible. However, they have not been reported in EOFAD.
- The presence of a mutation in the PSEN1 gene has complete penetrance by the age of 65, meaning that when a mutation is present symptoms present by this age. Mutations in PSEN2 have a penetrance of approximately 95%. The penetrance of APP is unknown.

Table 1

<table>
<thead>
<tr>
<th>EOFAD type D</th>
<th>Gene</th>
<th>Proportion of EOFAD cases</th>
<th>Average age of onset</th>
<th>Likelihood of symptoms with a mutation</th>
</tr>
</thead>
<tbody>
<tr>
<td>Type 1</td>
<td>APP</td>
<td>No more than 10% to 15%</td>
<td>40's to 50's (occasionally 60s)</td>
<td>Unknown</td>
</tr>
<tr>
<td>Type 3</td>
<td>PSEN1</td>
<td>30% to 70%</td>
<td>40 to early 50's</td>
<td>~100% by 65</td>
</tr>
<tr>
<td>Type 4</td>
<td>PSEN2</td>
<td>Less than 5%</td>
<td>40 to 75</td>
<td>~95% by 80s</td>
</tr>
</tbody>
</table>

Test information

- EOFAD is clinically diagnosed based on family history and age of onset. Genetic testing can confirm a clinical diagnosis in symptomatic individuals. Positive results also allow reliable presymptomatic predictive testing for at-risk family members. Sequence analysis is necessary for acceptable detection rates. Experts suggest that testing should start with PSEN1 sequencing and deletion/duplication analysis, as PSEN1 is the most commonly involved gene.
• Sequence analysis is available for each gene individually or as panel. In addition to sequencing, APP gene testing includes specialized deletion/duplication studies that explain a small percentage of cases.

• APP gene duplication FISH studies may be available as a separate test, but this test alone has limited clinical application.

• Once the disease-causing mutation is identified, predictive testing of adult first-degree relatives (primarily siblings and adult offspring) may be considered. The detection rate for a known familial mutation is greater than 99%. Because of the implications of predictive testing, “Those seeking testing should be counseled about possible problems that they may encounter with regard to health, life, and disability insurance coverage, employment and educational discrimination, and changes in social and family interaction.”

Guidelines and evidence

• The Amyloid Imaging Task Force, Society of Nuclear Medicine and Molecular Imaging, and the Alzheimer’s Association (2013) reference genetic testing in their recommendations:
  o "The use of amyloid PET in lieu of genotyping for suspected autosomal dominant mutation carriers is considered inappropriate. The optimal clinical evaluation in these cases is careful collection of a family history, followed (if appropriate) by genetic counseling prior to and after genetic testing for known mutations. Future use of amyloid PET in autosomal dominant mutation carriers could include determination of whether the amyloid deposition phase of their illness has begun. In the setting of a complete clinical evaluation, including serial neuropsychological testing, this information may be useful in identifying one disease-related milestone that, along with the genetic information, aids decision making."

• A 2012 expert-authored review states that:
  o “EOFAD is diagnosed in families with multiple affected individuals with mean age of onset before 65 years and/or with a documented disease-causing mutation in one of the genes known to be associated with EOFAD.”
  o “Establishing the diagnosis in a proband requires molecular genetic testing to identify a disease-causing mutation in one of the three genes known to be associated with EOFAD.”
  o “Predictive testing for at-risk asymptomatic adult family members requires prior identification of the disease-causing mutation in the family.”

• American College of Medical Genetics and The National Society of Genetic Counselors (2011):
  o “Testing for genes associated with early-onset autosomal dominant AD should be offered in the following situations:”
• “A symptomatic individual with EOAD in the setting of a family history of dementia or in the setting of an unknown family history (e.g., adoption).”
• “Autosomal dominant family history of dementia with one or more cases of EOAD.”
• “A relative with a mutation consistent with EOAD.”

The European Federation of Neurological Societies (2010)⁶ Alzheimer’s diagnosis and management guidelines address genetic testing: “Screening for known pathogenic mutations can be undertaken in patients with appropriate phenotype or a family history of an autosomal dominant dementia.” (No evidence level assigned.) They add, “Testing of patients with familial dementia and of unaffected at-risk-relatives should be accompanied by neurogenetic counseling and undertaken only after full consent and by specialist centres. Pre-symptomatic testing may be performed in at risk member of family-carrying mutation. It is recommended that the Huntington’s disease protocol is followed for pre-symptomatic testing.”

Criteria

PSEN1, PSEN2, or APP known familial mutation testing

• Clinical Consultation:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous genetic testing of PSEN1, PSEN2, or APP, and
  o PSEN1, PSEN2, or APP mutation identified in a 1ˢᵗ  or 2ⁿᵈ degree biological relative, AND
• Diagnostic Testing for Symptomatic Individuals:
  o Dementia diagnosed ≤65 years of age, OR
• Predictive Testing
  o Age 18 years or older, and
  o No previous genetic testing of PSEN1, PSEN2, or APP, and
  o PSEN1, PSEN2, or APP mutation identified in 1ˢᵗ  or 2ⁿᵈ degree biological relative

PSEN1 full sequence and deletion/duplication analysis

• Clinical Consultation:
Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  - No previous PSEN1 sequencing or deletion/duplication analysis, and
  - No known PSEN1, PSEN2, or APP mutation in the family, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Dementia diagnosed ≤65 years of age, and
  - Family history of dementia in 1st or 2nd degree relative

**APP sequence and deletion/duplication analysis**

- Criteria for PSEN1 analysis are met, AND
- No previous genetic testing for APP, AND
- No mutations detected in PSEN1 analysis

**PSEN2 full sequence analysis**

- Criteria for PSEN1 analysis are met, AND
- No previous genetic testing for PSEN2, AND
- No mutations detected in PSEN1 or APP analysis

**References**


EGFR Testing for Non-Small Cell Lung Cancer TKI Response

Procedures addressed

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What is EGFR testing in non-small cell lung cancer

Definition

Non-small cell lung cancer (NSCLC) is the most common type of lung cancer, and is associated with exposure to cigarette smoking.1

- About 80-85% of NSCLC tumors express the epidermal growth factor receptor (EGFR).1 EGFR is a cell surface receptor that causes activation of the intracellular tyrosine kinase domain. Overexpression of EGFR results in increased proliferation and survival of cells, leading to the growth of tumors.1

- The drugs erlotinib (Tarceva®), afatinib (Gilotrif®), and gefitinib (Iressa®) are used in the treatment of people with advanced NSCLC.1 These drugs are tyrosine kinase inhibitors (TKIs). They directly inhibit the EGFR pathway by binding to the epidermal growth factor receptor and blocking downstream signaling resulting in reduced tumor growth.1,2

  - People can develop resistance to erlotinib, afatinib and gefitinib therapy through the development of an EGFR T790M mutation. A third-generation TKI, osimertinib (Tagrisso®), can be used to treat people who have developed this specific mutation.3

- The presence of a mutation in a specific region of the EGFR gene is associated with positive response to TKIs. About 10-15% of Caucasian and up to 40% of Asian NSCLC patients have mutations in EGFR. Mutations occur more often in patients with adenocarcinoma, women, and patients who never smoked.1,2,4

- Testing an NSCLC patient for EGFR mutations can be helpful to select patients who are more likely to respond to TKI therapy.1

  - Patients with either small deletions in exon 19 or a point mutation in exon 21 (L858R) of the tyrosine kinase domain of the EGFR gene are considered good candidates for treatment with erlotinib, afatinib, or gefitinib. Patients found to be
wild type are unlikely to respond to erlotinib, afatinib, or gefitinib and there is insufficient evidence at this time to support their use in individuals with other exon mutations. Other treatment options should be considered for these patients.\textsuperscript{1,5-7}

- Patients who develop an EGFR T790M mutation become resistant to treatment with the first- and second-generation TKIs erlotinib, afatinib and gefitinib. This mutation frequently develops after a median of 9 to 13 months of initial TKI therapy and may respond to treatment with the third-generation TKI osimertinib.\textsuperscript{3,8}

- EGFR is upstream from another gene, KRAS, in the signaling pathway. Overlapping EGFR mutations and KRAS mutations occur in <1% of patients with lung cancer: patients with NSCLC may have an EGFR mutation or a KRAS mutation, but co-occurrence of both mutations is very rare.\textsuperscript{1}

**Test information**

- Targeted analysis of the EGFR gene can be performed by two different methods:
  - **Mutation panels** check specifically for the two most common EGFR mutations, E19del and L858R. These mutations account for up to 85% of all EGFR mutations.\textsuperscript{1}
  - **Sequencing of specific exons (18-21)** will find any mutation in the region (tyrosine kinase domain). The E19del mutation is located on exon 19, the T790M mutation is located on exon 20 and the L858R mutation is located on exon 21.\textsuperscript{1,8}

- Testing by either method is sensitive and accurate,\textsuperscript{1} and both methods are commonly used by commercial laboratories doing testing.

- EGFR activity can also be measured by fluorescence in situ hybridization (FISH) and immunohistochemistry (IHC) though targeted mutation analysis is the more routinely recommended test.\textsuperscript{1,2}

**Guidelines and evidence**

- The National Comprehensive Cancer Network (NCCN, 2018) guidelines recommend:\textsuperscript{1}
  - “...the NCCN Panel recommends that biomarker testing be done as part of broad molecular profiling using a validated test(s) that assesses a minimum of the following potential genetic alterations: EGFR mutations, BRAF mutations, ALK rearrangements, and ROS1 rearrangements."
  - “Testing for ALK gene rearrangements and EGFR mutations is recommended (category 1 for both) in the NSCLC algorithm for patients with nonsquamous..."
NSCLC or NSCLC not otherwise specified (NOS) so that patients with these genetic abnormalities can receive effective treatment with targeted agents.”

- “The predictive effects of the drug-sensitive EGFR mutations are well defined. Patients with these mutations have a significantly better response to erlotinib, gefitinib, afatinib, or osimertinib.”

- “Data suggests that patients without sensitizing EGFR mutations should not be treated with EGFR TKIs in any line of therapy.”

- “Although rare, patients with ALK rearrangements or EGFR mutations can have mixed squamous histology. Therefore, testing for ALK rearrangements and EGFR mutations can be considered in select patients with squamous cell histology if they are never smokers, small biopsy specimens were used for testing, or mixed histology was reported. EGFR, KRAS, ROS1, and ALK genetic alterations do not usually overlap.”

- “In patients with squamous cell carcinoma, the observed incidence of EGFR mutations is 2.7% with a confidence that the true incidence of mutations is less than 3.6%. This frequency of EGFR mutations does not justify routine testing of all tumor specimens.”

- “Thus the NCCN Panel strongly advises broader molecular profiling (also know as precision medicine) to identify rare driver mutations to ensure that patients receive the most appropriate treatment; patients may be eligible for clinical trials for some of these targeted agents.”

- The National Comprehensive Cancer Network (NCCN, 2018) states the following in regards to liquid biopsy testing for EGFR T790M (acquired resistance to EGFR TKI therapy) in patients with non-small cell lung cancer:¹

  - In patients with a sensitizing EGFR mutation at progression, tissue biopsy testing for EGFR T790M testing should be performed.

  - “Data suggest that plasma genotyping (also known as liquid biopsy or plasma biopsy) may be considered instead of tissue biopsy to detect whether patients have T790M; however, if the plasma biopsy is negative, then tissue biopsy is recommended if feasible.”

- The College of American Pathologists, International Association for the Study of Lung Cancer, and Association for Molecular Pathology (CAP/IASLC/AMP, 2013) Guidelines state:⁹

  - “EGFR molecular testing should be used to select patients for EGFR-targeted TKI therapy, and patients with lung adenocarcinoma should not be excluded from testing on the basis of clinical characteristics.”

  - “EGFR and ALK testing is recommended for adenocarcinomas and mixed lung cancers with an adenocarcinoma component, regardless of histologic grade. In the setting of fully excised lung cancer specimens, EGFR and ALK testing is not recommended in lung cancers that lack any adenocarcinoma component…”
• “For patients with multiple, apparently separate, primary lung adenocarcinomas, each tumor may be tested but testing of multiple different areas within a single tumor is not necessary.”

• “In the setting of more limited lung cancer specimens (eg, biopsies, cytology) where an adenocarcinoma component cannot be completely excluded, EGFR and ALK testing may be performed in cases showing squamous cell histology. Clinical criteria (eg, young age, lack of smoking history) may be useful to select a subset of these samples for testing.”

The American Society of Clinical Oncology (ASCO, 2014) Endorsement of the College of American Pathologists/International Association for the Study of Lung Cancer/Association for Molecular Pathology Guideline states that:

• “The ASCO ad hoc panel reviewed the CAP/IASLC/AMP guideline and concurs that the recommendations are clear, thorough, based on the most relevant scientific evidence in this content area, and present options that will be acceptable to patients, clinicians, and pathologists.”

• “The ASCO review panel endorses the CAP/IASLC/AMP guideline. This guideline represents an important advance toward standardization of EGFR and ALK testing practices and is of major clinical relevance in advancing the care of patients with lung cancer.”

• EGFR is listed as an FDA-approved biomarker for erlotinib, afatinib, gefitinib, and osimertinib.

• Product labeling for erlotinib, afatinib, and gefitinib address EGFR testing. All three products are listed as “first line treatments for patients with metastatic non-small cell lung cancer whose tumors have epidermal growth factor receptor (EGFR) exon 19 deletions or exon 21 (L858R) substitution mutations as detected by an FDA-approved test.”

• Product labeling for osimertinib notes the product is “a kinase inhibitor indicated for the treatment of patients with metastatic epidermal growth factor receptor (EGFR) T790M mutation-positive non-small cell lung cancer (NSCLC), as detected by an FDA-approved test, whose disease has progressed on or after EGFR TKI therapy.”

Criteria

EGFR targeted mutation testing is indicated in individuals with metastatic non-small cell lung cancer of nonsquamous cell type prior to initiation of treatment with erlotinib, afatinib, or gefitinib therapy.

For patients whose disease progresses either on or after TKI therapy, repeat EGFR testing to identify the emergence of a T790M mutation may be considered to determine whether further treatment with osimertinib would be indicated.
Exceptions and other considerations

Analysis for other mutations within exons 18-24, or other applications related to NSCLC, is considered experimental/investigational because the safety and/or effectiveness of this service cannot be established by review of the available published peer-reviewed literature.

References

10. FDA. Table of valid genomic biomarkers in the context of approved drug labels. Available at
http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm.
Ehlers-Danlos Syndrome Testing

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Known Familial Mutation Analysis</td>
<td>81403</td>
</tr>
<tr>
<td>ADAMTS2 Sequencing</td>
<td>81479</td>
</tr>
<tr>
<td>ADAMTS2 Deletion/Duplication Analysis</td>
<td>81479</td>
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<td>B3GALT6 Sequencing</td>
<td>81479</td>
</tr>
<tr>
<td>B3GALT6 Deletion/Duplication Analysis</td>
<td>81479</td>
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<tr>
<td>B4GALT7 Sequencing</td>
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<tr>
<td>B4GALT7 Deletion/Duplication Analysis</td>
<td>81479</td>
</tr>
<tr>
<td>C1R Sequencing</td>
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<tr>
<td>C1R Deletion/Duplication Analysis</td>
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</tr>
<tr>
<td>C1S Sequencing</td>
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<tr>
<td>C1S Deletion/Duplication Analysis</td>
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<td>CHST14 Sequencing</td>
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<td>CHST14 Deletion/Duplication Analysis</td>
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<tr>
<td>COL1A1 Sequencing</td>
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<td>COL1A1 Deletion/Duplication Analysis</td>
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<td>COL1A2 Sequencing</td>
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<tr>
<td>COL1A2 Deletion/Duplication Analysis</td>
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<tr>
<td>COL12A1 Sequencing</td>
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<td>COL12A1 Deletion/Duplication Analysis</td>
<td>81479</td>
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<td>COL3A1 Sequencing</td>
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<tr>
<td>COL3A1 Deletion/Duplication Analysis</td>
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<tr>
<td>COL5A1 Sequencing</td>
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<tr>
<td>COL5A1 Deletion/Duplication Analysis</td>
<td>81479</td>
</tr>
<tr>
<td>COL5A2 Sequencing</td>
<td>81479</td>
</tr>
</tbody>
</table>
What is Ehlers-Danlos Syndrome

Definition

Ehlers-Danlos syndrome (EDS) is a heterogeneous group of connective tissue disorders. Although all types of EDS affect the joints and skin, additional features vary by type.¹

- An unusually large range of joint movement (hypermobility) occurs with most forms of EDS, and is especially prominent in the hypermobile type.¹
  - Joint hypermobility is typically assessed using a 9-point scale called the Beighton criteria. Adults with a Beighton score of ≥5, and children with a Beighton score ≥6, are considered to have generalized joint hypermobility.²³
  - Generalized joint hypermobility is relatively common, occurring in 2-57% of different populations.²
  - Joint hypermobility can be a feature of other connective tissue disorders (e.g. Marfan syndrome, skeletal dysplasias, and other disorders), myopathic disorders, and other chromosomal and molecular disorders. Joint hypermobility may also occur as an isolated, nonsyndromic finding.³
Joint hypermobility may be asymptomatic, or associated with musculoskeletal complications such as chronic pain and disturbed proprioception. Individuals with symptomatic joint hypermobility who do not have hypermobile EDS or another causative genetic disorder are considered to have “hypermobility spectrum disorders (HSDs).”

- The combined prevalence of all types of EDS may be about 1 in 5,000 individuals worldwide, with the most common being the hypermobile type.
- Six types of EDS were originally delineated in 1997. In 2017, clinical criteria were updated and revised to include thirteen EDS types:
  - Classical EDS (formerly types I/II)
  - Classical-like EDS (formerly tenascin X deficiency EDS)
  - Cardiac-valvular EDS
  - Vascular EDS (formerly type IV)
  - Hypermobile EDS (formerly type III; “joint hypermobility syndrome” is also now considered to be the same clinical entity as hypermobile EDS)
  - Arthrochalasia EDS (formerly types VIIA and VIIIB)
  - Dermatosparaxis EDS (formerly type VIIC)
  - Kyphoscoliotic EDS (formerly type VIA)
  - Brittle cornea syndrome (formerly type VIB)
  - Spondylodysplastic EDS
  - Musculocontractural type (formerly type VIB)
  - Myopathic EDS
  - Periodontal EDS (formerly type VIII)

**Genetics of Ehlers-Danlos Syndrome**

Genetics of EDS (summarized in the table below):

- Some EDS types follow an autosomal dominant pattern, meaning only one mutation is required to cause disease. In these cases, children of an affected person have a 50% chance of inheriting the disease-causing mutation.
- Other types are autosomal recessive. Two mutations are required to cause recessive types, and usually only siblings are affected. There is no parent-to-child transmission.

<table>
<thead>
<tr>
<th>EDS Type</th>
<th>Inheritance</th>
<th>Genetic basis</th>
<th>Protein</th>
</tr>
</thead>
<tbody>
<tr>
<td>Classical EDS</td>
<td>Autosomal dominant</td>
<td>Major: COL5A1, COL5A2</td>
<td>Type V collagen</td>
</tr>
</tbody>
</table>

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<table>
<thead>
<tr>
<th>EDS Type</th>
<th>Inheritance</th>
<th>Genetic basis</th>
<th>Protein</th>
</tr>
</thead>
<tbody>
<tr>
<td>Classical-like EDS</td>
<td>Autosomal recessive</td>
<td>COL1A2 (biallelic mutations that lead to COL1A2 NMD &amp; absence of pro α2(I) collagen chains)</td>
<td>Type I collagen</td>
</tr>
<tr>
<td>Cardiac valvular EDS</td>
<td>Autosomal recessive</td>
<td>COL1A2 (biallelic mutations that lead to COL1A2 NMD &amp; absence of pro α2(I) collagen chains)</td>
<td>Type I collagen</td>
</tr>
<tr>
<td>Vascular EDS</td>
<td>Autosomal dominant</td>
<td>Major: COL3A1 Rare: COL1A1 c.934C&gt;T, c.1720C&gt;T, c.3227C&gt;T</td>
<td>Type III collagen Type I collagen</td>
</tr>
<tr>
<td>Hypermobile EDS</td>
<td>Autosomal dominant</td>
<td>Unknown</td>
<td>Unknown</td>
</tr>
<tr>
<td>Arthrochalasia EDS</td>
<td>Autosomal dominant</td>
<td>COL1A1 COL1A2</td>
<td>Type I collagen</td>
</tr>
<tr>
<td>Dermatosparaxis EDS</td>
<td>Autosomal recessive</td>
<td>ADAMTS2</td>
<td>ADAMTS-2</td>
</tr>
<tr>
<td>Kyphoscoliotic EDS</td>
<td>Autosomal recessive</td>
<td>PLOD1 FKBP14</td>
<td>LH1 FKBP22</td>
</tr>
<tr>
<td>Brittle cornea syndrome</td>
<td>Autosomal recessive</td>
<td>ZNF469 PRDM5</td>
<td>ZNF469 PRDM5</td>
</tr>
<tr>
<td>Spondylodysplastic EDS</td>
<td>Autosomal recessive</td>
<td>B4GALT7 B3GALT6 SLC9A13</td>
<td>β4GalT7 β3GalT6 ZIP13</td>
</tr>
<tr>
<td>Musculocontractural EDS</td>
<td>Autosomal recessive</td>
<td>CHST14 DSE</td>
<td>D4ST1 DSE</td>
</tr>
<tr>
<td>Myopathic EDS</td>
<td>Autosomal recessive or dominant</td>
<td>COL12A1</td>
<td>Type XII collagen</td>
</tr>
<tr>
<td>Periodontal type</td>
<td>Autosomal dominant</td>
<td>C1R C1S</td>
<td>C1r C1s</td>
</tr>
</tbody>
</table>
Test information

- Clinical genetic testing is available for most types of EDS (see table above), and is used to confirm the final diagnosis when it is clinically suspected.\(^5\)
  - Hypermobile EDS (hEDS) continues to require a clinical diagnosis, since the genetic etiology of this type is not yet known.\(^5\)

- **Single gene analysis** — EDS genetic testing may be performed with Sanger sequencing or next generation sequencing (NGS). If no mutations are identified (or, in the case of an autosomal recessive EDS, only one mutation is identified), deletion/duplication analysis may be considered. Mutation detection rates vary by type:
  - >90% of individuals with classical EDS have a mutation in COL5A1 or COL5A2.\(^5\)
  - >95% of individuals with vascular EDS have a mutation in COL3A1.\(^6\)
  - Mutation detection rates for the rarer EDS types are mostly unknown.

- **Multi-gene panel testing** — With the availability of NGS technology, EDS genetic testing is increasingly performed as a panel test that includes multiple EDS genes. In addition, these panels often include other hereditary connective tissue disorders with overlapping phenotypes. Panel testing is addressed in the guideline: *Hereditary Connective Tissue Disorder Testing*.

Guidelines and evidence

- An expert-authored review (updated in 2016)\(^7\) states the following regarding hEDS: “If a patient’s personal or family history is suggestive of one of the other types of EDS or another hereditary disorder of connective tissue or arterial fragility syndrome, analysis of an associated gene or multi-gene connective tissue disease panel may be appropriate. Failure to identify a pathogenic variant with such multiple gene testing reduces the likelihood of an arterial fragility syndrome, but does not completely rule it out, especially in the setting of a positive personal or family history of arterial fragility. Negative testing for an arterial fragility syndrome also does not confirm a diagnosis of EDS, hypermobility type. Therefore, such testing is not recommended in the absence of specific suggestive signs, symptoms, or family history.”

- According to the International Consortium on the Ehlers-Danlos Syndromes (2017):\(^5\)
  - “In view of the vast genetic heterogeneity and phenotypic variability of the EDS subtypes, and the clinical overlap between many of these subtypes, but also with other hereditary connective tissue disorders, the definite diagnosis relies for all subtypes, except hEDS, on molecular confirmation with identification of (a) causative variant(s) in the respective gene.”
Molecular diagnostic strategies should rely on NGS technologies, which offer the potential for parallel sequencing of multiple genes. Targeted resequencing of a panel of genes is a time- and cost-effective approach for the molecular diagnosis of the genetically heterogeneous EDS. When no mutation (or in case of an autosomal recessive condition only one mutation) is identified, this approach should be complemented with a copy number variant (CNV) detection strategy to identify large deletions or duplications, for example Multiplex Ligation-dependent Probe Amplification (MLPA), qPCR, or targeted array analysis.

The diagnosis of hEDS remains clinical as there is yet no reliable or appreciable genetic etiology to test for in the vast majority of patients.

2017 International Criteria for Classical EDS

Minimal criteria suggestive for Classical EDS (cEDS):

- Major criterion 1, PLUS either:
  - Major criterion 2, and/or
  - At least three minor criteria.

<table>
<thead>
<tr>
<th>Major criteria for cEDS</th>
<th>Minor criteria for cEDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Skin hyperextensibility and atrophic scarring</td>
<td>1. Easy bruising</td>
</tr>
<tr>
<td>2. Generalized joint hypermobility</td>
<td>2. Soft, doughy skin</td>
</tr>
<tr>
<td></td>
<td>3. Skin fragility (or traumatic splitting)</td>
</tr>
<tr>
<td></td>
<td>4. Molluscoid pseudotumors</td>
</tr>
<tr>
<td></td>
<td>5. Subcutaneous spheroids</td>
</tr>
<tr>
<td></td>
<td>6. Hernia (or history thereof)</td>
</tr>
<tr>
<td></td>
<td>7. Epicanthal folds</td>
</tr>
<tr>
<td></td>
<td>8. Complications of joint hypermobility (e.g., sprains, luxation/subluxation, pain, flexible flatfoot)</td>
</tr>
<tr>
<td></td>
<td>9. Family history of a first-degree relative who meets clinical criteria</td>
</tr>
</tbody>
</table>

2017 International Criteria for Classical-like EDS

Minimal criteria suggestive for Classical-like EDS (clEDS):

- All three major criteria, AND
• A family history compatible with autosomal recessive transmission.

<table>
<thead>
<tr>
<th>Major criteria for cIEDS</th>
<th>Minor criteria for cIEDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Skin hyperextensibility, with velvety skin texture and absence of atrophic scarring</td>
<td>1. Foot deformities: broad/plump forefoot, brachydactyly with excessive skin; pes planus; hallux valgus; piezogenic papules</td>
</tr>
<tr>
<td>2. Generalized joint hypermobility with or without recurrent dislocations (most commonly shoulder and ankle)</td>
<td>2. Edema in the legs in absence of cardiac failure</td>
</tr>
<tr>
<td>4. Axonal polyneuropathy</td>
<td>5. Atrophy of muscles in hands and feet</td>
</tr>
<tr>
<td>6. Acrogeric hands, mallet finger(s), clinodactyly, brachydactyly</td>
<td>7. Vaginal/uterus/rectal prolapse</td>
</tr>
</tbody>
</table>

2017 International Criteria for Cardiac-Valvular EDS

Minimal criteria suggestive for Cardiac-Valvular EDS (cvEDS)

• Major criterion 1, AND

• A family history compatible with autosomal recessive inheritance, PLUS either:
  o One other major criterion, and/or
  o At least two minor criteria.

<table>
<thead>
<tr>
<th>Major criteria for cvEDS</th>
<th>Minor criteria for cvEDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Severe progressive cardiac-valvular problems (aortic valve, mitral valve)</td>
<td>1. Inguinal hernia</td>
</tr>
<tr>
<td>2. Skin involvement: skin hyperextensibility, atrophic scars, thin skin, easy bruising</td>
<td>2. Pectus deformity (especially pectus excavatum)</td>
</tr>
<tr>
<td>3. Joint hypermobility (generalized or restricted to small joints)</td>
<td>3. Joint dislocations</td>
</tr>
<tr>
<td>4. Axonal polyneuropathy</td>
<td>4. Foot deformities: pes planus, pes planovalgus, hallux valgus</td>
</tr>
</tbody>
</table>

2017 International Criteria for Vascular EDS

Minimal criteria suggestive for Vascular EDS (vEDS):

• A family history of the disorder, and/or
• Arterial rupture or dissection in individuals less than 40 years of age, and/or
• Spontaneous pneumothorax in the presence of other features consistent with vEDS, and/or
• A combination of the other minor clinical features listed below.

<table>
<thead>
<tr>
<th>Major criteria for vEDS</th>
<th>Minor criteria for vEDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Family history of vEDS with documented causative variant in COL3A1</td>
<td>1. Bruising unrelated to identified trauma and/or in unusual sites such as cheeks and back</td>
</tr>
<tr>
<td>2. Arterial rupture at a young age</td>
<td>2. Thin, translucent skin with increased venous visibility</td>
</tr>
<tr>
<td>3. Spontaneous sigmoid colon perforation in the absence of known diverticular disease or other bowel pathology</td>
<td>3. Characteristic facial appearance</td>
</tr>
<tr>
<td>4. Uterine rupture during the third trimester in the absence of previous C-section and/or severe peripartum perineum tears</td>
<td>4. Spontaneous pneumothorax</td>
</tr>
<tr>
<td>5. Carotid-cavernous sinus fistula (CCSF) formation in the absence of trauma</td>
<td>5. Acrogeria</td>
</tr>
<tr>
<td>6. Talipes equinovarus</td>
<td>6. Talipes equinovarus</td>
</tr>
<tr>
<td>12. Early onset varicose veins (under 30 and nulliparous if female)</td>
<td>12. Early onset varicose veins (under 30 and nulliparous if female)</td>
</tr>
</tbody>
</table>

2017 International Criteria for Hypermobile EDS

Diagnosis of Hypermobile EDS (hEDS) requires the simultaneous presence of criteria 1 AND 2 AND 3:
• Criteria 1: Generalized joint hypermobility
• Criterion 2: Two or more among the features (A-C) listed in the table below must be present (for example: A and B; A and C; B and C; A and B and C).
• Criterion 3: All of the following prerequisites must be met:
  o Absence of unusual skin fragility, and
  o Exclusion of other heritable and acquired connective tissue disorders, including autoimmune rheumatologic conditions, and
- Exclusion of alternative diagnoses that may also include joint hypermobility by means of hypotonia and/or connective tissue laxity.

<table>
<thead>
<tr>
<th>Feature A</th>
<th>Feature B</th>
<th>Feature C</th>
</tr>
</thead>
<tbody>
<tr>
<td>A total of 5 must be present:</td>
<td>Positive family history, with one or more first degree relatives independently meeting the current diagnostic criteria for hEDS.</td>
<td>Must have at least one</td>
</tr>
<tr>
<td>1. Unusually soft or velvety skin</td>
<td></td>
<td>1. Musculoskeletal pain in two or more limbs, recurring daily for at least 3 months.</td>
</tr>
<tr>
<td>2. Mild skin hyperextensibility</td>
<td></td>
<td>2. Chronic, widespread pain for ≥ 3 months</td>
</tr>
<tr>
<td>3. Unexplained striae</td>
<td></td>
<td>3. Recurrent joint dislocations or frank joint instability, in the absence of trauma:</td>
</tr>
<tr>
<td>4. Bilateral piezogenic papules of the heel</td>
<td></td>
<td>a. Three or more atraumatic dislocations in the same joint or two or more atraumatic dislocations in two different joints occurring at different times, or</td>
</tr>
<tr>
<td>5. Recurrent or multiple abdominal hernia(s)</td>
<td></td>
<td>b. Medical confirmation of joint instability at two or more sites not related to trauma</td>
</tr>
<tr>
<td>6. Atrophic scarring involving at least two sites</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7. Pelvic floor, rectal, and/or uterine prolapses in children, men or nulliparous women without a history of morbid obesity or other known predisposing medical condition</td>
<td></td>
<td></td>
</tr>
<tr>
<td>8. Dental crowding and high or narrow palate</td>
<td></td>
<td></td>
</tr>
<tr>
<td>9. Arachnodactyly</td>
<td></td>
<td></td>
</tr>
<tr>
<td>10. Arm span-to-height ≥ 1.05</td>
<td></td>
<td></td>
</tr>
<tr>
<td>11. Mitral valve prolapse (MVP)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>12. Aortic root dilatation with Z-score &gt; +2</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**2017 International Criteria for Arthrochalasia EDS**

Minimal criteria suggestive for Arthrochalasia EDS (aEDS):

- Major criterion 1, PLUS either:
Major criteria for aEDS | Minor criteria for aEDS
---|---
1. Congenital hip dislocation | 1. Muscle hypotonia
2. Severe generalized joint hypermobility, with multiple dislocations/subluxations | 2. Kyphoscoliosis
3. Skin hyperextensibility | 3. Radiologically mild osteopenia
| 4. Tissue fragility, including atrophic scars
| 5. Easy bruisable skin

### 2017 International Criteria for Dermatosparaxis EDS

Minimal criteria suggestive for Dermatosparaxis EDS (dEDS):

- Major criterion 1, AND
- Major criterion 2, PLUS either:
  - One other major criterion, and/or
  - Three minor criteria.

#### Major criteria for dEDS | Minor criteria for dEDS
---|---
1. Extreme skin fragility with congenital or postnatal skin tears | 1. Soft and doughy skin texture
2. Characteristic craniofacial features, which are evident at birth or early infancy, or evolve later in childhood | 2. Skin hyperextensibility
3. Redundant, almost lax skin, with excessive skin folds at the wrist and ankles | 3. Atrophic scars
4. Increased palmar wrinkling | 4. Generalized joint hypermobility
5. Severe bruisability with a risk of subcutaneous hematomas and hemorrhage | 5. Complications of visceral fragility (e.g., bladder rupture, diaphragmatic rupture, rectal prolapse)
6. Umbilical hernia | 6. Delayed motor development
7. Postnatal growth retardation | 7. Osteopenia
8. Short limbs, hands and feet | 8. Hirsutism
9. Perinatal complications due to connective tissue fragility | 9. Tooth abnormalities
10. Refractive errors (myopia, astigmatism) | 11. Strabismus

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2017 International Criteria for Kyphoscoliotic EDS

Minimal criteria suggestive for Kyphoscoliotic EDS (kEDS):

- Major criterion 1, AND
- Major criterion 2, PLUS either:
  - Major criterion 3, and/or
  - Three minor criteria (either general or gene-specific criteria).

### Major criteria for kEDS

1. Congenital muscle hypotonia
2. Congenital or early onset kyphoscoliosis (progressive or non-progressive)
3. Generalized joint hypermobility with dislocations/subluxations (shoulders, hips, and knees in particular)

### Minor criteria for kEDS

1. Skin hyperextensibility
2. Easy bruised skin
3. Rupture/aneurysm of a medium-sized artery
4. Osteopenia/osteoporosis
5. Blue sclerae
6. Hernia (umbilical or inguinal)
7. Pectus deformity
8. Marfanoid habitus
9. Talipes equinovarus
10. Refractive errors (myopia, hypermetropia)

### Gene-specific minor criteria for kEDS

- PLOD1
  1. Skin fragility
  2. Scleral and ocular fragility/rupture
  3. Microcornea
  4. Facial dysmorphology
- FKBP14
  1. Congenital hearing impairment (any type)
  2. Follicular hyperkeratosis
  3. Muscle atrophy
  4. Bladder diverticula

2017 International Criteria for Brittle Cornea Syndrome

Minimal criteria suggestive for Brittle Cornea Syndrome (BCS):

- Major criterion 1, PLUS either:
  - At least one other major criterion, and/or
  - Three minor criteria.

### Major criteria for BCS

1. Thin cornea, with or without rupture (central corneal thickness often <400 µm)

### Minor criteria for BCS

1. Enucleation or corneal scarring as a result of previous rupture
2. Progressive loss of corneal stromal
### 2017 International Criteria for Spondylodysplastic EDS

Minimal criteria suggestive for Spondylodysplastic EDS (sEDS):

- Major criterion 1, AND
- Major criterion 2, PLUS
- Characteristic radiographic findings and at least 3 other minor criteria (general or type-specific).

<table>
<thead>
<tr>
<th>Major criteria for sEDS</th>
<th>Minor criteria for sEDS</th>
<th>Gene-specific minor criteria for sEDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Short stature</td>
<td>1. Skin hyperextensibility, soft, doughy skin, thin translucent skin</td>
<td></td>
</tr>
<tr>
<td>3. Bowing of limbs</td>
<td>3. Delayed motor development</td>
<td></td>
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<td></td>
<td>4. Osteopenia</td>
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<tr>
<td></td>
<td>B4GALT7</td>
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</tr>
<tr>
<td></td>
<td>1. Radioulnar synostosis</td>
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<td></td>
<td>2. Bilateral elbow contractures or limited elbow movement</td>
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</tr>
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<td></td>
<td>3. Generalized joint hypermobility</td>
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<td></td>
<td>4. Single transverse</td>
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</tr>
<tr>
<td>Major criteria for sEDS</td>
<td>Minor criteria for sEDS</td>
<td>Gene-specific minor criteria for sEDS</td>
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<tr>
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</tr>
<tr>
<td>5. Delayed cognitive development</td>
<td>palmar curve</td>
<td>1. Protuberant eyes with bluish sclerae</td>
</tr>
<tr>
<td></td>
<td>5. Characteristic craniofacial features</td>
<td>2. Hands with finely wrinkled palms</td>
</tr>
<tr>
<td></td>
<td>6. Characteristic radiographic findings</td>
<td>3. Atrophy of the thenar muscles, tapering fingers</td>
</tr>
<tr>
<td></td>
<td>7. Severe hypermetropia</td>
<td>4. Hypermobility of distal joints</td>
</tr>
<tr>
<td></td>
<td>8. Clouded cornea</td>
<td>5. Characteristic radiologic findings</td>
</tr>
<tr>
<td></td>
<td>SLC39A13</td>
<td>B3GALT6</td>
</tr>
<tr>
<td></td>
<td>1. Kyphoscoliosis (congenital or early onset)</td>
<td>1. Kyphoscoliosis (congenital or early onset)</td>
</tr>
<tr>
<td></td>
<td>2. Joint hypermobility</td>
<td>2. Joint hypermobility</td>
</tr>
<tr>
<td></td>
<td>4. Peculiar fingers</td>
<td>4. Peculiar fingers</td>
</tr>
<tr>
<td></td>
<td>5. Talipes equinovarus</td>
<td>5. Talipes equinovarus</td>
</tr>
<tr>
<td></td>
<td>6. Characteristic craniofacial features</td>
<td>6. Characteristic craniofacial features</td>
</tr>
<tr>
<td></td>
<td>7. Tooth discoloration, dysplastic teeth</td>
<td>7. Tooth discoloration, dysplastic teeth</td>
</tr>
<tr>
<td></td>
<td>8. Characteristic radiographic findings</td>
<td>8. Characteristic radiographic findings</td>
</tr>
<tr>
<td>Major criteria for sEDS</td>
<td>Minor criteria for sEDS</td>
<td>Gene-specific minor criteria for sEDS</td>
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<td></td>
<td></td>
<td>fractures Ascending aortic aneurysm</td>
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<td></td>
<td></td>
<td>10. Lung hypoplasia, restrictive lung disease</td>
</tr>
</tbody>
</table>

2017 International Criteria for Musculocontractural EDS

Minimal criteria suggestive for Musculocontractural EDS (mcEDS):

- At birth or in early childhood:
  - Major criterion 1, AND
  - Major criterion 2
- In adolescence and in adulthood:
  - Major criterion 1, AND
  - Major criterion 3.

<table>
<thead>
<tr>
<th>Major criteria for mcEDS</th>
<th>Minor criteria for mcEDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Congenital multiple contractures and/or talipes equinovarus (clubfoot)</td>
<td>1. Recurrent/chronic dislocations</td>
</tr>
<tr>
<td>2. Characteristic craniofacial features, which are evident at birth or in early infancy</td>
<td>2. Pectus deformities (flat, excavated)</td>
</tr>
<tr>
<td>3. Characteristic cutaneous features including skin hyperextensibility, easy bruisability, skin fragility with atrophic scars, increased palmar wrinkling</td>
<td>3. Spinal deformities (scoliosis, kyphoscoliosis)</td>
</tr>
<tr>
<td>4. Peculiar fingers (tapering, slender, cylindrical)</td>
<td>4. Peculiar fingers (tapering, slender, cylindrical)</td>
</tr>
<tr>
<td>5. Progressive talipes deformities</td>
<td>5. Progressive talipes deformities</td>
</tr>
<tr>
<td>6. Large subcutaneous hematomas</td>
<td>6. Large subcutaneous hematomas</td>
</tr>
<tr>
<td>7. Chronic constipation</td>
<td>7. Chronic constipation</td>
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<tr>
<td>8. Colonic diverticula</td>
<td>8. Colonic diverticula</td>
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<tr>
<td>11. Cryptorchidism in males</td>
<td>11. Cryptorchidism in males</td>
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<tr>
<td>12. Strabismus</td>
<td>12. Strabismus</td>
</tr>
<tr>
<td>13. Refractive errors (myopia, astigmatism)</td>
<td>13. Refractive errors (myopia, astigmatism)</td>
</tr>
</tbody>
</table>
2017 International Criteria for Periodontal EDS

Minimal criteria suggestive for Periodontal EDS (pEDS):

- Major criterion 1, OR major criterion 2, PLUS
  - At least two other major criteria and one minor criterion.

<table>
<thead>
<tr>
<th>Major criteria for pEDS</th>
<th>Minor criteria for pEDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Severe and intractable periodontitis of early onset (childhood or adolescence)</td>
<td>1. Easy bruising</td>
</tr>
<tr>
<td>2. Lack of attached gingiva</td>
<td>2. Joint hypermobility, mostly distal joints</td>
</tr>
<tr>
<td>3. Pretibial plaques</td>
<td>3. Skin hyperextensibility and fragility, abnormal scarring (wide or atrophic)</td>
</tr>
<tr>
<td>4. Family history of a first-degree relative who meets clinical criteria</td>
<td>4. Increased rate of infections</td>
</tr>
<tr>
<td></td>
<td>5. Hernias</td>
</tr>
<tr>
<td></td>
<td>6. Marfanoid facial features</td>
</tr>
<tr>
<td></td>
<td>7. Acrogeria</td>
</tr>
<tr>
<td></td>
<td>8. Prominent vasculature</td>
</tr>
</tbody>
</table>

Criteria

EDS Known Familial Mutation Analysis

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous testing of the requested gene, AND
- Diagnostic Testing for an Autosomal Dominant EDS:
  - Known mutation identified in 1st degree biological relative. (Note: 2nd or 3rd degree relatives may be considered when 1st degree relatives are unavailable or unwilling to be tested), OR
- Carrier Screening for an Autosomal Recessive EDS:
  - Known mutation(s) identified in 1st, 2nd, or 3rd degree biologic relative(s), OR
- Prenatal Testing for At-Risk Pregnancies:
Family history of an autosomal dominant type of EDS with a known mutation identified in a previous child or either parent, or

Both parents carry a known mutation for an autosomal recessive type of EDS, AND

Rendering laboratory is a qualified provider of service per the Health Plan policy.

**EDS Gene Sequencing**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous sequencing of the requested gene, AND

- The member does not have a known genetic cause for their symptoms, AND

- The member does not have a family history of a known EDS gene mutation, AND

- The member meets the above 2017 minimal criteria suggestive for an EDS type associated with the requested gene test:
  - For COL5A1 and/or COL5A2 sequencing: criteria for classical EDS met, or
  - For TNXB sequencing: criteria for classical-like EDS met, or
  - For COL1A1* sequencing: criteria met for one of the following EDS types:
    - Classical EDS, or
    - Vascular EDS, or
    - Arthrochalasia EDS, or
    - Member displays one or more of the following:^{5}
      - Arterial rupture at a young age, or
      - Spontaneous sigmoid colon perforation in the absence of known diverticular disease or other bowel pathology, or
      - Uterine rupture during the third trimester in the absence of previous C-section and/or severe peripartum perineum tears, or
      - Carotid-cavernous sinus fistula (CCSF) formation in the absence of trauma, OR
  - For COL1A2* sequencing: criteria met for one of the following EDS types:
    - Cardiac valvular EDS, or
- Arthrochalasia EDS, or
  - For COL3A1* sequencing: criteria for vascular EDS met, or
    - Member displays one or more of the following:\(^5\)
      - Arterial rupture at a young age, or
      - Spontaneous sigmoid colon perforation in the absence of known diverticular disease or other bowel pathology, or
      - Uterine rupture during the third trimester in the absence of previous C-section and/or severe peripartum perineum tears, or
      - Carotid-cavernous sinus fistula (CCSF) formation in the absence of trauma, OR
  - For ADAMTS2 sequencing: criteria for dermatosparaxis EDS met, or
  - For PLOD1 and/or FKBP14 sequencing: criteria for kyphoscoliotic EDS met, or
  - For ZNF469 and/or PRDM5 sequencing: criteria for brittle cornea syndrome met, or
  - For B3GALT6, B4GALT7, and/or SLC39A13 sequencing: criteria for spondylodysplastic EDS met, or
  - For CHST14 and/or DSE sequencing: criteria for musculocontractural EDS met, or
  - For COL12A1 sequencing: criteria for myopathic EDS met, or
  - For C1R and/or C1S sequencing: criteria for periodontal EDS met, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

* For non-EDS indications, refer to any available disorder-specific guidelines or general guidelines, *Hereditary Connective Tissue Disorder Testing* or *Genetic Testing for Non-Cancer Conditions*, as appropriate. COL1A1 and COL1A2 are also associated with osteogenesis imperfecta, Caffey disease, and skeletal dysplasias. COL3A1 is also associated with familial thoracic aortic aneurysm and dissection (TAAD).

Panel testing is addressed in the guideline: *Hereditary Connective Tissue Disorder Testing*.

**EDS Gene Deletion/Duplication Analysis**

- The member meets above criteria for sequencing of the requested gene, AND
- Sequencing failed to identify a causative mutation (or, in the case of an autosomal recessive EDS, no more than one causative mutation was identified), AND
• No previous deletion/duplication analysis of the requested gene.

Exceptions and other considerations

The following are specifically non-reimbursable indications for EDS gene sequencing and deletion/duplication analysis:

• Hypermobile EDS or the related clinical entity, “joint hypermobility syndrome”
• Nonsyndromic joint hypermobility, including both asymptomatic and symptomatic forms (e.g., “hypermobility spectrum disorders”)

References


EndoPredict for Breast Cancer Prognosis

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedure addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>EndoPredict Breast Cancer Assay</td>
<td>81599</td>
</tr>
<tr>
<td></td>
<td>S3854</td>
</tr>
</tbody>
</table>

What is EndoPredict for breast cancer prognosis

Definition

EndoPredict® is a commercial multigene expression profiling assay designed to assess prognosis in early-stage breast cancer patients.¹

- The assay combined with results of the tumor size and nodal status is intended to predict the likelihood of women with early stage, node-negative, hormone receptor positive, and HER2 negative breast cancer of developing metastasis within 10 years of initial diagnosis.¹
- This test identifies 12 genes related to tumor proliferation and hormone receptor activity, but does not assess ER or HER2 status.¹
- Test results of the 12-gene risk score are designed to guide decisions regarding adjuvant systemic chemotherapy in women with early-stage invasive breast cancer with known hormone receptor and human epidermal growth factor receptor 2 (HER2) status following surgical management of breast cancer.¹

Test information

- The EndoPredict assay analyzes the gene expression level of 8 breast-cancer related genes and 4 reference genes (12 genes in total) within a breast tumor to determine an EndoPredict score (EP), ranging from 0 to 15. Each score corresponds to a specific likelihood of breast cancer recurrence within 10 years after the initial diagnosis. Based on the calculated score, the patient is categorized as follows:
  - Low risk: 0 to <5
• High risk: 5 to 15 for distant recurrence under endocrine therapy.¹

• When combining the score with clinical risk factors, such as tumor size and node status, a combined molecular and clinical risk score, EPclin, is established. The EPclin score assigns patients into low- and high-risk groups. Patients placed in the high-risk group may be recommended to have chemotherapy, but those in the low-risk group may be able to forego chemotherapy and be spared its associated complications.¹

Guidelines and evidence

• The American Society of Clinical Oncology (ASCO, 2016) published a clinical practice guideline regarding the use of biomarkers to guide clinical decision-making on adjuvant systemic therapy among women with early-stage invasive breast cancer.² Based on a review of the peer-reviewed scientific evidence, the following recommendations were published:

  o If a patient has ER/PgR-positive, HER2-negative (node-negative) breast cancer, the clinician may use the 12-gene risk score (EndoPredict; Sividon Diagnostics, Koln, Germany) to guide decisions on adjuvant systemic chemotherapy. Type: evidence based. Evidence quality: intermediate. Strength of recommendation: moderate.

  o If a patient has ER/PgR-positive, HER2-negative (node-positive) breast cancer, the clinician should not use the 12-gene risk score (EndoPredict) to guide decisions on adjuvant systemic chemotherapy. Type: evidence based. Evidence quality: insufficient. Strength of recommendation: moderate.

  o If a patient has HER2-positive breast cancer or TN breast cancer, the clinician should not use the 12-gene risk score (EndoPredict) to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: insufficient. Strength of recommendation: strong.

• Two clinical validation studies were identified that used archived specimens from previous prospective RCTs (retrospective-prospective study).³,⁴ Of the studies identified, these two prospective-retrospective studies are considered moderate quality evidence (Simon Level I evidence; category B; prospective using archived samples).⁵

  o Filipits et al. (2011) evaluated two groups of patients derived from two independent RCTs (ABC哲6 and ABC哲8) to assess the validity of both the EP and EPclin.³ ABC哲6 was a phase 3 RCT comparing tamoxifen alone for 5 years with tamoxifen in combination with aminoglutethimide for the first 2 years of treatment in postmenopausal women. In ABC哲8, postmenopausal breast cancer patients were randomly assigned to receive tamoxifen for either 5 or 2 years followed by anastrozole for 3 years. Filipits et al. (2011) included women who had participated in the ABC哲6 trial (n=378; tamoxifen-only arm; mean follow-up, 97.4 months) or the ABC哲8 trial (n=1324; mean follow-up, 72.3
months). All tumor specimens were collected at the time of surgery before adjuvant therapy. Assessors of samples, qRT-PCR analyses and score calculations were blinded to clinical and outcome data. The primary outcome measure was distant disease recurrence. Study authors reported that qRT-PCR was successfully analyzed in ~96% and ~99% of the two patient groups.

- EPclin had significantly greater prognostic power compared with clinical pathology factors alone (c indices: 0.76 vs 0.75; P=0.024 [ABCSG-6]; 0.726 vs 0.70; P=0.003 [ABCSG-8]).

- At 10 years, the distant recurrence rates were as follows:
  - Low EP and High EP (ABCSG-6): 8% (95% CI, 3-13%) and 22% (95% CI, 15-29%) (P<0.001)
  - Low EP and High EP (ABCSG-8): 6% (95% CI, 2-9%) and 15% (95% CI, 11-20%) (P<0.001)
  - Low EPclin and High EP (ABCSG-6): 4% (95% CI, 1-8%) and 28% (95% CI, 20-36%) (P<0.001)
  - Low EPclin and High EP (ABCSG-8): 4% (95% CI, 2-5%) and 22% (95% CI, 15-29%) (P<0.001)

  Buus et al. (2016) conducted a prospective-retrospective study to estimate the risk of distant recurrence in women with early-stage breast cancer (ER+/HER2-) considering adjuvant therapy. Patients were evaluated in the prospective RCT (ATAC) evaluating the safety and efficacy of anastrozole compared with tamoxifen in postmenopausal women. Women with either node-positive or node-negative disease were included (n=928). The majority of the population had node-negative disease (n=680; 73%). (Study results that focused on node-negative disease will be discussed in this section.) Among node-positive patients, 59 had disease recurrence (8.6%). EP and EPclin were predictive of recurrence at 10 years of follow-up in both low- and high-risk groups. At 10 years, the distant recurrence rates based on Kaplan-Meier plots, stratified by pre-specified cut-off points, were as follows:
  - Low EP and High EP: 3% (95% CI, 1.5-6%) and 14.6% (95% CI, 11.3-18.8%)
  - EP hazard ratio (HR; 95% CI): 5.15 (2.44-10.85) (P<0.001)
  - Low EPclin and High EP: 5.9% (95% CI, 4-8.6%) and 20% (95% CI, 14.6-27%)
  - EPclin HR (95% CI): 3.90 (2.33-6.52%) (P<0.001)

- A prospective-retrospective study was conducted to evaluate the ability of the EndoPredict assay to identify those patients who would achieve the most benefit from continuing hormonal therapy after 5 years. The study used archived samples from a population of ER-positive/HER2-negative post-menopausal women (node positive and node negative) from the ABCSG-6 (n=378; tamoxifen-only arm) and
ABCSG-8 trials (n=1324) described in the earlier Dubsky study. Patients were retrospectively classified to low- and high-risk EP categories based on the incidence of late recurrence. Based on Kaplan–Meier analysis of distant metastasis, assignment to the EP low-risk group was associated with a significantly reduced risk of recurrence between 0 to 5 years (HR 2.80; 95% CI, 1.81-4.34, P<0.001) and greater than 5 years (HR 3.28; 95% CI 1.47-7.24, P=0.002). Values for the EP high-risk group were not reported by study authors. When EndoPredict and clinical parameters were combined, the prediction of late recurrence was improved as evidenced by the improved c-index; the EPclin score had the highest c-index (0.786) in predicting late recurrence.

- In a prospective-retrospective study, Martin et al. (2014) evaluated the EP score in node-positive breast cancer patients (ER+/HER2-) who were treated with adjuvant chemotherapy followed by hormone therapy. The study also evaluated whether EP scores could predict the efficacy of incorporating weekly paclitaxel into anthracycline-based regimens. Patients enrolled in the RCT (n=555; GEICAM 9906) were evaluated for distant metastasis-free survival (MFS). Rates of MFS at 10 years of follow-up were 93% for the EP low-risk group and 70% for the EP high-risk group, with an absolute risk reduction of 23% (HR 4.8; 95% CI 2.5-9.6; P<0.0001). Adding weekly paclitaxel treatment did not have an effect on the risk of relapse. The EPclin score c-index estimate of 0.70 was the highest compared with other risk factors.

- No direct evidence regarding clinical utility of EndoPredict to improve clinical decision making (e.g., predicting recurrence and/or selecting treatment approaches based on test results) and improve patient health outcomes in women with early-stage breast cancer considering adjuvant chemotherapy was identified. Weak indirect evidence from one small study (n=167) that evaluated treatment decisions assessed retrospectively suggests a potential for the test’s clinical utility. When pre-and post-test decisions were compared, a change of therapy was observed in nearly 38%. In addition, 16 patients (~12%) changed to a treatment strategy of additional chemotherapy; 33 patients (~25%) of patients changed to endocrine treatment alone. In addition to limitations of the retrospective study design, it is unclear how these projected and altered treatment recommendations would translate into improved morbidity and mortality outcomes in this patient population.

- There is adequate evidence in the peer-reviewed literature from two retrospective-prospective studies of moderate quality to support testing with EndoPredict in women with early stage (ER+/HER2-) node-negative breast cancer who are considering adjuvant chemotherapy. Moderate quality evidence indicates that use of the EndoPredict test may improve predictions regarding an individual’s long-term prognosis up to 10 years and determine if they can safely avoid adjuvant chemotherapy.

- There is currently insufficient evidence in the peer-reviewed literature regarding the use of EndoPredict in women with early stage (ER+/HER2-) node-positive breast cancer who are considering adjuvant chemotherapy.

- There is currently insufficient evidence in the peer-reviewed literature regarding the use of EndoPredict in women with early stage (ER+/HER2-) node-negative or node-
positive breast cancer who are disease-free at 5 years after initial diagnosis, currently receiving adjuvant hormonal therapy, and who are considering continuing hormonal therapy.

• **Ongoing clinical trials:**
  
  o NCT Number: NCT02773004\(^{10}\)
    - Title: Prospective Study Assessing EndoPredict\(^{®}\) Genomic Test Impact on Shared Decision of Adjuvant Chemotherapy in Patients With ER-positive, Her2-negative Early Breast Cancer (Active, not recruiting)
  
  o NCT Number: NCT01805271\(^{11}\)
    - Title: Safety Study of Adding Everolimus to Adjuvant Hormone Therapy in Women With High Risk of Relapse, ER+ and HER2- Primary Breast Cancer, Free of Disease After Receiving at Least One Year of Adjuvant Hormone Therapy

**Criteria**

• Previous Testing:
  
  o No repeat EndoPredict testing on the same sample when a result was successfully obtained, and
  
  o No previous gene expression assay (e.g. OncotypeDx Breast) performed on the same sample when a result was successfully obtained, AND

• Required Clinical Characteristics:
  
  o Primary invasive breast cancer meeting all of the following criteria:
    
    o Unilateral tumor
      - Tumor size >0.5cm (5mm) in greatest dimension (T1b-T3)
      - Hormone receptor positive (ER+ or PR+), and
      - HER2 negative, and
    
    o Patient has no regional lymph node metastasis (pN0) or only micrometastases (pN1mi, malignant cells in regional lymph node(s) not greater than 2.0mm), and
Adjuvant endocrine systemic chemotherapy is a planned treatment option for the patient or results from this EndoPredict test will be used in making adjuvant chemotherapy treatment decisions, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Other considerations**

- Testing Multiple Samples:
  - When more than one ipsilateral breast cancer primary is diagnosed, testing should be performed on the tumor with the most aggressive histologic characteristics. If an exception is requested, the following criteria will apply:
    - There should be reasonable evidence that the tumors are distinct (e.g., different quadrants, different histopathologic features, etc.), AND
    - There should be no evidence from either tumor that chemotherapy is indicated with or without knowledge of the EndoPredict test result (e.g., histopathologic features or previous EndoPredict result of one tumor suggest chemotherapy is indicated), AND
    - If both tumors are to be tested, both tumors must independently meet the required clinical characteristics

**References**


Expanded Carrier Screening Panels

Introduction

Expanded carrier screening panels are addressed by this guideline.

Procedures addressed

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<tr>
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<td>SERPINA1 Targeted Mutation Analysis</td>
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### What are expanded carrier screening panels

**Definition**

Expanded carrier screening panels, also known as multiplex carrier screening panels, are designed to identify carrier status or predict risk for multiple genetic diseases in a single test. It is typically offered to individuals planning a pregnancy or currently pregnant.

**Prevalence**

The genetic diseases that are tested for range in severity from lethal in infancy to so mild an affected individual may never develop symptoms. Some conditions are quite common, especially in certain ethnic groups, while others are rare.

It is generally believed that all people carry several recessive gene mutations. An estimated 1 in 580 births has an autosomal recessive condition and 1 in 2000 have an X-linked condition.¹
Inheritance

A carrier has a single recessive gene mutation that does not cause symptoms for the person with the mutation.

Most commonly, both parents have to be carriers of the same genetic condition to have an affected child, which is autosomal recessive inheritance. In this case, each pregnancy has a 25% risk to be affected when both parents are carriers of mutations in the same gene.

Expanded carrier screening panels may include mutations for some X-linked conditions as well. In this case, a mother can be an unaffected carrier but is at risk to have a son with the genetic disease if she passes on that mutation. The father does not need to be a carrier to have an affected child in this situation.

Common uses

Carrier screening is most commonly done for reproductive planning, to identify couples at risk for having a child with a recessive inherited disorder. Carrier screening for a specific disorder may be indicated when there is a positive family history, when a reproductive partner is a carrier of or affected with a recessive disorder, or when there is a known increased risk based on ethnicity or other factors.

Test information

Introduction

Expanded carrier screening panels determine carrier status for numerous genetic conditions simultaneously for the purposes of reproductive planning.

Expanded carrier screening panels

Several expanded carrier screening panels are available. Each test has a unique set of diseases included in novel and proprietary genetic testing platforms. The number of mutations tested varies considerably by condition, ranging from a single mutation for rare conditions to over 100 mutations for cystic fibrosis. Complete testing information, including a list of all conditions screened, can be found at a laboratory's website.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to expanded carrier screening.
American College of Obstetrics and Gynecology

The American College of Obstetrics and Gynecology (ACOG, 2017) published a committee opinion that stated the following regarding Expanded Carrier Screening:

- “Ethnic-specific, panethnic, and expanded carrier screening are acceptable strategies for prepregnancy and prenatal carrier screening. Each obstetrician–gynecologist or other health care provider or practice should establish a standard approach that is consistently offered to and discussed with each patient, ideally before pregnancy. After counseling, a patient may decline any or all carrier screening.”

  “Given the multitude of conditions that can be included in expanded carrier screening panels, the disorders selected for inclusion should meet several of the following consensus-determined criteria: have a carrier frequency of 1 in 100 or greater, have a well-defined phenotype, have a detrimental effect on quality of life, cause cognitive or physical impairment, require surgical or medical intervention, or have an onset early in life. Additionally, screened conditions should be able to be diagnosed prenatally and may afford opportunities for antenatal intervention to improve perinatal outcomes, changes to delivery management to optimize newborn and infant outcomes, and education of the parents about special care needs after birth.”

- “Carrier screening panels should not include conditions primarily associated with a disease of adult onset.”

American College of Medical Genetics and Genomics

The American College of Medical Genetics and Genomics (ACMG, 2013) published a position statement on prenatal and preconception carrier screening. This statement did not provide evidence-based guidance for specific tests or conditions. Rather, it provides general considerations for disease inclusion, clinical relevance, laboratory performance, reporting, and genetic counseling.

Concerns with large panels

Although large panels may be less expensive than doing each carrier screening test individually, most of the included tests are not indicated for each person being tested. Issues with expanded carrier screening include:

- Many included tests have not been recommended for population-based carrier screening and should therefore only be performed when there is a specific known increased risk, such as a family history of the condition.
- Some conditions included in expanded carrier screens are exceedingly rare except in certain ethnicities.
- Mutation analysis may not be the preferred initial screening test for some conditions. For example, a CBC with RBC indices is the initial screening test for
beta-thalassemia followed by hemoglobin analysis for individuals with microcytic anemia.\(^3\) Measuring hexosaminidase A activity may be preferable to mutation analysis for Tay-Sachs carrier screening, especially in non-Jewish populations.\(^4\)

- Some expanded carrier screens include testing for conditions that are relatively mild, treatable, or have onset in adulthood.
- Depending on ethnicity, current expanded carrier screening panels are expected to identify up to 40% of people tested as carriers of a recessive gene mutation. Therefore, if this screening is routinely offered, many patients will require counseling for a positive result, and partner testing must be offered. The most complete partner testing is often by full gene sequencing. Availability of partner testing, cost, turnaround time, and the possibility of identifying a variant of unknown significance by sequencing make this a complex clinical scenario to manage in the routine reproductive setting.

### Criteria

#### Introduction

Requests for expanded carrier screening panels are reviewed using these criteria.

#### Individually billed gene tests

Individual gene tests included in expanded carrier screening panels that will be separately billed should be evaluated based on the medical necessity criteria for each gene test.

Any gene tests that are separately billed and do not meet medical necessity criteria are not a reimbursable service. It will be at the laboratory, provider, and patient’s discretion to determine if a multi-gene panel remains the preferred testing option, recognizing that only a portion of the panel may be reimbursed by insurance.

#### Coverage guidance

This table describes coverage guidance around the most commonly performed carrier screening tests. It also includes the test types addressed by population-based carrier screening guidelines. When the test is not addressed in this table, refer to the general guideline: Genetic Testing for Carrier Status. For these additional tests to be medically necessary, there will generally need to be a specific known increased risk for that condition such as a known family history or a reproductive partner who is known to be a carrier of or affected with the condition.
### Coverage Guidance for Genes Included in Expanded Carrier Screening Multi-Gene Panels

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</table>

**Note** The single Ashkenazi Jewish Carrier Screening guideline should be sufficient to assess the appropriateness of all tests in this category in most circumstances.
available individual gene test policies are provided should additional information be useful.

References

Introduction

These references are cited in this guideline.


Factor II/Prothrombin Testing for Thrombophilia

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
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What is prothrombin thrombophilia

Definition

Prothrombin thrombophilia is a genetic disorder that increases one's risk for developing abnormal blood clots (venous thromboembolism or VTE).¹

- Prothrombin thrombophilia is caused by a genetic change, or mutation, in the F2 gene called G20210A.¹⁻³
  - The F2 gene produces a protein that helps to initiate the formation of blood clots.¹
  - The prothrombin mutation shifts the F2 gene into overdrive, increasing one’s risk of VTE.¹
  - The prothrombin mutation is one of several mutations linked to an increase risk for blood clotting.²⁻³
- The formation of abnormal blood clots can lead to conditions like deep vein thrombosis (DVT) and pulmonary embolism.¹⁻²
- Prothrombin thrombophilia is also linked to an increased risk of miscarriage or other pregnancy complications like preeclampsia, slow fetal growth, and placental abruption.¹⁻²
- About 2% of Caucasians have at least one prothrombin mutation.¹⁻²
  - Inheriting one prothrombin mutation increases one’s risk for developing VTE threefold.¹
  - Inheriting two prothrombin mutations increases one’s risk twentyfold.¹
• Inheriting a prothrombin mutation with other genetic risk factors such as Factor V Leiden also significantly increases the risk for developing VTE.\(^1\)

• Definitive diagnosis of prothrombin thrombophilia relies on both clinical and genetic testing.\(^2,3\)

Test information

• Factor II mutation analysis looks for the G20210A mutation, and determines how many copies of that mutation are present.\(^2,3\) Understanding the number of prothrombin mutations in a suspected case is essential for proper diagnosis, management, and screening. The detection rate for prothrombin mutation analysis is virtually 100%.\(^2,4\)

• Individuals with the prothrombin mutation often have mildly elevated prothrombin levels. These levels can be measured directly in suspected cases of prothrombin thrombophilia.\(^2\) However, levels vary among individuals and even overlap significantly with the normal range.\(^2\) Prothrombin levels are therefore not reliable for the diagnosis of prothrombin thrombophilia, and mutation analysis remains the best choice for definitive diagnosis.\(^2\)

Guidelines and evidence

• Consensus guidelines from the College of American Pathologists (CAP, 2002) related to diagnostic issues in thrombophilia have been issued. These guidelines were obtained by evaluating the literature since 1996 and were accepted if 70% consensus were reached. The guidelines are summarized below:\(^3\)

  o Prothrombin G20210A testing should be performed in the following individuals:
    • A first VTE before age 50 years
    • A first unprovoked VTE at any age
    • A history of recurrent VTE
    • Venous thrombosis at unusual sites such as the cerebral, mesenteric, portal, or hepatic veins
    • VTE during pregnancy or the puerperium
    • VTE associated with the use of oral contraceptives or hormone replacement therapy (HRT)
    • A first VTE at any age in an individual with a first-degree family member with a VTE before age 50 years
    • Women with unexplained fetal loss after the first trimester

  o Prothrombin G20210A testing may be considered in the following individuals/circumstances, but is more controversial:
- Selected women with unexplained early-onset severe preeclampsia, placental abruption, or significant intrauterine growth retardation
- A first VTE related to tamoxifen or other selective estrogen receptor modulators (SERM)
- Female smokers under age 50 years with a myocardial infarction
- Individuals older than age 50 years with a first provoked VTE in the absence of malignancy or an intravascular device
- Asymptomatic adult family members of people with one or two known prothrombin G20210A alleles, especially those with a strong family history of VTE at a young age
- Asymptomatic female family members of people with known prothrombin thrombophilia who are pregnant or considering oral contraception or pregnancy

- Prothrombin G20210A testing is not recommended for the following:
  - General population screening
  - Routine initial testing during pregnancy
  - Routine initial testing prior to the use of oral contraceptives, HRT, or SERMs
  - Prenatal or newborn testing
  - Routine testing in asymptomatic children
  - Routine initial testing in adults with arterial thrombosis

- A consensus statement from the American College of Medical Genetics (ACMG, 2001) on factor V Leiden mutation analysis also provided guidance about prothrombin testing. These older guidelines generally agree with the CAP guidelines of 2002.4

- An Agency for Health Care Research and Quality supported systematic review (AHRQ, 2009) found that, while mutation analysis is effective at identifying prothrombin mutations, “the incremental value of testing individuals with VTE for these mutations is uncertain. The literature does not conclusively show that testing individuals with VTE or their family members for FVL or prothrombin G20210A confers other harms or benefits. If testing is done in conjunction with education, it may increase knowledge about risk factors for VTE.”5

- Evaluation of Genomic Applications in Practice and Prevention Working Group (EGAPP, 2011) found sufficient evidence to recommend against Prothrombin mutation analysis in the following scenarios:6
  
a) Adult with idiopathic VTE,
  
b) Asymptomatic adult family members of patient with VTE and a Prothrombin gene mutation for the purpose of considering primary prophylactic anticoagulation.
Criteria
Testing is indicated in individuals who meet ANY of the following criteria:3,6-9

- Provoked venous thromboembolism (VTE) at a young age (<50 years); or
- Recurrent VTE; or
- Unusual VTE site, such as those involving the hepatic, portal, mesenteric, or cerebral veins; or
- VTE associated with pregnancy or oral contraceptive use; or
- VTE associated with hormone replacement therapy, selective estrogen receptor modulators (SERMs), or tamoxifen; or
- Personal and close family history of VTE; or
- Unprovoked VTE at any age; or
- Family history of venous thrombosis at a young age (<50 years); or
- Women experiencing recurrent pregnancy loss; or
- Women with a history of other unexplained poor pregnancy outcomes, including severe preeclampsia, placental abruption, fetal growth retardation, and stillbirth; or
- Family history of prothrombin gene mutation, particularly when results may impact oral contraceptive use or pregnancy management; or
- Myocardial infarction before age 50, particularly in female smokers

References


**Factor V Leiden Testing for Thrombophilia**

**MOL.TS.167.A**

v2.0.2018

**Procedures addressed**

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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<tr>
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</table>

**What is Factor V Leiden thrombophilia**

**Definition**

About 1 in 1000 people in the U.S. experiences a first venous thromboembolism (VTE) each year, and about one-third of symptomatic patients will develop pulmonary embolism (PE). VTE is a multifactorial condition, usually arising from a combination of genetic, acquired and circumstantial events and risk factors.

- A variant in the factor V gene (F5), called factor V Leiden (FVL), is the most common genetic risk factor for thrombophilia (hypercoagulability) among Caucasians.
  - F5 plays a critical role in forming blood clots.
  - A molecule called activated protein C (APC) keeps the size of clots in check by turning off F5 when clots have formed completely.
  - The FVL variant prevents APC from inactivating F5, increasing the chance of developing abnormal blood clots.
  - The FVL variant is one of several changes in the F5 gene that are reportedly linked to an increase risk of blood clotting.

- The risk for FVL-related thrombosis depends on whether one or two FVL variants are present and additional risk factors, such as prothrombin gene variants.
  - A single FVL variant increases the risk for initial VTE up to 3-8 fold. Two FVL variants increases the risk more dramatically at 18-80 fold. While the risk of subsequent VTE is significantly increased in anyone with a history of VTE, the risk for recurrent VTE attributable to a FVL variant after a first event is much more modest with a pooled odds ratio of 1.56 for single variant and 2.65 for two variants.
The increased risk for pregnancy-related VTE is estimated at 8 fold with a single FVL variant and 20-40 fold with two variants.³

The risk for oral contraceptive-related VTE is estimated at 16 fold with a single FVL variant and over 100 fold with two variants.³

FVL mutations have also appeared to have a small but significant association with some poor pregnancy outcomes in retrospective studies. However, more recent prospective data does not support an increased incidence of pregnancy loss among those with an FVL variant.⁵ There has been conflicting evidence about the association of these variants with other pregnancy complications, such as severe preeclampsia, intrauterine growth restriction, and placental abruption.³⁵

Inheriting an FVL variant with other genetic risk factors also significantly increases the risk for developing VTE. For example, inheriting both a single FVL variant and a single prothrombin variant appears to increase the risk for VTE 20 fold.³

The frequency of FVL varies by ethnicity with about 5% of Caucasians, 2% of Hispanics, and 1% of African Americans in the US having one FVL variant.⁴ About 1 in 1500 Caucasian people have two variants.⁴

Test information

- Factor V Leiden genotyping looks specifically for the Leiden variant (1691G>A; R506Q) in the F5 gene. The detection rate for genotyping is virtually 100%.³ Genotyping can determine how many Leiden variants a person has and therefore can provide information about relative risk of clotting. Understanding the number of Leiden variants in a suspected case is essential for proper diagnosis and management.

- In addition to factor V Leiden genotyping, the modified APC resistance assay is available to detect factor V Leiden thrombophilia. This assay makes use of the fact that the Leiden variant creates a protein that resists inactivation by activated protein C (APC). The APC resistance assay is effective, but does not determine how many copies of the Leiden variant are present. Therefore, if positive, factor V Leiden genotyping is recommended to confirm the findings and quantify the number of variants present.³

- Proposed uses for a positive test result include:
  - Treatment decisions for preventing recurrent VTE in an affected person
  - Primary prevention of VTE in at-risk relatives
  - Decisions about use of oral contraceptives, hormone replacement therapy, or other estrogen-containing therapies
Management decisions for preventing VTE or other possibly associated complications in pregnancy

Guidelines and evidence

- Early consensus statements from the American College of Medical Genetics (ACMG, 2001)\(^6\) and the College of American Pathologists (CAP, 2002)\(^7\) recommended factor V Leiden (FVL) variant testing in the populations most likely to have a mutation. These included:
  - VTE at a young age (<50 years)
  - Recurrent VTE
  - Unusual VTE site, such as those involving the hepatic, portal, mesenteric, or cerebral veins
  - VTE associated with pregnancy or oral contraceptive use
  - VTE associated with hormone replacement therapy, selective estrogen receptor modulators (SERMs), or tamoxifen
  - Personal and close family history of VTE
  - Unprovoked VTE at any age
  - Family history of VTE at a young age (<50 years)

- An Agency for Health Care Research and Quality (AHRQ, 2009) supported systematic review found that, while variant analysis is effective at identifying FVL variants, “the incremental value of testing individuals with VTE for these mutations is uncertain. The literature does not conclusively show that testing individuals with VTE or their family members for FVL or prothrombin G20210A confers other harms or benefits. If testing is done in conjunction with education, it may increase knowledge about risk factors for VTE.”\(^8\)

- The Evaluation of Genomic Applications in Practice and Prevention (EGAPP, 2011), an initiative of the CDC Office of Public Health Genomics, evaluated the clinical utility evidence for two limited scenarios:
  a) anticoagulation duration to prevent recurrence in people with idiopathic VTE and
  b) primary VTE prevention in their at-risk relatives. They specifically exclude individuals with other risk factors for VTE, such as estrogen-containing therapy use. EGAPP makes the following recommendations:\(^4\)
    - [EGAPP] found adequate evidence to recommend against routine testing for Factor V Leiden (FVL) and/or prothrombin 20210G>A (PT) in the following circumstances: (1) adults with idiopathic venous thromboembolism (VTE). In such cases, longer term secondary prophylaxis to avoid recurrence offers similar benefits to patients with and without one or more of these mutations. (2) Asymptomatic adult family members of patients with VTE and an FVL or...
PT mutation, for the purpose of considering primary prophylactic anticoagulation. Potential benefits are unlikely to exceed potential harms.

- Because anticoagulation is associated with significant risks and these mutations are associated with relatively low absolute VTE risk, the potential harms of overtreatment in these scenarios appears to outweigh the benefits of testing. However, test results may be used for other treatment decisions, such as anticoagulation in high-risk situations (e.g., surgery, pregnancy, long-distance travel), avoidance of estrogen-containing therapies, or the use of low-risk preventive measures (e.g., compression hose, activity counseling, smoking cessation). The authors noted that the evidence was insufficient to determine if testing might have utility in some situations, such as for influencing patient behavior or identifying those with homozygous mutations or combined thrombophilias. Therefore, these findings have limited application to the broader decision about who should be tested.

- Several other organizations have issued guidelines that help inform a decision about clinical utility by defining the change, or lack of change, in management of patients with known FVL thrombophilia in specific clinical circumstances.
  
  o VTE management:
    
    - The American College of Chest Physicians (ACCP, 2008) recommends the same management for unprovoked VTE or VTE associated with a transient (reversible) risk factor (such as estrogen-containing therapies) irrespective of FVL results.  
    
    - These guidelines add “The presence of hereditary thrombophilia has not been used as a major factor to guide duration of anticoagulation for VTE in these guidelines because evidence from prospective studies suggests that these factors are not major determinants of the risk of recurrence.”
    
    - Also note that the above referenced EGAPP (2011) study specifically addresses this test use and finds “There is no evidence that knowledge of FVL/PT mutation status in patients with VTE affects anticoagulation treatment to avoid recurrence.” “There is convincing evidence that anticoagulation beyond 3 months reduces recurrence of VTE, regardless of mutation status.”

  o Pregnancy management:
    
    - The American College of Chest Physicians (ACCP, 2008) recommends the same management for VTE in a current pregnancy or for those with a prior VTE history during or outside of pregnancy irrespective of FVL results. However, if a higher risk thrombophilia is present, such as two Leiden variants or a combination of a Leiden and prothrombin variant, ACCP recommends some form of treatment and not simply surveillance.
Thrombophilia in pregnancy guidelines from the American College of Obstetricians and Gynecologists (ACOG, 2013) state:

- Testing is controversial and is "is useful only when results will affect management decisions, and is not useful in situations where treatment is indicated for other risk factors." However, they add that screening "may be considered" for those with "A personal history of venous thromboembolism that was associated with a non-recurrent risk factor (e.g. fractures, surgery, and prolonged immobilization). The recurrence risk among untreated pregnant women with such a history and a thrombophilia was 16% (odds ratio, 6.5; 95% confidence interval, 0.8–56.3)."5

- They add "Testing for inherited thrombophilias in women who have experienced recurrent fetal loss or placental abruption is not recommended because it is unclear if anticoagulation therapy reduces recurrence. Although there may be an association in these cases, there is insufficient clinical evidence that antepartum prophylaxis with unfractionated heparin or low molecular weight heparin (LMWH) prevents recurrence in these patients"

Estrogen-containing therapy decisions:

- American College of Obstetricians and Gynecologists (ACOG, 2006) contraceptive use guidelines state “Combination contraceptives are not recommended for women with a documented history of unexplained venous thromboembolism or venous thromboembolism associated with pregnancy or exogenous estrogen use, unless they are taking anticoagulants.”11 Therefore, estrogen-containing drugs are contraindicated based on a history of VTE alone irrespective of FVL results.

- American Association of Clinical Endocrinologists (AACE, 2011) menopause guidelines says only the following about menopausal hormone therapy (MHT): “Estrogen therapy has been associated with an increased risk of venous thromboembolic disease within 1 to 2 years after initiation of therapy. The increased relative risk (RR) is high, but the increased absolute risk is quite small…The incidence was greater with increasing age, obesity, and factor V Leiden mutations (45 [EL 1; RCT]). Women with a history of venous thromboembolic disease should be carefully advised about this risk when MHT is being considered.”12

Family history of a Leiden variant:

- The above referenced EGAPP (2011) statement specifically addresses this test use for VTE prophylaxis and found “There is no evidence that knowledge of FVL/PT mutation status among asymptomatic family members of patients with VTE leads to anticoagulation aimed at avoiding initial episodes of VTE.”4
• American College of Obstetricians and Gynecologists (ACOG, 2010) states that testing is controversial and should only be done when the results will change management. However, they add that screening “may be considered” for those with “A first-degree relative (eg, parent or sibling) with a history of high-risk thrombophilia.”

• Generally, estrogen-containing drugs must be approached with caution in anyone with a significant family history of VTE or known FVL and/or PT mutations, but no US evidence-based guidelines were identified that addressed testing in this scenario. Guidelines from the British Society for Haematology (BSH, 2010) most directly address FVL and PT testing in at-risk relatives for the purposes of deciding about estrogen-containing therapies. They recommend considering “alternative contraceptive or transdermal HRT [hormone replacement therapy]” when a first-degree relative: “has not been tested or is negative… Testing for heritable thrombophilia will provide an uncertain estimate of risk and is not recommended (1C).” or “has been tested and the result is positive… Offer alternative contraception, counsel that negative result would not exclude increased risk. However, testing may assist in counseling of selected women particularly if a high risk thrombophilia has been identified in the symptomatic relative (C).”

• The evidence supporting an association between FVL variants and thrombosis is adequate (clinical validity). However, there are no clinical situations in which FVL testing is either mandatory or specifically recommended in guidelines due to generally insufficient clinical utility data. Factor V Leiden genotyping may have some utility in limited circumstances where there is a recognized increased risk to have at least one mutation based on established risk factors, where the results will be used to direct management beyond the current VTE, and particularly when individuals are found to have a combination of more than one factor V Leiden mutation or additional genetic thrombophilias (despite the absence of reliable indicators). If testing is performed, there should be a specific plan for how the results will impact management.

Criteria

• Genetic Counseling
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous genetic testing for Factor V Leiden mutation, AND

• Individual has at least one of the following risk factors suggesting a higher likelihood of having one or more factor V Leiden variants:
- Unprovoked/idiopathic venous thromboembolism at any age, or
- History of recurrent venous thromboembolism, or
- Venous thrombosis at an unusual site (e.g., cerebral, mesenteric, hepatic, and portal veins), or
- Venous thromboembolism during pregnancy or the puerperium, or
- Venous thromboembolism associated with the use of estrogen-containing therapies (e.g., oral contraceptives or hormone replacement therapy), or
- A personal history of any venous thromboembolism combined with a first-degree family member with venous thromboembolism before the age of 50 years, or
- Known factor V Leiden variant(s) identified in at least one 1st degree relative (parent, sibling, child). (Note: 2nd or 3rd degree relatives may be considered when 1st degree relatives are unavailable or unwilling to be tested), AND

- Test results will be used for guiding management decisions beyond simply therapy of a current first venous thrombosis event or related future prophylaxis decisions, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

The following factor V Leiden genotyping test applications are specifically considered investigational and/or experimental:

- Testing without clear evidence of an increased likelihood of having at least one factor V Leiden variant. This includes but is not limited to:
  - Testing performed as part of expanded cardiovascular disease screening
  - Testing based on the presence of conditions with unclear evidence including stroke, myocardial infarction, pregnancy loss, and pregnancy complications

References


Familial Adenomatous Polyposis Testing

Procedures addressed

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<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>APC Known Familial Mutation Analysis</td>
<td>81202</td>
</tr>
<tr>
<td>APC Sequencing</td>
<td>81201</td>
</tr>
<tr>
<td>APC Deletion/Duplication Analysis</td>
<td>81203</td>
</tr>
</tbody>
</table>

What is Familial Adenomatous Polyposis (FAP)

Definition

FAP is an inherited colorectal cancer syndrome that accounts for up to 1 in 200 colorectal cancers.¹

• FAP is clinically diagnosed when a person has 100 or more colorectal adenomatous polyps or fewer than 100 polyps and a family member with FAP. Polyposis typically begins before age 40. Virtually all people with classic FAP will develop colorectal cancer without intervention. Other clinical manifestations include:¹
  
  o Modestly increased risk for other malignancies including cancers of the thyroid, small bowel, stomach, liver (hepatoblastoma, typically seen in children under 5), pancreas, brain (medulloblastoma), and bile duct.
  
  o Additional gastrointestinal manifestations including duodenal adenomas and gastric polyps.
  
  o Non-gastrointestinal manifestations including osteomas (often of the mandible or skull), dental abnormalities (supernumerary teeth, odontomas), desmoid tumors, soft tissue tumors (epidermoid cysts, fibromas), adrenal masses (adenomas), and congenital hypertrophy of retinal epithelium (CHRPE).¹ Isolated CHRPE may be found in the general population, but multiple or bilateral CHRPE in an at-risk family member may be suspicious for FAP.
FAP with osteomas or soft tissue tumors suggests the Gardner syndrome variant. FAP with medulloblastoma suggests the Turcot syndrome variant.

- Attenuated FAP (AFAP) is a milder form characterized by the presence of 10-99 polyps. Colon cancer generally presents at a later age than classic FAP. Individuals with 100 or more polyps occurring at later ages (35 to 40 years or older) may be found to have AFAP. A personal history of colorectal cancer before age 60 (without polyposis) and a family history of multiple adenomatous polyps may also be seen with AFAP. Currently, there is no consensus regarding precise diagnostic criteria for AFAP.\(^1,2\)

- Almost all cases of FAP and some cases of AFAP are due to mutations in the adenomatous polyposis coli (APC) gene, a tumor suppressor gene. Most people inherit an APC mutation from an affected parent, but up to 1 in 4 people with FAP have a new mutation with no known affected family members. Parents of someone with FAP may also be unaffected due to germline mosaicism (a mix of normal and mutated copies of the APC gene are confined to the parent's eggs or sperm).\(^1\)

- Management and prevention strategies for those affected with or at-risk for FAP/AFAP include annual flexible sigmoidoscopy or colonoscopy screening beginning at 10-15 years for FAP and every 2-3 years beginning in the late teens for AFAP. Prophylactic colectomy is generally recommended when sufficient polyps emerge such that polyposis cannot be managed endoscopically.\(^3\)

**Test information**

- APC sequence analysis is used to identify disease-causing mutations in those clinically diagnosed with FAP/AFAP.\(^3-6\) Testing may be considered for close relatives of someone with FAP when an affected relative is unavailable for testing.\(^5\)

  - Sequence analysis detects a mutation in up to 90% of individuals clinically diagnosed with FAP.\(^1\) The mutation detection rate is lower for those with AFAP than classic FAP.\(^2\)

- APC deletion/duplication testing is typically performed in reflex to negative analysis. Deletion/duplication testing detects an additional 8-12% of mutations in those with clinical suspicion of FAP.\(^1\)

- Molecular genetic testing of MUTYH should be considered next if no APC mutation is found.\(^1\)

- “Another strategy is to perform concurrent genetic testing of two or more genes known to be associated with colon cancer predisposition."\(^1\)

  - “Concurrent molecular genetic testing for both APC and MUTYH may be considered. These two genes may also be represented together on a multi-gene panel."
“Multi-gene panels can be used for the simultaneous analysis of some or all of the genes known to be associated with intestinal polyposis conditions. These panels vary by methods used and genes included.”

- Once a disease-causing mutation has been identified, at-risk family members can be tested for that known familial mutation. This may be called single site mutation analysis. Those proven not to have inherited a known family mutation through genetic testing can avoid the additional screening required for those at-risk for FAP.¹

- A common variant in the APC gene, called I1307K, may mildly increase the risk for colorectal cancer, but **does not cause FAP**. Testing for this variant is not widely accepted.

### Guidelines and evidence

- **Consensus guidelines from the American Gastroenterological Association (AGA, 2001) recommend:**⁴,⁵
  - APC gene testing in individuals age 10 or older to confirm the diagnosis of FAP or AFAP, or to provide presymptomatic screening in individuals age 10 or older with a first-degree relative with FAP or AFAP.
  - First testing an affected family member to establish if a detectable mutation is present in the family.

- **Evidence- and consensus-based guidelines from the National Comprehensive Cancer Network (NCCN, 2016) state:**³
  - “APC genetic testing is recommended in a proband to confirm a diagnosis of FAP and allow for mutation specific testing in family members. Additionally knowing the location of the mutation in the APC gene can be helpful for predicting severity of polyposis, rectal involvement and desmoid tumors.”
  - When the family mutation is known, APC gene testing is recommended for at-risk family members (defined as first-degree relatives or more distant relatives if closer relatives are unavailable or unwilling to be tested).
  - When the family mutation is not known, APC gene testing may be considered for first-degree relatives when an affected family member is not available or not willing to test first.
These recommendations are Category 2A, defined as “lower-level evidence with uniform NCCN consensus.”

- Evidence-based guidelines from the American College of Gastroenterology (ACG, 2009) recommend:⁶
  - Patients with classic FAP (>100 adenomas) should be advised to pursue genetic counseling and genetic testing, if they have siblings or children who could potentially benefit from this testing.”[Grade 2B: “weak recommendation, moderate-quality evidence”].

- The American College of Gastroenterology (ACG, 2015) clinical guidelines state that “Individuals who have a personal history of >10 cumulative colorectal adenomas, a family history of one of the adenomatous polyposis syndromes, or a history of adenomas and FAP-type extracolonic manifestations (duodenal/ampullary adenomas, desmoid tumors, papillary thyroid cancer, congenital hypertrophy of the retinal pigment epithelium, epidermal cysts, osteomas) should undergo assessment for the adenomatous polyposis syndrome.”⁷

- Note that NCCN excluded I1307K variant testing from the guideline “because there is very little evidence to date indicating what kind of screening should be offered to individuals with this mutation.”³

Criteria

APC Known Familial Mutation Analysis

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous genetic APC mutation testing, AND

- Diagnostic or Predisposition Testing:
  - Family History:
    - Known family mutation in APC identified in 1st degree relative(s). (Note: 2nd or 3rd degree relatives may be considered when 1st degree relatives are unavailable or unwilling to be tested), AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

APC Sequencing

- Genetic Counseling:
Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous APC mutation testing, and
  o No known familial mutation, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Personal history:\textsuperscript{5,7}
    ▪ More than 10 cumulative adenomas (known or suspected diagnosis of FAP – 100 or more adenomas or AFAP – 10 to 100 adenomas), or
    ▪ A desmoid tumor, hepatoblastoma, cribriform-morular variant of papillary thyroid cancer, or multifocal/bilateral CHRPE, OR

• Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  o Family history:
    ▪ First degree relative of an individual with a diagnosis of FAP or AFAP. (Note: Whenever possible, an affected family member should be tested first), AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

**APC Duplication/Deletion Analysis**

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous large rearrangement testing, and
  o Previous APC sequencing performed and no mutations found, and
  o No known familial mutation, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

**References**


Familial Hypercholesterolemia Genetic Testing

Procedures addressed

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<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
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<td>LDLR Known Familial Mutation</td>
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</tr>
<tr>
<td>APOB Known Familial Mutation</td>
<td>81403</td>
</tr>
<tr>
<td>PCSK9 Known Familial Mutation</td>
<td>81403</td>
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<tr>
<td>LDLR Sequencing</td>
<td>81406</td>
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<td>LDLR Deletion/Duplication</td>
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<tr>
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<td>APOB Sequencing</td>
<td>81479</td>
</tr>
<tr>
<td>PCSK9 Sequencing</td>
<td>81406</td>
</tr>
</tbody>
</table>

What is familial hypercholesterolemia

Definition

Familial hypercholesterolemia (FH) is a genetic disorder characterized by very high levels of low-density lipoprotein (LDL) cholesterol.

- Familial hypercholesterolemia (FH) is a genetic disorder characterized by very high levels of low-density lipoprotein (LDL) cholesterol: usually >190 mg/dL in untreated adults. This leads to an increased risk for coronary heart disease (CHD), including heart attacks, at an early age.1,2,3
  - Men with untreated FH have a 50% risk for a coronary event by age 50.3,4
  - Women with untreated FH have a 30% risk for a coronary event by age 60.3,4
- People with untreated FH have about a 20 fold increase for coronary heart disease.3
- Early and aggressive LDL-lowering with high doses of potent statins or statin combination therapy significantly lowers CHD morbidity and mortality for people with FH.5,6 Statins are contraindicated during pregnancy due to concerns for
teratogenicity and should be discontinued prior to conception. Because there is considerable overlap between the LDL levels of those with FH and common multifactorial hypercholesterolemia, FH often goes undiagnosed until middle age, when much of the preventive value of cholesterol-lowering therapy is lost.

- For FH patients who are not adequately controlled with statin therapy, or with intolerance to statins, PCSK9 inhibitors (e.g. evolocumab, alirocumab) may be an effective alternative treatment.
- Less than 10% of people with FH are adequately treated.
- Various criteria for identifying FH clinically have been developed and are described below:

### Diagnosis: MEDPED criteria

MEDPED criteria

**Total Cholesterol (LDL), mg/dL**

<table>
<thead>
<tr>
<th>Patient’s age</th>
<th>Patient has 1(^{st}) degree relative with FH</th>
<th>Patient has 2(^{nd}) degree relative with FH</th>
<th>Patient has 3(^{rd}) degree relative with FH</th>
<th>General population</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;18</td>
<td>220 (155)</td>
<td>230 (165)</td>
<td>240 (170)</td>
<td>270 (200)</td>
</tr>
<tr>
<td>20</td>
<td>240 (170)</td>
<td>250 (180)</td>
<td>260 (185)</td>
<td>290 (220)</td>
</tr>
<tr>
<td>30</td>
<td>270 (190)</td>
<td>280 (200)</td>
<td>290 (210)</td>
<td>340 (240)</td>
</tr>
<tr>
<td>40+</td>
<td>290 (205)</td>
<td>300 (215)</td>
<td>310 (225)</td>
<td>360 (260)</td>
</tr>
</tbody>
</table>

### Diagnosis: Dutch criteria

Definitive FH: 8 points or more; Probable FH: 6-7 points; Possible FH: 3-5 points

<table>
<thead>
<tr>
<th>Points</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>1 point</td>
<td>First-degree relative with premature cardiovascular disease or LDL &gt;95th percentile, or personal history of premature peripheral or cerebrovascular disease or LDL 155-189 mg/dL</td>
</tr>
<tr>
<td>2 points</td>
<td>First-degree relative with tendinous xanthoma or corneal arcus, or first-degree relative age &lt;18 with LDL &gt;95th percentile, or personal history of coronary artery disease</td>
</tr>
<tr>
<td>3 points</td>
<td>LDL 190-249 mg/dL</td>
</tr>
<tr>
<td>4 points</td>
<td>Corneal arcus in patient age &lt;45 years</td>
</tr>
<tr>
<td>5 points</td>
<td>LDL 250-329 mg/dL</td>
</tr>
</tbody>
</table>
### Points Description

<table>
<thead>
<tr>
<th>Points</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>6 points</td>
<td>Tendon xanthoma</td>
</tr>
<tr>
<td>8 points</td>
<td>LDL ≥330 mg/dL**</td>
</tr>
</tbody>
</table>

**Note** Please note that these are LDL level cut offs for untreated individuals.

### Diagnosis: Simon Broome criteria

#### Definitive FH
- Total cholesterol (LDL): 290 (190) mg/dL in adults or 260 (155) mg/dL in pediatric patients and:
- DNA mutation

#### Probable FH
- Total cholesterol (LDL): 290 (190) mg/dL in adults or 260 (155) mg/dL in pediatric patients and:
- Tendon xanthoma in patient or in first- or second-degree relative

#### Possible FH
- Total cholesterol (LDL): 290 (190) mg/dL in adults or 260 (155) mg/dL in pediatric patients and:
- Family history of myocardial infarction (MI) at age <50 in second-degree relative or at age <60 in first-degree relative or family history of total cholesterol >290 mg/dL in first- or second-degree relative

### Prevalence

About 1 in 200-250 people worldwide have FH. The risk is much higher in some South African Afrikaner, Amish, Lebanese, and Finnish populations.

Approximately 1 in 300 to 500 people have heterozygous FH, which means they have one copy of the gene mutation.

Approximately one in 1 million people have homozygous FH, which means they have 2 copies of the gene mutation. This is much more severe than heterozygous FH. People with this type of FH typically have severe coronary heart disease by their mid-20s; the rate of death or the need for surgical treatment of heart problems by the teenage years is high.

### Cause

Most cases of FH are caused by mutations in one of three genes: LDLR, APOB, PCSK9. However, mutations in these genes only account for approximately 60%-80% of FH.
There are likely other genes that are not known at the present time that make up the remaining 20%-40% of cases of FH. Therefore, a negative genetic test does not rule out a diagnosis of FH.

Inheritance

FH is an autosomal dominant condition, meaning that only one gene mutation is needed to cause the condition.

A person with heterozygous FH has a 50% chance to pass the mutation to each child.

Although not included in this guideline, it is important to note that there is an autosomal recessive form of hypercholesterolemia which is caused by mutations in the LDLRAP1 gene. There is also a milder autosomal dominant form, Familial Combined Hyperlipidemia, which is usually caused by mutations in the LPL gene.3

Test information

• A clinical diagnosis of FH is suspected based on some combination of personal and family history of very high cholesterol, premature CHD, and cholesterol deposits, such as tendon xanthomas and corneal arcus.2 At least three organizations have attempted to define clinical diagnostic criteria for FH, but all criteria have recognized limitations.10 The three different criteria are described above.

• Genetic testing for FH can confirm a diagnosis of FH, particularly in borderline clinical cases.9,11,12

• Laboratories may offer evaluation of the LDLR, APOB, or PCSK9 genes individually, as panels, or with reflex options.
  
  o LDLR: Over 1000 mutations have been characterized so sequence analysis is required. Major gene deletions and rearrangements account for an estimated 9% of LDLR mutations and require specialized deletion testing to detect them.13
  o APOB: FH-causing APOB mutations are primarily found in a limited region of the gene, with the R3500Q mutation being most common.13 Laboratory testing may be done by targeted mutation analysis for a limited number of APOB mutations or sequencing of the gene region where these mutations are generally found.3 According to GeneReviews, as of 2016 there has been only one reported case of a deletion in APOB causing FH.3
o **PCSK9**: “Gain of function mutations in PCSK9 cause fewer than 5% of cases in most studies.” According to GeneReviews, as of 2016 there have been no deletions or duplications reported in PCSK9 that cause FH.  

- Once a mutation is found in an affected person, single-site testing should be offered to at-risk family members to allow for appropriately early intervention.

**Genetic testing for FH**

Proportion of FH attributed to each gene.

**Molecular Genetic Testing for FH**

<table>
<thead>
<tr>
<th>Gene</th>
<th>Proportion of FH Attributed to Mutations in Gene</th>
<th>Test Method</th>
</tr>
</thead>
<tbody>
<tr>
<td>LDLR</td>
<td>60%-80%</td>
<td>Sequence Analysis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deletion/Duplication</td>
</tr>
<tr>
<td>APOB</td>
<td>1%-5%</td>
<td>Targeted Analysis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Sequencing Analysis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deletion/Duplication</td>
</tr>
<tr>
<td>PCSK9</td>
<td>0%-3%</td>
<td>Targeted Analysis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Sequencing Analysis</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Deletion/Duplication</td>
</tr>
<tr>
<td>Unknown</td>
<td>20%-40%</td>
<td>NA</td>
</tr>
</tbody>
</table>

**Guidelines and evidence**

**Guidelines and evidence - genetic testing**

- Evidence-based guidelines by the National Institute for Clinical Excellence of UK (NICE, 2008 (reaffirmed 2016)) support genetic testing for FH as follows:
  - “A diagnosis of FH should be made using the Simon Broome criteria, which include a combination of family history, clinical signs (specifically tendon xanthomata), cholesterol concentration and DNA testing (see appendix E of the NICE guideline).”
  - “Healthcare professionals should offer people with a clinical diagnosis of FH a DNA test to increase the certainty of their diagnosis and to aid diagnosis among their relatives.”
  - “Healthcare professionals should inform all people who have an identified mutation diagnostic of FH that they have an unequivocal diagnosis of FH even if
their LDL-C concentration does not meet the diagnostic criteria (see appendix E).

- “In a family where a DNA mutation is identified, not all family members may have inherited the mutation. When DNA testing has excluded FH in a member of a family, healthcare professionals should manage the person’s coronary heart disease risk as in the general population.”

- “In families in which a mutation has been identified, the mutation and not LDL-C concentration should be used to identify affected relatives. This should include at least the first- and second- and, when possible, third-degree biological relatives.”

- “In children at risk of FH because of one affected parent, the following diagnostic tests should be carried out by the age of 10 years or at the earliest opportunity thereafter.

  - A DNA test if the family mutation is known.”

  - “LDL-C concentration measurement if the family mutation is not known. When excluding a diagnosis of FH a further LDL-C measurement should be repeated after puberty because LDL-C concentrations change during puberty.”

• Consensus-based guidelines from The Cardiac Society of Australia and New Zealand (CSANZ, 2013) state: “Although the clinical picture of FH will be clear-cut in many instances, the diagnostic criteria suggest that genetic testing can provide certainty of diagnosis in some cases where confounding factors such as borderline cholesterol levels, inconclusive family histories or tendon injuries have resulted in a diagnostic dilemma.”

• The National Lipid Association expert panel on Familial Hypercholesterolemia (2011) made the following recommendations regarding genetic testing:

  - “Genetic screening for FH is generally not needed for diagnosis or clinical management but may be useful when the diagnosis is uncertain.”

  - “Identification of a causal mutation may provide additional motivation for some patients to implement appropriate treatment.”

  - “Importantly, a negative genetic test does not exclude FH, since approximately 20% of clinically definite FH patients will not be found to have a mutation despite an exhaustive search using current methods.”

Guidelines and evidence - drug treatment

• The US Food and Drug Administration approved the following PCSK9 inhibitors as treatment for FH. However, there have been no guidelines recommending that genetic testing should be performed for the sole purpose of treatment decisions (i.e. PCSK9 inhibitors) in the absence of a clinical suspicion of FH:
“Praluent (alirocumab) injection in adult patients with heterozygous familial hypercholesterolemia or patients with clinical atherosclerotic cardiovascular disease such as heart attacks or strokes, who require additional lowering of LDL cholesterol.”\textsuperscript{17}

“Repatha (evolocumab) injections for use in additional to diet and maximally-tolerated statin therapy in adult patients with heterozygous hypercholesterolemia, homozygous hypercholesterolemia, or clinical atherosclerotic cardiovascular disease, such as heart attacks or strokes, who require additional lowering of LDL cholesterol.”\textsuperscript{18}

Criteria

**LDLR, APOB, PCSK9 Known Familial Mutation Testing**

- Clinical Consultation:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous genetic testing of LDLR, APOB, or PCSK9, and
  - LDLR, APOB, or PCSK9 mutation identified in 1\textsuperscript{st}, 2\textsuperscript{nd} or 3\textsuperscript{rd} degree biological relative, AND
- Diagnostic Testing:
  - LDL cholesterol of >120 mg/dL in the absence of treatment

**LDLR Full Sequence and Deletion/Duplication Analysis**

- Clinical Consultation:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Testing:
  - No previous LDLR sequencing or deletion/duplication testing, and
  - No known LDLR, APOB, or PCSK9 mutation in the family, AND
- Diagnostic Testing:
  - Member meets the MEDPED criteria or either the Dutch criteria or the Simon Broome criteria for possible or probable FH, and
Genetic testing is necessary because there is uncertainty in the clinical diagnosis

**APOB Targeted Mutation Analysis or Full Sequence Analysis**

- Criteria for LDLR sequencing and deletion/duplication analysis is met, AND
- No previous full sequence analysis of APOB, AND
- No mutations detected in full sequencing or deletion/duplication testing of LDLR or PCSK9 sequencing

**PCSK9 Full Sequence Analysis**

- Criteria for LDLR sequencing and deletion/duplication analysis is met, AND
- No previous genetic testing for PCSK9, AND
- No mutations detected in full sequencing or deletion/duplication analysis of LDLR or APOB sequencing

**Exclusions**

Genetic testing for the sole purpose of treatment decisions (i.e. PCSK9 inhibitors) in the absence of a clinical suspicion supported by either the MEDPED, Dutch, or Simon Broome criteria is considered investigational and/or experimental.

**References**


Familial Malignant Melanoma Testing

Procedures addressed

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<tr>
<td>CDK4 Known Familial Mutation Analysis</td>
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<td>CDKN2A Sequencing</td>
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<td>CDKN2A Deletion/Duplication Analysis</td>
<td>81479</td>
</tr>
<tr>
<td>CDK4 Sequencing</td>
<td>81479</td>
</tr>
</tbody>
</table>

What is familial malignant melanoma

Definition

Familial malignant melanoma (FMM) is a strongly inherited form of melanoma.

- The lifetime risk of melanoma for someone born in the U.S may reach 1 in 55. The incidence continues to rise dramatically.
- Most melanoma is sporadic. It usually is the result of a combination of genetic susceptibility (probably from several relatively low risk gene variants such as those involved with pigment) and environmental risk factors such as sun exposure.
- About 4-8% of people with melanoma have a family history of at least one first-degree relative (parent, child, sibling) with melanoma. Less than 1% to 2% have multiple affected relatives, which suggests a stronger genetic susceptibility.
- FMM is most likely in a family when there are three or more close relatives diagnosed with melanoma. Other factors that may also suggest FMM include:
  - Melanoma diagnosed younger than usual (average diagnosis age 30s versus 50s in people without FMM)
  - More than one melanoma primary in the same individual
  - Melanoma and pancreatic cancer in the same family
- Multiple, atypical moles, called dysplastic nevi that are often larger than 5mm in diameter with irregular borders. Melanoma with multiple nevi has also been called familial atypical mole-malignant melanoma syndrome. However, the presence or absence of such moles is no longer viewed as a reliable predictor of FMM in a family.

- Several genes have been linked to a higher risk of melanoma in families. CDKN2A gene mutations account for most of the currently identifiable FMM mutations, followed by CDK4 mutations.\(^6\)

- FMM is an autosomal dominant condition, meaning that only one gene mutation is needed to increase susceptibility to melanoma. A person with FMM has a 50% chance to pass the mutation to each child.

- People who inherit an FMM mutation do not always develop melanoma. Data for CDKN2A mutations suggest that in Europe the melanoma risk is 5% by age 40 and 60% by age 80.\(^4\) The likelihood may vary with geographic location and sun exposure.\(^5\)

- Familial melanoma is also associated with some other inherited cancer syndromes, like Li Fraumeni syndrome, inherited retinoblastoma, and xeroderma pigmentosum.\(^2\) Additionally, germline mutations in the BAP1 gene have been identified in families with cutaneous and ocular melanoma.\(^7\)

### Test information
- **CDKN2A Sequencing:** Identifies the majority of FMM-causing mutations, and is usually the first step in testing. The likelihood that genetic testing will identify an FMM mutation varies with the personal and family history. The chance of finding a CDKN2A mutation is:
  - 20-40% of people with melanoma from a family with at least 3 affected first-degree relatives.\(^2,6\)
  - Less than 5% of those with only 2 affected first-degree relatives\(^2\)
  - 15% in someone with multiple melanoma primaries and no known family history\(^2\)
  - 25-40% in people diagnosed with familial atypical mole-malignant melanoma syndrome - a subset of FMM characterized by >50 atypical nevi with characteristic microscopy features\(^8\)
  - 74% of families with FMM and pancreatic cancer\(^6\)

- **CDKN2A Deletion/Duplication Analysis:** Tests for large deletions that cannot be identified by sequencing.

- **CDK4 Sequencing:** Sequencing, sometimes of only exon 2, is also available, but mutations are uncommon, accounting for only 2-3% of FMM cases.\(^6\)
• CDKN2A Known Familial Mutation Analysis: When the family mutation is known, testing for only the family mutation can be performed in at-risk relatives. Test accuracy approaches 100%.\textsuperscript{2}

• CDK4 Known Familial Mutation Analysis: When the family mutation is known, testing for only the family mutation can be performed in at-risk relatives. Test accuracy approaches 100%.\textsuperscript{2}

Guidelines and evidence

• No evidence-based U.S. guidelines were identified.

• FMM genetic testing outside of the research setting is not currently recommended for several reasons, including:
  
o Currently available testing does not detect a mutation in a significant number of people who appear to have FMM. Therefore, a negative result cannot rule out FMM and should not change the prevention and screening plan for at-risk people.\textsuperscript{2}
  
o Individuals with FMM mutations need essentially the same prevention and screening as anyone at high risk for melanoma (family history, pigmentation, multiple moles, history of blistering sunburn).\textsuperscript{2} Therefore, identifying an FMM-causing mutation is also not expected to change screening or treatment.\textsuperscript{5}
  
o When a family FMM mutation has been found, other relatives who test negative for that mutation at best only return to the background risk for melanoma (which may be as high as 1 in 25) and still need regular skin screening.\textsuperscript{2}
  
o A significant percentage of people with recognized FMM mutations do not develop melanoma, which is especially true when sun exposure is limited by geography or prevention.\textsuperscript{4}

• The Melanoma Genetics Consortium (GenoMEL), an international research collaborative group, published a consensus statement in 1999 stating, “DNA testing for mutations in known melanoma susceptibility genes should only rarely be performed outside of defined research programs. With this general proviso, two distinct clinical situations need further consideration: families in which a CDKN2A mutation has been identified in a proband as part of a research study and families for which no prior testing of affected individuals has been conducted.”\textsuperscript{2}
  
o “Individuals who choose to undergo genetic testing [in a research setting] should have a second independent diagnostic (as distinct from research) DNA test performed in an accredited genetic testing laboratory.”\textsuperscript{2}
For at-risk relatives with a known familial mutation, test sensitivity is virtually 100%. However, the likelihood of developing melanoma in mutation-positive individuals is largely unknown and there is "lack of proved efficacy of prevention and surveillance strategies based on DNA testing, even for mutation carriers." They do acknowledge potential benefits could include enhanced motivation to adhere to prevention and screening guidelines, earlier melanoma diagnosis if the biopsy threshold is lower, and lower anxiety for those who learn they are negative for a known family mutation.²

- The National Comprehensive Cancer Network (NCCN) Melanoma Guideline (updated 2017) includes family history as a melanoma risk factor and alters management based on this risk. However, these guidelines do not address genetic testing for FMM.¹

Criteria
- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  - In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References


Flow Cytometry

Introduction

Flow cytometry testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Flow cytometry, cell cycle or DNA analysis</td>
<td>88182</td>
</tr>
<tr>
<td>Flow cytometry, cell surface, cytoplasmic, or nuclear marker, technical component only; first marker</td>
<td>88184</td>
</tr>
<tr>
<td>Flow cytometry, cell surface, cytoplasmic, or nuclear marker, technical component only; each additional marker (List separately in addition to code for first marker)</td>
<td>88185</td>
</tr>
<tr>
<td>Flow cytometry, interpretation; 2 to 8 markers</td>
<td>88187</td>
</tr>
<tr>
<td>Flow cytometry, interpretation; 9 to 15 markers</td>
<td>88188</td>
</tr>
<tr>
<td>Flow cytometry, interpretation; 16 or more markers</td>
<td>88189</td>
</tr>
</tbody>
</table>

What is flow cytometry

Definition

Flow cytometry is a method that counts the number and proportion of cells with a variety of characteristics, including cell surface or cytoplasmic stains/antibodies for specific biomarkers, as they move single file through a laser beam. Specimens are most commonly fluids such as blood or bone marrow, but it is also possible to test solid samples.
Flow cytometry procedure coding

The following combination(s) of CPT codes may be used unless more specific CPT codes exist (e.g., 86355-86367, 86828-86835). Any deviation from these CPT coding standards is subject to review and denial if not properly coded.

- 88184 is used to describe the technical component of the first marker applied (maximum one unit).
- 88185 is used for each additional marker applied and billed with the applicable number of units. Therefore, 88185 should not be billed without 88184.
- Because these two codes describe only the technical component, there are three other interpretation codes that may be applied based on the number of markers assessed (each billed with a maximum of one unit):
  - 88187 for evaluating 2 to 8 markers
  - 88188 for evaluating 9 to 15 markers
  - 88189 for evaluating 16 or more markers

Common uses

A variety of disorders are associated with distinct biomarker patterns, which can be used to diagnose or subtype these disorders. The following are common uses of flow cytometry in medicine.

Hematopoietic neoplasm evaluation and monitoring

Hematopoietic neoplasm evaluation and monitoring is the most common use, which includes leukemia and lymphoma phenotyping and minimal residual disease detection. Minimal residual disease detection is when individuals with acute leukemia appear to be in remission having levels of disease below detection on bone marrow samples but detectable through flow cytometry. Initial evaluation is generally limited and extensive flow cytometry evaluation should be reserved for those cases with a higher likelihood of neoplasia based on initial evaluation.

HIV infection monitoring

Flow cytometry is used for HIV infection monitoring to accurately and reliably evaluate the number of CD4 positive T cells.

Organ transplantation matching and monitoring

Flow cytometry is used for organ transplantation matching and monitoring. It identifies problematic antibodies and evaluates biomarkers and activation antigens that can distinguish transplant rejection from other issues.
**Immunodeficiencies**

Flow cytometry is used for immunodeficiencies, which may be associated with absent or impaired cell proteins (primary disease), leukocyte dysfunction, and markers of immune status in lymphocytes (secondary disease).¹

**Autoimmune conditions**

Autoimmune conditions may be associated with certain characteristic, detectable auto-antibodies.¹

**Paroxysmal nocturnal hemoglobinuria**

Paroxysmal nocturnal hemoglobinuria (PNH), a rare stem cell disorder, is diagnosed through the detection of deficient proteins by flow cytometry.¹

**Criteria**

**Introduction**

Requests for flow cytometry testing are reviewed using these criteria. This guideline addresses common clinical applications of flow cytometry-based tests that are billed using CPT codes 88184-88189. It is not intended to encompass flow cytometry-based tests billed using more specific CPT codes (e.g., 86355-86367, 86828-86835).

**Hematopoietic Neoplasm Evaluation and Monitoring**

*Medical necessity requirements:*

Because the flow cytometry markers used to evaluate a sample are necessarily different based on clinical indication, information from other evaluations (e.g., morphology), sample type, and the laboratory setting, this guideline addresses general principles of marker panel selection.²

- In the initial evaluation of suspected hematopoietic neoplasm:
  - Common non-neoplastic causes of the clinical presentation (e.g., infection or asplenia with leukocytosis, etc.) should be reasonably ruled out before flow cytometry is employed.
  - A limited but sufficient number of markers should be used in the initial evaluation that allows identification of all major categories of neoplasia (B, T, myeloid, or plasma cell lineages) under consideration based on the clinical indication.
Testing with additional markers is indicated to further characterize disease when the initial evaluation is suggestive.

- For staging or evaluating residual disease in patients with a known diagnosis of hematopoietic neoplasm, a limited panel of markers characteristic of that neoplasm should be used.

**Billing and reimbursement considerations:**

Most presentations, even non-specific indications that require evaluation of several lineages (e.g., anemia, thrombocytopenia, etc.), should rarely require more than 23 flow cytometry markers and monitoring of a known hematopoietic neoplasia diagnosis requires fewer. Therefore:

- In addition to the one marker represented by CPT 88184, reimbursement will routinely be limited to 22 units of CPT 88185.
- ICD code information may be compared with units billed to identify cases with possible excess units that will require post-service medical necessity review. Expected unit number is based on the required cell lineage evaluation by medical indication outlined in the 2006 Bethesda flow cytometry guidelines.²
- When a laboratory routinely bills more than an average of 14 markers, claims from that laboratory will be subject to post-service medical necessity review.

**HIV Monitoring**

**Medical necessity requirements:**

- Flow cytometry is an important method for determining the percentage of lymphocytes that express antigens used to identify CD4+ T cells, and to directly measure absolute T cell counts in the case of single-platform technology (SPT).³
- Four antibodies are routinely required (CD45, CD3, CD4, CD8), which may be applied in three- or four-color antibody panels.
- For pediatric patients, additional antibodies may be required to determine CD19+ B-cell values, which is an indicator of immune status in this population.

**Billing and reimbursement considerations:**

- The most commonly required flow cytometry studies for HIV are represented by marker-specific CPT codes (e.g., 86355-86367). The non-specific flow cytometry codes should not be used when a more specific code exists.
- Therefore, the non-specific CPT codes addressed in this policy should not routinely be required for HIV monitoring. Post-service medical necessity review may be employed when such codes are used for HIV monitoring as indicated by the following ICD codes:
Non-Reimbursed Clinical Indications

Medical necessity requirements:
Flow cytometry procedures will not be reimbursed for the evaluation of the following indications:

- Detection of sexually transmitted organisms, such as human papillomavirus
- Hypertension or cardiovascular disease risk

Billing and reimbursement considerations:
Flow cytometry will not be reimbursed when billed with any of the following ICD codes:

- ICD10 Codes:
  - ICD10 Codes Indicating Testing for STIs on page 374
  - ICD10 Codes Indicating Testing for Hypertension or Cardiovascular Disease Screening on page 377

Other Clinical Indications

Medical Necessity Requirements:
Flow cytometry has a variety of applications that cannot all be adequately addressed by this guideline. All flow cytometry studies must be performed for well validated and medically necessary indications.

Billing and reimbursement considerations:
When flow cytometry is billed with ICD codes that do not suggest one of the other clinical indications addressed in this policy, post-service medical necessity review may be employed. See the Reimbursement Policy that addresses Post-Service Medical Necessity Review for more information.

ICD10 Codes

ICD10 codes in this section may be used to support or refute medical necessity as described in the above criteria.

ICD10 Codes Indicating HIV Positive Status

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>B20</td>
<td>Human immunodeficiency virus [HIV] disease</td>
</tr>
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</table>
### ICD10 Code or Range

<table>
<thead>
<tr>
<th>Code or Range</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>B97.35</td>
<td>Human immunodeficiency virus, type 2 [HIV-2]</td>
</tr>
<tr>
<td>O98.7X</td>
<td>Human immunodeficiency virus [HIV] disease complicating pregnancy, childbirth and the puerperium</td>
</tr>
<tr>
<td>R75</td>
<td>Inconclusive laboratory evidence of human immunodeficiency virus [HIV]</td>
</tr>
<tr>
<td>Z21</td>
<td>Asymptomatic human immunodeficiency virus [HIV] infection status</td>
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</table>

### ICD10 Codes Indicating Testing for STIs

<table>
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<th>Description</th>
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<td>Congenital syphilis</td>
</tr>
<tr>
<td>A51.X</td>
<td>Early syphilis</td>
</tr>
<tr>
<td>A52.X</td>
<td>Late syphilis</td>
</tr>
<tr>
<td>A53.X</td>
<td>Other and unspecified syphilis</td>
</tr>
<tr>
<td>A54.X</td>
<td>Gonococcal infection</td>
</tr>
<tr>
<td>A55</td>
<td>Chlamydial lymphogranuloma (venereum)</td>
</tr>
<tr>
<td>A56.X</td>
<td>Other sexually transmitted chlamydial diseases</td>
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<tr>
<td>A57</td>
<td>Chancroid</td>
</tr>
<tr>
<td>A58</td>
<td>Granuloma inguinale</td>
</tr>
<tr>
<td>A59.X</td>
<td>Trichomoniasis</td>
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<tr>
<td>A60.X</td>
<td>Anogenital herpesviral [herpes simplex] infections</td>
</tr>
<tr>
<td>A63.X</td>
<td>Other predominantly sexually transmitted diseases, not elsewhere classified</td>
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<tr>
<td>A64</td>
<td>Unspecified sexually transmitted disease</td>
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<td>A74.89</td>
<td>Other chlamydial diseases</td>
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<td>A74.9</td>
<td>Chlamydial infection, unspecified (includes childbirth and postpartum)</td>
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<tr>
<td>B37.3</td>
<td>Candidiasis of vulva and vagina</td>
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<tr>
<td>B37.4X</td>
<td>Candidiasis of other urogenital sites</td>
</tr>
<tr>
<td>B97.7</td>
<td>Papillomavirus as the cause of diseases classified elsewhere</td>
</tr>
<tr>
<td>ICD10 Code or Range</td>
<td>Description</td>
</tr>
<tr>
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<td>-------------</td>
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<tr>
<td>L29.3</td>
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<tr>
<td>M02.30</td>
<td>Reiter's disease, unspecified site</td>
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<tr>
<td>N34.X</td>
<td>Urethritis and urethral syndrome</td>
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<td>N35.111</td>
<td>Postinfective urethral stricture, not elsewhere classified, male, meatal</td>
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<tr>
<td>N37</td>
<td>Urethral disorders in diseases classified elsewhere</td>
</tr>
<tr>
<td>N39.0</td>
<td>Urinary tract infection, site not specified</td>
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<tr>
<td>N39.9</td>
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<td>N70.X</td>
<td>Salpingitis and oophoritis</td>
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<td>N71.X</td>
<td>Inflammatory disease of uterus, except cervix</td>
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<tr>
<td>N72</td>
<td>Inflammatory disease of cervix uteri</td>
</tr>
<tr>
<td>N73.X</td>
<td>Other female pelvic inflammatory diseases</td>
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<tr>
<td>N74</td>
<td>Female pelvic inflammatory disorders in diseases classified elsewhere</td>
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<tr>
<td>N75.X</td>
<td>Diseases of Bartholin's gland</td>
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<tr>
<td>N76.X</td>
<td>Other inflammation of vagina and vulva</td>
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<tr>
<td>N77.X</td>
<td>Vulvovaginal ulceration and inflammation in diseases classified elsewhere</td>
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<tr>
<td>N94.1</td>
<td>Dyspareunia</td>
</tr>
<tr>
<td>O09.X</td>
<td>Supervision of high risk pregnancy</td>
</tr>
<tr>
<td>O23.X</td>
<td>Infections of genitourinary tract in pregnancy</td>
</tr>
<tr>
<td>O86.1X</td>
<td>Other infection of genital tract following delivery</td>
</tr>
<tr>
<td>O86.2X</td>
<td>Urinary tract infection following delivery</td>
</tr>
<tr>
<td>R87.5</td>
<td>Abnormal microbiological findings in specimens from female genital organs</td>
</tr>
<tr>
<td>R87.6X</td>
<td>Abnormal cytological findings in specimens from female genital organs</td>
</tr>
<tr>
<td>R87.8X</td>
<td>Other abnormal findings in specimens from female genital organs</td>
</tr>
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<td>Z00.00</td>
<td>Encounter for general adult medical</td>
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<tr>
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<td>Description</td>
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<td>---------------------</td>
<td>-------------</td>
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<tr>
<td>Z00.8</td>
<td>Encounter for other general examination</td>
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<tr>
<td>Z01.4X</td>
<td>Encounter for gynecological examination</td>
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<td>Z11.3</td>
<td>Encounter for screening for infections with a predominantly sexual mode of transmission</td>
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<tr>
<td>Z11.51</td>
<td>Encounter for screening for human papillomavirus (HPV)</td>
</tr>
<tr>
<td>Z11.59</td>
<td>Encounter for screening for other viral diseases</td>
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<tr>
<td>Z11.8</td>
<td>Encounter for screening for other infectious and parasitic diseases</td>
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<td>Z11.9</td>
<td>Encounter for screening for infectious and parasitic diseases, unspecified</td>
</tr>
<tr>
<td>Z12.4</td>
<td>Encounter for screening for malignant neoplasm of cervix</td>
</tr>
<tr>
<td>Z20.2</td>
<td>Contact with and (suspected) exposure to infections with a predominantly sexual mode of transmission</td>
</tr>
<tr>
<td>Z20.6</td>
<td>Contact with and (suspected) exposure to human immunodeficiency virus [HIV]</td>
</tr>
<tr>
<td>Z20.818</td>
<td>Contact with and (suspected) exposure to other bacterial communicable diseases</td>
</tr>
<tr>
<td>Z20.828</td>
<td>Contact with and (suspected) exposure to other viral communicable diseases</td>
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<tr>
<td>Z20.89</td>
<td>Contact with and (suspected) exposure to other communicable diseases</td>
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<td>Contact with and (suspected) exposure to unspecified communicable disease</td>
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<td>Z30.X</td>
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</tr>
<tr>
<td>Z31.X</td>
<td>Encounter for procreative management</td>
</tr>
<tr>
<td>Z32.X</td>
<td>Encounter for pregnancy test and childbirth and childcare instruction</td>
</tr>
<tr>
<td>Z33.X</td>
<td>Pregnant state</td>
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<tr>
<td>Z34.X</td>
<td>Encounter for supervision of normal pregnancy</td>
</tr>
</tbody>
</table>
### ICD10 Code or Range

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Z36</td>
<td>Encounter for antenatal screening of mother</td>
</tr>
<tr>
<td>Z39.X</td>
<td>Encounter for maternal postpartum care and examination</td>
</tr>
<tr>
<td>Z64.0</td>
<td>Problems related to unwanted pregnancy</td>
</tr>
<tr>
<td>Z64.1</td>
<td>Problems related to multiparity</td>
</tr>
<tr>
<td>Z71.7</td>
<td>Human immunodeficiency virus [HIV] counseling</td>
</tr>
<tr>
<td>Z72.5X</td>
<td>High risk sexual behavior</td>
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<tr>
<td>Z77.9</td>
<td>Other contact with and (suspected) exposures hazardous to health</td>
</tr>
<tr>
<td>Z97.5</td>
<td>Presence of (intrauterine) contraceptive device</td>
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</tbody>
</table>

### ICD10 Codes Indicating Testing for Hypertension or Cardiovascular Disease Screening

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
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<tbody>
<tr>
<td>I10</td>
<td>Essential (primary) hypertension</td>
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<tr>
<td>I11.X</td>
<td>Hypertensive heart disease</td>
</tr>
<tr>
<td>I12.X</td>
<td>Hypertensive chronic kidney disease</td>
</tr>
<tr>
<td>I13.X</td>
<td>Hypertensive heart and chronic kidney disease</td>
</tr>
<tr>
<td>I15.X</td>
<td>Secondary hypertension</td>
</tr>
<tr>
<td>I20.X</td>
<td>Angina pectoris</td>
</tr>
<tr>
<td>I21.X</td>
<td>ST elevation (STEMI) and non-ST elevation (NSTEMI) myocardial infarction</td>
</tr>
<tr>
<td>I22.X</td>
<td>Subsequent ST elevation (STEMI) and non-ST elevation (NSTEMI) myocardial infarction</td>
</tr>
<tr>
<td>I23.X</td>
<td>Certain current complications following ST elevation (STEMI) and non-ST elevation (NSTEMI) myocardial infarction (within the 28 day period)</td>
</tr>
<tr>
<td>I24.X</td>
<td>Other acute ischemic heart diseases</td>
</tr>
<tr>
<td>I25.X</td>
<td>Chronic ischemic heart disease</td>
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<tr>
<td>R07.2</td>
<td>Precordial pain</td>
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<tr>
<td>R07.8X</td>
<td>Other chest pain</td>
</tr>
<tr>
<td>ICD10 Code or Range</td>
<td>Description</td>
</tr>
<tr>
<td>---------------------</td>
<td>-------------</td>
</tr>
<tr>
<td>R07.9</td>
<td>Chest pain, unspecified</td>
</tr>
<tr>
<td>R09.89</td>
<td>Other specified symptoms and signs involving the circulatory and respiratory systems</td>
</tr>
<tr>
<td>R94.3X</td>
<td>Abnormal results of cardiovascular function studies</td>
</tr>
<tr>
<td>Z13.6</td>
<td>Encounter for screening for cardiovascular disorders</td>
</tr>
<tr>
<td>Z82.4X</td>
<td>Family history of ischemic heart disease and other diseases of the circulatory system</td>
</tr>
</tbody>
</table>

**References**

**Introduction**

These references are cited in this guideline.


Fragile X Associated Tremor/Ataxia Syndrome Testing

MOL.TS.171.A
v2.0.2018

Procedures addressed

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<table>
<thead>
<tr>
<th>Procedure addressed by this guideline</th>
<th>Procedure code</th>
</tr>
</thead>
<tbody>
<tr>
<td>FMR1 Expansion Analysis</td>
<td>81243</td>
</tr>
</tbody>
</table>

What is fragile X-associated tremor/ataxia syndrome

Definition

Fragile X-associated tremor/ataxia syndrome (FXTAS) is a neurodegenerative disorder characterized by progressive cerebellar ataxia and/or intention tremor usually presenting after age 50 in individuals with a premutation allele in the gene for fragile X (FMR1).¹

- Fragile X syndrome, FXTAS, and other related disorders are caused by a type of genetic mutation called a triplet repeat. A triplet repeat is a sequence of three nucleotide building blocks (CGG) that is variably repeated within the FMR1 gene. The number of triplet repeats determines whether the gene is normal, intermediate, or has a premutation or full mutation.³,⁴ Premutation carriers — the group at risk for FXTAS — have 55 to 200 CGG repeats.¹
- Both male and female premutation carriers are at risk for FXTAS. Approximately 40% of males over the age of 50, with a premutation allele, will develop FXTAS. The risk to female premutation carriers appears to be lower.¹,²
- Other neurologic findings of FXTAS include:¹
  - Short term memory loss
  - Executive function deficits
  - Cognitive decline
  - Dementia
  - Parkinsonism
  - Peripheral neuropathy
Lower limb proximal weakness

A diagnosis is confirmed by the presence of a FMR1 premutation and white matter lesions on MRI in the middle cerebellar peduncles and/or brain stem, with intention tremor and/or gait ataxia.¹

Test information

FMR1 CGG expansion analysis measures the number of CGG repeat copies within the FMR1 gene. Repeat number classifies results as normal, intermediate, premutation, or full mutation.²,³ The same analysis can be used for diagnostic, carrier, and prenatal testing.

Guidelines and evidence

Consensus guidelines from the American College of Medical Genetics (ACMG, 2005) recommend FXTAS testing for the following people:

- Men and women who are experiencing late onset intention tremor and cerebellar ataxia of unknown origin, especially if they have (a) a family history of movement disorders, (b) a family history of fragile X syndrome, or (c) male or female relatives with undiagnosed mental retardation.”³

Evidence-based guidelines from the European Federation of Neurological Societies (EFNS, 2010) state:

- “Recommendations for FXTAS genetic testing: Genetic testing for the X-linked FXTAS is recommended when there is a clinical suspicion, and it is readily available in many laboratories (Class B).”⁴ [Class B rating = “(probably effective, ineffective, or harmful) requires at least one convincing class II study or overwhelming class III evidence”⁵]

Criteria

Targeted mutation analysis for CGG trinucleotide repeat expansion in FMR1

- Genetic Counseling:
  - Medical evaluation by a physician familiar with FXTAS, and
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous molecular genetic testing of FMR1, AND

- Diagnostic Testing for Symptomatic Individuals:
• Intellectual disability (ID), or
  • Males and females ≥50 years with progressive intention tremor and cerebellar ataxia of unknown origin, OR

• Prenatal Testing for At-Risk Pregnancies
  • CGG trinucleotide repeat expansion in FMR1 identified in biological mother,** OR

• Carrier Screening and Predictive Testing for Presymptomatic/Asymptomatic At Risk Individuals:
  • Known CGG trinucleotide repeat expansion in FMR1 in 1st, 2nd, or 3rd degree biologic relative, or
  • Personal or family history of premature ovarian failure (cessation of menses before age of 40 years), or
  • Family history of movement disorder, and
    ▪ Cerebellar ataxia has been ruled out, and
    ▪ Other movement disorders have been ruled out, or
  • Family history of intellectual disability with an unknown cause, or
  • Prior cytogenetic test suspicious for Fragile X, and
  • Age 18 years or older
  • Intellectual disability, AND

• Possibility of X-linked inheritance has not been ruled out by male to male transmission

** Note: CVS must be interpreted with caution. The number of CGG repeats in the fetus can be accurately determined; however, often the methylation status of FMR1 is not yet established in chorionic villi at the time of sampling. CVS results may lead to a situation in which follow-up amniocentesis is necessary to resolve an ambiguous result.

References


Fragile X Syndrome Testing

Procedures addressed

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What is fragile X syndrome

Definition

Fragile X syndrome is the most common cause of inherited intellectual disability affecting approximately 1 in 4,000 males and 1 in 8,000 females.\(^1\)\(^2\) Because the mutation is on the X-chromosome, males tend to be more severely affected than females.

- Symptoms vary widely and may include the following:\(^1\)\(^2\)
  - Intellectual disability
  - Autism
  - Large head
  - Long face
  - Prominent forehead and chin
  - Protruding ears
  - Loose joints
  - Large testes in postpubertal males
  - Motor and language delays
o Behavioral differences

- Fragile X syndrome is caused by a type of genetic mutation called a triplet repeat. A triplet repeat is a sequence of three nucleotide building blocks (CGG) that is variably repeated within the FMR1 gene. A full mutation (>200 repeats) usually causes the gene to be abnormally methylated, turning it off. The number of CGG repeat copies within the FMR1 gene can expand from one generation to the next, a property known as anticipation.\(^2,3\)

- Predictive (carrier) testing can be performed for at-risk relatives when there is a family history of fragile X, intellectual disability of unknown etiology, or some other characteristic conditions.\(^3\)

- A woman carrying a premutation or full mutation is at risk to have a child affected with fragile X. The actual risk depends on the number of repeats in her FMR1 gene.\(^1,4\) Prenatal testing is available for pregnancies at-risk.

Test information

- FMR1 CGG expansion analysis measures the number of CGG repeat copies within the FMR1 gene. Repeat number classifies results as normal, intermediate, premutation, or full mutation.\(^2,3\) The same analysis can be used for diagnostic, carrier, and prenatal testing.

- FMR1 CGG methylation analysis is typically assessed in those with a premutation or full mutation.\(^1,5\) Abnormal methylation, causing a disruption in FMR1 protein production, is the mechanism responsible for features of Fragile X. Non-classic presentations due to size and methylation mosaicism have been reported.

- Prenatal diagnosis must be undertaken with caution. Expansion analysis is equally accurate on fetal samples from amniocentesis and CVS. However, methylation analysis on a CVS sample may yield an ambiguous result and amniocentesis may be needed for follow up.\(^5\)

Guidelines and evidence

- Consensus guidelines from the American Academy of Pediatrics (AAP, 2011) that address health supervision of fragile X syndrome:
  
  - "Because children with fragile X syndrome may not have apparent physical features, any child who presents with developmental delay, borderline intellectual abilities, or mental retardation or has a diagnosis of autism without a specific etiology should undergo molecular testing for fragile X syndrome to determine the number of CGG repeats (Fig 1). Fragile X testing should also be considered in patients in whom there is suspected, but not molecularly proven, Sotos syndrome or Prader-Willi syndrome. On the other hand, fragile X testing, is not routinely warranted for children with isolated attention-deficit/hyperactivity disorder."\(^6\)
• Practice guidelines from the American College of Medical Genetics (ACMG, 2005) recommend diagnostic testing for fragile X syndrome for “Individuals of either sex with mental retardation, developmental delay, or autism, especially if they have (a) any physical or behavioral characteristics of fragile X syndrome, (b) a family history of fragile X syndrome, or (c) male or female relatives with undiagnosed mental retardation.”

• Practice guidelines from the American College of Medical Genetics (ACMG, 2005) and the American College of Obstetricians and Gynecologists (ACOG, 2010) support carrier screening for fragile X syndrome:
  - ACMG: “Individuals seeking reproductive counseling who have (a) a family history of fragile X syndrome or (b) a family history of undiagnosed mental retardation.”
  - ACOG: “Women with a family history of fragile X-related disorders, unexplained mental retardation or developmental delay, autism, or premature ovarian insufficiency are candidates for genetic counseling and fragile X premutation carrier screening.”

• Practice guidelines from the American College of Medical Genetics (ACMG, 2005) and the American College of Obstetricians and Gynecologists (ACOG, 2010) support prenatal screening for fragile X syndrome:
  - ACMG: Fragile X testing is appropriate in “Fetuses of known carrier mothers.”
  - ACOG: “Prenatal testing for fragile X syndrome by amniocentesis or CVS should be offered to known carriers of the fragile X premutation or full mutation. Although amniocentesis and CVS are reliable for determining the number of triplet repeats, CVS may not adequately determine the methylation status of the FMR1 gene.”

Criteria

Targeted mutation analysis for CGG trinucleotide repeat expansion in FMR1

• Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  - No previous molecular genetic testing of FMR1, AND

• Diagnostic Testing for Symptomatic Individuals:
  - Males and females with unexplained speech and/or language delay, motor development delay, intellectual disability (ID), or autism, or
- Female with premature ovarian failure (cessation of menses before age of 40 years), or
- Males and females ≥50 years with progressive intention tremor and cerebellar ataxia of unknown origin, OR

- Prenatal Testing for At-Risk Pregnancies:
  - CGG trinucleotide repeat expansion in FMR1 identified in biologic mother,** OR

- Carrier Screening and Predictive Testing for Presymptomatic/Asymptomatic At Risk Individuals:
  - Known CGG trinucleotide repeat expansion in FMR1 in 1st, 2nd, or 3rd degree biologic relative, or
  - Family history of premature ovarian failure (cessation of menses before age of 40 years), or
  - Family history of movement disorder and
    - Cerebellar ataxia has been ruled out
    - Other movement disorders have been ruled out, or
  - Family history of undiagnosed intellectual disability, or
  - Prior cytogenetic test suspicious for Fragile X, AND

- Possibility of X-linked inheritance has not been ruled out by male to male transmission

**Methylation analysis**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - CGG expansion analysis result showing a premutation or full allele size (typically greater than 55 repeats), AND

- Diagnostic Testing for Symptomatic Individuals:
  - Males and females with speech and/or language delay, motor development delay, intellectual disability (ID), or autism, or
  - Female with premature ovarian failure (cessation of menses before age of 40 years), or
- Males and females ≥50 years with progressive intention tremor and cerebellar ataxia of unknown origin, OR

- Prenatal Testing for At-Risk Pregnancies:
  - CGG trinucleotide repeat expansion in FMR1 identified in biologic mother**

** Note: CVS must be interpreted with caution. The number of CGG repeats in the fetus can be accurately determined; however, often the methylation status of FMR1 is not yet established in chorionic villi at the time of sampling. CVS results may lead to a situation in which follow-up amniocentesis is necessary to resolve an ambiguous result.

References


Gaucher Disease Testing

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What is Gaucher Disease

Definition

Gaucher disease is a genetic disease that affects multiple organs and tissues.

- There are several types of Gaucher disease, each with varying signs and symptoms:\(^1,2\)
  - Type 1 is the most common type of Gaucher Disease. Unlike other types, type 1 does not affect the central nervous system (CNS). Symptoms include enlargement of the liver and spleen (hepatosplenomegaly), anemia, low blood platelets, lung disease, and bone abnormalities.
  - Type 2/Type 3. These types are rarer, usually more severe, and affect the brain and CNS. Common symptoms include seizures, hyperextension of the spine, and lockjaw, in addition to the symptoms listed above for type 1. Type 2 is more severe, and affected individuals usually do not survive past childhood. Type 3 affected individuals have more slowly progressing symptoms and can survive into adulthood.
  - Perinatal lethal. The most severe form of Gaucher disease has symptoms that begin during pregnancy or in early infancy, including swelling, dry/scaly skin (ichthyosis), and serious neurological problems. Affected infants usually survive only a few days after birth.
  - Cardiovascular. This type has mainly heart manifestations. Symptoms include the hardening of heart valves, eye abnormalities, bone disease, and enlarged spleen.
These subtypes are identified through clinical symptoms and do not correlate well with the different mutations that cause Gaucher disease.2

- Gaucher disease is relatively common in Ashkenazi Jewish populations, affecting about 1 in 500 to 1 in 1,000 people.1 It is much less common in the general population, affecting about 1 in 50,000 to 1 in 100,000 people.1

- Gaucher disease is caused by changes, or mutations to the GBA gene.1-3 The GBA gene makes the enzyme beta-glucosylceramidase, also called acid beta-glucocerebrosidase. This enzyme helps break down fatty substances in cells. Mutations in GBA lead to a buildup of these fatty substances to toxic levels. This buildup damages tissues and organs, leading to symptoms of Gaucher disease.1-3

- Gaucher disease is an autosomal recessive disorder. An affected person inherits two GBA gene mutations -- one from each parent.1,2
  - People who have only one GBA mutation are called carriers. Carriers do not show symptoms of Gaucher disease, but have a 50% chance of passing the mutation on to their children.
  - Two carriers of Gaucher disease have a 25% chance of having a child affected with the disease.

- Clinical findings alone are insufficient for a definitive diagnosis of Gaucher disease.2

- If Gaucher disease is suspected in a symptomatic person, beta-glucosylceramidase enzyme testing should be performed first. People affected with Gaucher disease have 0-15% the normal level of beta-glucosylceramidase compared to healthy individuals. Measuring beta-glucosylceramidase levels is a reliable way to confirm a suspected case of Gaucher disease.2,4,5 Beta-glucosylceramidase levels within the normal range rule out Gaucher disease.

- Genetic testing can be used to identify the disease-causing mutations in an affected person diagnosed by enzyme analysis.1 This is done primarily for reproductive purposes when parents of an affected child need to know the mutations for preimplantation genetic diagnosis or prenatal diagnosis. Mutation analysis can also confirm disease-causing mutations when a diagnosis by enzyme analysis is inconclusive.1 Enzyme testing is not appropriate to identify unaffected carriers.2

**Test information**

- **GBA Mutation Panel:** Clinically-available testing panels look for four or more of most common mutations in the GBA gene.
  - Four mutations (N370S, L444P, 84GG, IVS2+1) account for about 90% of mutations in the Ashkenazi Jewish population and about 50%-60% of mutations in the non-Ashkenazi Jewish population.1
  - Some laboratories include several other common mutations in their panels.
o Carrier screening by GBA mutation panel for Gaucher disease is widely available as part of an "Ashkenazi Jewish Panel" that includes several other genetic disease that are more common in this population. (See Ashkenazi Jewish Carrier Screening for more information.)

- **GBA Sequence Analysis:** This test analyzes the entire coding region of the GBA gene and will find mutations that the GBA mutation panel could not.\(^1\)
  o The detection rate of sequencing is about 99%.
  o This test is indicated in people with Gaucher disease who have one or no mutations identified by mutation panel testing.
  o This test is also indicated for reproductive partners of individuals who have a GBA mutation.

- **GBA known familial mutation testing:** When there is a family history of Gaucher disease, the family mutations should be identified prior to carrier testing in at-risk family members when possible. A mutation panel can be used if the family mutations are included in the panel. If the family mutations are not included in the panel and were identified through sequencing, then GBA known familial mutation testing is necessary.\(^2\)

- **Prenatal or preimplantation genetic diagnosis:** This testing is possible in at-risk pregnancies if the parental mutations are known.

**Guidelines and evidence**

- No US evidence-based diagnostic guidelines have been identified.
- A 2015 expert-authored review recommends the following testing strategy for diagnosis of an affected person:\(^2\)
  o "Assay of glucosylceramidase enzyme activity in leukocytes or other nucleated cells is the confirmatory diagnostic test."
  o "Molecular genetic testing and the identification of two disease-causing alleles provide an alternative means of confirming the diagnosis. There is broad heterogeneity in causative variants; in individuals in whom genetic testing identifies a novel GBA variant, biochemical testing to confirm the diagnosis should be considered."
  o "Targeted analysis for pathogenic variants in a proband originally diagnosed by biochemical testing may be considered for genetic counseling purposes, primarily to identify the pathogenic variants and permit carrier detection among at-risk relatives."
- Reviews published in peer-reviewed medical literature support this and offer some considerations for genotyping:
  o Archives of Internal Medicine (1998):\(^4\)
“The most efficient and reliable method of establishing the diagnosis of Gaucher disease is the assay of β-glucocerebrosidase activity.”
“Knowledge of the genotype may be helpful in predicting the severity and rate of progression of clinical symptoms in patients. For example, the homozygous N370S allele is usually associated with a generally less severe phenotype, although with wide clinical variability; the heterozygous state for N370S is protective against central nervous system involvement; and the L444P allele in the homozygous state is associated with early neurologic symptoms common in the types 2 and 3 clinical classifications.”

The Brazilian Study Group on Gaucher Disease (2009):5

“Definitive diagnosis of [Gaucher disease] requires confirmation by the acid β-glucosidase enzyme assay in leukocytes or fibroblasts.”
“N370S homozygotes generally present with a less severe phenotype, whereas L444P and D409H homozygosity confers neurologic involvement. Despite these general genotype-phenotype correlations, disease severity, and clinical outcomes cannot be predicted on the basis of genotype.”

Professional guidelines generally support Gaucher disease carrier screening for those at increased risk.6,7

Consensus guidelines from the American College of Obstetricians and Gynecologists (ACOG, 2009) address carrier screening and prenatal diagnosis for Gaucher disease:6

“Individuals with a positive family history of one of these disorders [including Gaucher disease] should be offered carrier screening for the specific disorder and may benefit from genetic counseling.”
Carrier screening for Ashkenazi Jewish people is routinely recommended for some disorders (i.e., Tay-Sachs, Canavan, cystic fibrosis, familial dysautonomia). However, for testing of a group of other disorders more common in this population (including Gaucher disease), ACOG simply states: “Individuals of Ashkenazi Jewish descent may inquire about the availability of carrier screening for other disorders.”
“If it is determined that this individual [an Ashkenazi Jewish descent partner] is a carrier, the other partner should be offered screening.”
o “When both partners are carriers of one of these disorders, they should be referred for genetic counseling and offered prenatal diagnosis.”

• Consensus guidelines from the American College of Medical Genetics (2008) recommend routine carrier screening for a group of disorders that includes Gaucher disease when at least one member of the couple is Ashkenazi Jewish and that couple is pregnant or planning pregnancy.”

Criteria
Carrier Testing

GBA Known Familial Mutation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous molecular genetic testing of GBA, AND

• Carrier Screening:
  o GBA mutation(s) identified in 1st, 2nd, or 3rd degree biologic relative(s), OR

• Prenatal Testing for At-Risk Pregnancies:
  o GBA mutation(s) identified in both biologic parents.

GBA Targeted Mutation Analysis for Ashkenazi Mutations (Four Mutations)

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous GBA genetic testing, including Ashkenazi Jewish screening panels containing targeted mutation analysis for Gaucher disease, AND

• Carrier Screening:
  o Ashkenazi Jewish descent, regardless of disease status and results of glucosylceramidase assay, and
  o Intention to reproduce

Diagnostic and Expanded Carrier Testing

GBA Sequencing

• Genetic Counseling:
• Pre and post-test genetic counseling by an appropriate provider (as deemed by
the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous GBA full sequencing analysis, and
  o If Ashkenazi Jewish, testing for 4 common mutations is negative, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Glucosylceramidase enzyme activity in peripheral blood leukocytes is 0-15% of
    normal activity, and
  o Characteristic bone changes including osteopenia, focal lytic or sclerotic bone
    lesions or osteonecrosis, or
  o Hepatosplenomegaly and hematologic changes including anemia or
    thrombocytopenia, or
  o Primary neurologic disease which could include one or more of the following:
    cognitive impairment, bulbar signs, pyramidal signs, oculomotor apraxia, or
    seizures (progressive myoclonic epilepsy), OR

• Diagnostic Testing for Asymptomatic Carriers:
  o One mutation detected by targeted mutation analysis, and
  o Glucosylceramidase enzyme activity in peripheral blood leukocytes is 0-15% of
    normal activity, OR

• Testing for Individuals with Family History or Partners of Carriers:
  o 1st, 2nd, or 3rd degree biologic relative with Gaucher disease clinical diagnosis,
    family mutation unknown and testing unavailable, or
  o Partner is monoallelic or biallelic for GBA mutation, and has the potential and
    intention to reproduce with this partner.

References

   Genetic Conditions (database online). A Service of the US National Library of

2. Pastore GM, Hughes DA. (Updated February 2015). Gaucher Disease. In:
   online). Copyright, University of Washington, Seattle. 1997-2016. Available at:


Gene Expression Profiling Tests for Prostate Cancer

Procedures addressed

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What are gene expression profiling tests for prostate cancer

Definition

Prostate cancer (PC) is the most common cancer and a leading cause of cancer-related deaths worldwide. It is considered a heterogeneous disease with highly variable prognosis.¹

- High-risk prostate cancer (PC) patients treated with radical prostatectomy (RP) undergo risk assessment to assess future disease prognosis and determine optimal treatment strategies. Post-RP pathology findings, such as disease stage, baseline Gleason score, time of biochemical recurrence (BCR) after RP, and PSA doubling-time, are considered strong predictors of disease-associated metastasis and mortality. Following RP, up to 50% of patients have pathology or clinical features that are considered at high risk of recurrence and these patients usually undergo post-RP treatments, including adjuvant or salvage therapy or radiation therapy, which can have serious risks and complications. According to clinical practice guideline recommendations, high risk patients should undergo 6 to 8 weeks of radiation therapy (RT) following RP. However, approximately 90% of high-risk patients do not develop metastases or die of prostate cancer, and instead may be appropriate candidates for alternative treatment approaches, including active surveillance. As such, many patients may be subjected to unnecessary follow-up procedures and their associated complications, highlighting the need for improved methods of prognostic risk assessment.²,³
Several genomic biomarkers have been commercially developed to augment the prognostic ability of currently available routine clinical and pathological tests and identify those patients most and least likely to benefit from a specific treatment strategy. Prognostic genomic tests, including gene expression profiling tests, may help to avoid overtreatment by reclassifying those men originally identified as high risk, but who are unlikely to develop metastatic disease. Genomic biomarkers may also play a role in assisting clinicians to tailor personalized and more appropriate treatments for subgroups of PC patients, and improve overall health outcomes.¹²³

Four commercial genomic biomarker tests for assessing risk of PC recurrence and guiding treatment decisions in prostate cancer will be addressed in this guideline: Decipher Prostate Cancer Classifier, OncotypeDX Genomic Prostate Score, ProLaris, and ProMark Proteomic Prognostic Test.

Test information

Gene expression profiles (GEPs) evaluate the expression of several genes using one sample. Gene expression is determined through RNA analysis, using either reverse transcriptase (RT) polymerase chain reaction (PCR) or DNA microarrays.⁴

- Decipher® Prostate Cancer Classifier (GenomeDX Biosciences, Inc.)⁵
  - According to the manufacturer, the Decipher test is a tissue-based tumor genomic test that predicts the probability of metastasis within 5 years of RP, and provides an independent assessment of tumor aggressiveness, information that is distinct from that provided by the Gleason score or PSA.
  - Decipher analyzes a small tissue sample removed during surgery that is routinely archived or stored by the pathology lab. This test is intended for PC patients with stage T2 disease with positive margins, stage T3 disease, or rising serum PSA after RP. The test evaluates the expression of 1.4M RNA (44,000 genes) using RNA extracted from formalin-fixed paraffin-embedded (FFPE) tumor specimens of the index lesion, defined as the highest tumor stage or Gleason score.
  - The Decipher test result is expressed as a continuous risk score; a genomic classifier (GC) that ranges from 0 (lowest) to 1 (highest). Each score is associated with the probability of 5-year metastasis.

- Oncotype DX® Genomic Prostate Score (GPS) (Genomic Health)⁶
  - According to the manufacturer, Oncotype DX prostate cancer assay is a multi-gene expression profiling assay that produces a genomic prostate score (GPS), ranging from 0-100, representing tumor aggressiveness. The Oncotype DX GPS provides risk stratification to properly classify patients. This test is designed to help patients with newly diagnosed, early-stage PC make informed treatment decisions, including active surveillance.
Oncotype DX GPS uses quantitative RT-PCR for 12 prostate cancer-related genes and 5 control genes (total of 17 genes). It was developed for use with fixed paraffin-embedded (FPE) diagnostic prostate needle biopsies (≥1 mm prostate tumor).

- Prolaris® (Myriad® Genetics)
  - According to the manufacturer, Prolaris is a genomic test developed to predict PC-specific mortality in PC patients after needle biopsy, as well as post-RP patients to assess the risk of BCR. This test is designed to assist clinicians with predicting tumor aggressiveness combined with clinical and pathologic variables (Gleason score, PSA).

- ProMark Proteomic Prognostic Test (Metamark®)
  - According to the manufacturer, ProMark uses an 8-protein signature to predict PC aggressiveness (adverse prostate pathology of Gleason ≥ 4+3 and/or non-organ confined disease [T3a, T3b, N1, or M1]) in patients with biopsy Gleason Scores of 3+3 and 3+4. It is designed to provide a personalized prediction regarding if PC can be managed with or without aggressive forms of treatment.
  - ProMark scores range from 0 to 1 score, reflecting the probability of adverse pathology at radical prostatectomy.

Guidelines and evidence

- The National Comprehensive Cancer Network (NCCN) 2017 Clinical Practice Guidelines on Prostate Cancer state the following regarding molecular assays:  
  - “Men with clinically localized disease may consider the use of tumor-based molecular assays. Retrospective case cohort studies have shown that molecular assays performed on biopsy or prostatectomy specimens provide prognostic information independent of NCCN risk groups. These include, but are not limited to, likelihood of death with conservative management, likelihood of biochemical progression after radical prostatectomy or external beam therapy, and likelihood of developing metastasis after radical prostatectomy or salvage radiotherapy.”
  - “Several tissue-based molecular assays have been developed in an effort to improve decision-making in newly diagnosed men considering active surveillance and in treated men considering adjuvant therapy or treatment for recurrence.”
  - “Uncertainty about the risk of disease progression can be reduced if such molecular assays can provide accurate and reproducible prognostic or predictive information beyond NCCN risk group assignment and currently available life expectancy tables and nomograms. Retrospective case cohort studies have shown that these assays provide prognostic information independent of NCCN risk groups, which include likelihood of death with..."
conservative management, likelihood of biochemical recurrence after radical prostatectomy or radiotherapy, and likelihood of developing metastasis after operation or salvage radiotherapy. No randomized controlled trials have studied the utility of these tests."

- According to NCCN, the Molecular Diagnostic Services Program (MoIDX) recommendations stated the following:
  - Decipher: “Cover post-RP for 1) pT2 with positive margins; 2) any pT3 disease; 3) rising PSA (above nadir)”
  - Prolaris: “Cover post-biopsy for NCCN very-low and low-risk prostate cancer at diagnosis with at least 10 years life expectancy.”
  - Oncotype DX: “Cover post-biopsy for NCCN very-low and low-risk prostate cancer at diagnosis with at least 10-20 years life expectancy.”
  - ProMark: Not reviewed

- Decipher Literature Review

  - There is currently limited evidence in the peer-reviewed literature to support the widespread use of the Decipher tests to accurately provide prognostic risk stratification among patients with prostate cancer who have undergone RP in routine clinical practice. The relatively large evidence base, published primarily by the test manufacturer, consists of retrospective case-control and retrospective cohort studies evaluating the strength of the association between the Decipher score and incidence of disease recurrence (e.g., biochemical recurrence, metastasis) or PC-associated mortality. Hazard and odds ratios from univariate and multivariate logistic regression analyses show significant associations between the test and clinical endpoint. Also, study results indicate that Decipher consistently discriminates between men at 5-year risk of metastatic disease progression after RP and men without disease progression with reasonable AUC and c-index estimates. Several studies reported reclassification rates using the Decipher test, indicating that patient risk could be stratified differently based on Decipher results. These type of reclassification calculations are useful since the clinical usefulness of a prognostic test has been reported to be reliant on its ability to categorize patients into different and more accurate prognostic groups, providing accurate predictions about their future disease state, and ultimately guiding optimal treatment regimens. However, these various estimates may be subject to bias and confounders given the several limitations that weaken the quality of the individual studies, including publication bias; patient overlap; insufficient follow-up periods and small number of metastatic event cases; bias associated with retrospective analyses; lack of observer or investigator blinding; missing or flawed registry data; Decipher sampling issues; and considerable heterogeneity between cases and controls for various demographic, disease risk factors, and treatment regimens. Most importantly, it is not clear how results of the Decipher test will impact patient disease management and treatment strategies, and if any changes will translate into improved morbidity and mortality for high-risk PC patients. Ongoing
prospective clinical trials indicate that clinical utility studies of Decipher for PC patients are forthcoming in 2017. Results of new peer-reviewed studies of clinical utility will potentially provide higher quality evidence to better inform clinicians regarding patient selection criteria and appropriate use of the Decipher test among high-risk PC patients who are weighing the risk and benefits of various treatment options.

- Ongoing clinical trials
  - Three ongoing clinical trials, identified on ClinicalTrials.gov are currently undergoing patient recruitment or have completed recruitment and are ongoing.
    - Observational patient registry study (NCT02609269): Decipher Genomics Resource Information Database (GRID)
    - Prospective cohort study sponsored by GenomeDx (estimated completion date, March 2017) to study the influence of the Decipher test on urologist and patient treatment plan choices immediately after RP, and at the time of PSA risk or BCR (NCT02080689): Prospective Clinical Utility Study to Assess the Impact of Decipher on Treatment Decisions After Surgery (PRO-IMPACT).
  - Prolaris Literature Review
    - Although clinical studies suggest that Prolaris may have potential prognostic value following RP in patients with prostate cancer, a number of limitations characterizing the evidence base weaken the strength of these findings. The available studies focused on primarily evaluating associations between results of Prolaris and the incidence of disease recurrence or mortality, which represents a preliminary stage of development of prognostic tests. The most appropriate clinical decisions to be made based on Prolaris test results have not been clearly established since there are no published studies that have reported the ability of the Prolaris test to prospectively predict patient-relevant health outcomes by virtue of prognostic risk assessment or changes made to treatment recommendations. The evidence base may possibly be subject to publication bias. With one exception, the reviewed studies with consistently positive or favorable results were sponsored or funded by the test manufacturer. The single study not funded by the manufacturer reported that 20 of 52 patients were misclassified by the Prolaris test, indicating that use of the test may be misleading in some cases.
    - In some cases, study follow-up was very short, and may was not sufficiently long enough to capture metastatic event data. In addition, the total number of identified cases in each study was relatively small, which limited the power of
the analysis to properly assess diagnostic accuracy. Other flaws in study design that weaken the overall quality of the evidence base include the retrospective design aspect; the lack of investigator or observer blinding; presence of missing or flawed data in registry studies; sampling and specimen processing issues; and the presence of heterogeneity between cases and controls in terms of high risk PC features, and risk factors for metastasis. All of these introduce the possibility of bias and confounding, which weaken the overall confidence in the estimate of prognostic accuracy of the Prolaris test.

- Ongoing clinical trials

  - Two ongoing clinical trials, identified on ClinicalTrials.gov are currently undergoing patient recruitment or have completed recruitment and are ongoing.
    - Behavioral Registry Study to Measure the Impact of Adding Genomic Testing (NCT02454595): Outcome measures: the impact of genomic test results towards selecting a first-line therapy option for newly diagnosed, localized, prostate cancer patients (recruiting).43
    - Open Registry Measuring Impact of Genomic Testing on Treatment Decision after Biopsy in Newly Diagnosed Prostate Cancer Patients (NCT02209584). Outcome measures: Percentage change from the recorded PRE-Prolaris® test treatment option versus the ACTUAL treatment option of genomic risk assessment testing (Prolaris®); Percentage change from the recorded PRE-Prolaris® test treatment option versus the POST-Prolaris® test treatment plan (prior to patient consultation) (active, not recruiting).44
  - OncotypeDX Prostate Literature Review45-48

    - Oncotype DX may be useful to assist RP patients in deciding whether or not they should receive early adjuvant therapy after RP, and may be useful to assist AS patients in predicting the probability of adverse pathology that would help guide the need for subsequent intervention or AS. Oncotype DX was found to predict adverse prostate cancer pathology beyond currently used clinical parameters and nomograms in patients with very low, low-, and intermediate risk disease. Despite these preliminary clinical studies suggesting the potential benefit of Oncotype DX, additional well-designed studies are still needed to adequately determine if the test can allow for clinicians to offer active surveillance safely, thereby minimizing the risk of underestimating the risk of metastasis or local tumor spread. In addition, clinical utility studies in real-world urologic clinical practice are needed to evaluate if treatment practices change with test use, and if these changes result in improved patient-important outcomes.
Ongoing clinical trials

- One ongoing clinical trial, identified on ClinicalTrials.gov is currently undergoing patient recruitment.
  - Engaging Newly Diagnosed Men About Cancer Treatment Options (NCT02668276): Outcome measures: effect of adding Oncotype DX prostate cancer assay to usual counseling (standard NCCN counseling using recommendations based on currently accepted treatment approaches for cancer) on the proportion of men adopting active surveillance. Time frame: 3 to 6 months after making a treatment decision (recruiting).49

- ProMark Literature Review50

  - One clinical validity study suggests that the ProMark risk score offers additional prognostic information for patients compared with NCCN risk categories alone. However, the current evidence base consists of one clinical validity study and one analytical validity study, both published by the manufacturer. Additional clinical studies are needed to showcase consistency of ProMark test results to accurately predict disease severity following RP. Use of the test in clinical practice will shed light on whether test information is considered sufficient by the medical community to change treatment decision-making and if such changes result in improvement in patient-relevant outcomes, including morbidity and survival.

Criteria

- Decipher Prostate Cancer Classifier, OncotypeDX Genomic Prostate Score, Prolaris, and ProMark Proteomic Prognostic Test are considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References


5. Decipher website. Available at: http://deciphertest.com/


7. Prolaris website. Available at: https://prolaris.com/


43. ClinicalTrials.gov: Registry to Measure the Impact of Adding Genomic Testing. Available at: https://clinicaltrials.gov/ct2/show/NCT02454595?term=NCT02454595&rank=1


Genetic Testing for Epilepsy

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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<th>Procedures covered by this guideline</th>
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What is epilepsy

Definition

Epilepsy is a neurological condition that causes seizures. It affects approximately 1% of children and 1.8% of adults.¹

- Epilepsy can manifest in different ways, involving different types of seizures or multiple medical problems in addition to seizures. Seizure types include generalized seizures (absence seizures, tonic-clonic seizures) and focal seizures (simple focal seizures, complex focal seizures, secondary generalized seizures).
- An electroencephalography (EEG) can be used to help diagnose epilepsy and possibly give information as to the seizure type.
- Epilepsy has multiple causes. These include, but are not limited to, acquired causes such as stroke, brain tumor, head injury, and central nervous system infection.¹ There are also many known genetic conditions which are associated with an increased risk for epilepsy. It is estimated that approximately 40% of individuals with seizures have an underlying genetic basis for their condition (see Table 1 for a list of common genetic causes).²
• Inheritance patterns differ between the various epilepsy syndromes. Clinical heterogeneity is also seen in these conditions.

• Epileptic encephalopathy is a group of disorders in which seizures are accompanied by developmental delay or cognitive impairment. Knowledge regarding the genetic basis of these disorders has increased significantly in the last decade, resulting in wider availability of multi-gene panel testing. A recent study found that, “Targeted next-generation sequencing panels increased the genetic diagnostic yield from <10% to >25% in patients with epileptic encephalopathy.” The following are examples of epileptic encephalopathies:

  o Ohtahara syndrome (Early Infantile Epileptic Encephalopathy)
    ▪ “Characterized by early onset of intractable tonic spasms, suppression-burst pattern on interictal EEG, and poor prognosis.”
    ▪ “To date, various genes, which have essential roles in lower brain’s neuronal and interneuronal functions, have been reported to be associated with Ohtahara syndrome. For instance, syntaxin binding protein 1 (STXBP1) regulates synaptic vesicle release; aristaless-related homeobox (ARX) acts as a regulator of proliferation and differentiation of neuronal progenitors; solute carrier family 25 member 22 (SLC25A22) encodes a mitochondrial glutamate transporter; and potassium voltage-gated channel, KQT-like subfamily, member 2 (KCNQ2) plays a key role in a cell’s ability to generate and transmit electrical signals.”

  o Dravet Syndrome (Severe Myoclonic Epilepsy of Infancy)
    ▪ “Clinical cardinal features include febrile or afebrile generalized or hemiconvulsions starting in the first year of life, seizure evolution to a mixture of intractable generalized (myoclonic or atonic seizures, atypical absences) and focal seizures, normal early development, subsequent psychomotor retardation, and normal brain imaging at onset.”
    ▪ “In most of the cases with Dravet syndrome, one single gene has been involved, in contrast to other epileptic encephalopathy syndromes. SCN1A mutations have been shown in at least 80% of patients with Dravet syndrome.”

  o Infantile Spasms (West Syndrome and X-linked Infantile Spasms)
    ▪ “West syndrome is characterized by a specific seizure type, i.e., epileptic spasms, a unique interictal EEG pattern termed hypsarrhythmia, and psychomotor retardation. Spasms start within the first year of life, mainly between 4 and 6 months of age.”
“There are multiple genetic determinants of infantile spasms, which are usually explained by mutations in distinct genes. Genetic analysis of children with unexplained infantile spasms have demonstrated mutations on the X chromosome in genes such as ARX, cyclin-dependent kinase-like 5 (CDKL5), and UDP-N-acetylglucosaminytransferase subunit (ALG13) as well as de novo mutations in autosomal genes, including membrane-associated guanylate kinase, WW and PDZ domain containing protein 2 (MAGI2), STXBP1, sodium channel alpha 1 subunit (SCN1A), sodium channel protein type 2 subunit alpha (SCN2A), g-aminobutyric acid (GABA) A receptor, beta 3 (GABRB3), and dynamin 1 (DNM1).”

- Epilepsy and Intellectual Disability Limited to Females
  
  "Epilepsy and intellectual disability limited to females (EFMR) is an underrecognized disorder with X-linked inheritance but surprisingly only affecting females while sparing transmitting males. Seizure, cognitive, and psychiatric phenotypes show heterogeneity. Seizures start from the age of 6 to 36 months and may be precipitated by fever. Seizure types include GTCS, myoclonic and tonic seizures, absences, and focal seizures.”

  "Seven different mutations of PCDH19 (protocadherin 19), including missense, nonsense, and frameshift mutations, have been reported as the cause of EFMR.”

- Genetic testing for epilepsy is complicated by many factors. Epilepsy syndromes frequently have overlapping features, such as the types of seizures involved and/or additional clinical findings. Many epilepsy syndromes, including epileptic encephalopathy, are also genetically heterogeneous, and can be caused by mutations in a number of different genes. Sometimes, the inheritance pattern or the presence of pathognomonic features makes the underlying syndrome clear. However, in many cases, it can be difficult to reliably diagnose an epilepsy syndrome based on clinical and family history alone.

  - Clinical information (e.g. age of onset, seizure type, EEG results, etc) may be used in some cases to help narrow down the suspected cause. In these cases, it may be possible to identify a narrow subset of genes that may be responsible for a person’s epilepsy.
• Treatment for epilepsy ranges from antiepileptic drugs (AEDs) to ketogenic diets to vagus nerve stimulation to epilepsy surgery. Not all treatments will work for everyone and often, it takes multiple treatment trials to find a regimen that is successful. In some cases, knowing the genetic mutation that is responsible for the epilepsy can help to guide treatment.

Test information

• There are various methods used to test for mutations in genes which can cause epilepsy.
  o Single gene analysis
  o Chromosomal microarray analysis
  o Panel testing using next generation sequencing

• Chromosomal microarray (CMA) testing generally works by fluorescently tagging DNA from a patient test sample with one color and combining it with a control sample tagged in a different color. The two samples are mixed and then added to the array chip, where they compete to hybridize with the DNA fragments on the chip. By comparing the test sample versus the control, computer analysis can determine where genetic material has been deleted or duplicated in the patient.

• Until recently, most sequencing tests used the Sanger sequencing methodology that was originally developed in the 1970s. Sanger sequencing is labor intensive and did not lend itself to high-throughput applications.

• Next generation sequencing (NGS), which is also sometimes called massively parallel sequencing, has been developing since about 2005 to allow larger scale and more efficient gene sequencing. NGS relies on sequencing many copies of small pieces of DNA simultaneously and using bioinformatics to assemble the sequence.

• The efficiency of NGS has led to an increasing number of large, multi-gene testing panels. NGS panels that test several genes at once are particularly well-suited to conditions caused by more than one gene or where there is considerable clinical overlap between conditions, making it difficult to reliably narrow down likely causes. As a result, several laboratories have begun to combine genes involved in certain conditions, which often have both of those characteristics. However, NGS may not perform as well as Sanger sequencing in some applications. Results may also be obtained that cannot be adequately interpreted based on the current knowledgebase. When a sequence variation is identified that has not been previously characterized or shown to cause the disorder in question, it is called a variant of uncertain significance (VUS). VUSs are relatively common findings when sequencing large amounts of DNA with NGS.

• Epilepsy multi-gene panels include a wide variety of genes associated with epilepsy. The following are example panels (not intended to be a complete list):
o Ambry Genetics: EpiFirst-Neonate, EpiFirst-Fever, EpiFirst-IS, EpiFirst-Focal, PMEFirst/PMENext,EpilepsyNext
o GeneDx: Comprehensive Epilepsy Panel, Childhood-Onset Epilepsy Panel, STAT Epilepsy Panel, EpiXpanded Panel
o Invitae: Epilepsy Panel, Early Infantile Epileptic Encephalopathy Panel, Syndromic Neurodevelopmental Epilepsy Panel

- Multi-gene panels may also include genes believed to be associated with epilepsy, but with a lower impact on risk than recognized epilepsy syndromes. Results for such genes are of less clear value because there often are not clear management recommendation for mutation-positive individuals.
- Under certain circumstances, technologies used in multi-gene testing may fail to identify mutations that might be identifiable through single-gene testing. If high clinical suspicion remains for a particular syndrome after negative multi-gene test results, consultation with the testing lab and/or additional targeted genetic testing may be warranted.
- Multi-gene tests vary in technical specifications (e.g., depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis).
- Since genes can be easily added or removed from multi-gene tests over time by a given lab, medical records must document which genes were included in the specific multi-gene test used from each patient, and in which labs they were performed.
- Additionally, tests should be chosen that maximize the likelihood of identifying mutations in the genes of interest and that will alter patient management.

Guidelines and evidence

- No current U.S guidelines address the use of multi-gene panels in epilepsy.
- In 2016, a peer reviewed article on genetic testing for epileptic encephalopathy stated that following:
  o “Second line investigations: Targeted next generation sequencing panels of epileptic encephalopathy genes for individuals with epileptic encephalopathy.”
- In 2016, a peer reviewed article on genetic causes of early-onset epileptic encephalopathy stated the following:
  o “Molecular-based studies on early-onset epileptic encephalopathies should be performed, necessitating programmed genetical algorithms. If the phenotype could be determined with clinical findings, specific gene testing would be helpful in diagnosis. However, if the phenotype could not be determined because of overlapping phenotypes of different syndromes and the spectrum of phenotypes seen in different mutations, the use of gene panels for epilepsy would increase the probability of correct diagnosis. In a recent study, the rate of diagnosis with
targeted single gene sequencing has been reported as 15.4%, whereas the rate has increased to 46.2% with the utility of epilepsy gene panels.”

- A Task Force for the ILAE Commission of Pediatrics (2015) published recommendations for the management of infantile seizures. These recommendations included the following on treatments:

  - “for Dravet syndrome, strong evidence supports that stiripentol is effective (in combination with valproate and clobazam), whereas weak evidence supports that topiramate, zonisamide, valproate, bromide, and the ketogenic diet are possibly effective; and for Ohtahara syndrome, there is weak evidence that most antiepileptic drugs are poorly effective.”

  - “Genetic evaluation for Dravet syndrome and other infantile-onset epileptic encephalopathies should be available at tertiary and quaternary levels of care (optimal intervention would permit an extended genetic evaluation) (level of evidence—weak recommendation, level C)”

  - “Early diagnosis of some mitochondrial conditions may alter long-term outcome, but whether screening at quaternary level is beneficial is unknown (level of evidence U)”

Criteria

This policy applies to all epilepsy testing, including single genes as well as multi-gene panels, which are defined as assays that simultaneously test for more than one epilepsy gene. Medical necessity coverage generally relies on criteria established for testing individual genes.

Coverage criteria differ based on the type of testing being performed (i.e., individual epilepsy genes separately chosen versus pre-defined panels of epilepsy genes) and how that testing will be billed (one or more individual epilepsy gene procedure codes, specific panel procedure codes, or unlisted procedure codes).

Epilepsy single gene tests

Epilepsy single gene tests will be covered when the following criteria are met:

- The member has a condition that will benefit from information provided by the requested epilepsy gene testing based on at least one of the following:

  - The member displays clinical features of the condition for which testing is being requested and a particular treatment is being considered for the member that requires a genetic diagnosis, OR
A particular AED is being considered for the member and the AED is contraindicated for individuals with mutations in that gene by ONE of the following:

- A neurology therapy FDA label requires results from the genetic test to effectively or safely use or avoidance the therapy for the member’s epilepsy type and the member has not previously had a trial of the therapy, or
- An American neurological society specifically recommends the testing for the safe and effective use or avoidance of a therapy and the member has not previously had a trial of the therapy, OR

The member meets all criteria in a test-specific guideline, if available (see Table 1 for a list of genes, associated conditions, and applicable policy), AND

- The member does not have a known underlying cause for their seizures (e.g. tumor, head trauma, known genetic condition), AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Epilepsy multi-gene panels**

When separate procedure codes will be billed for individual epilepsy genes (e.g., Tier 1 MoPath codes 81200-81355 or Tier 2 MoPath codes 81400-81408), each individually billed test will be evaluated separately. The following criteria will be applied:

- The member has a condition that will benefit from information provided by the requested epilepsy gene testing based on at least one of the following:
  - The member displays clinical features of the condition for which testing is being requested and a particular treatment is being considered for the member that requires a genetic diagnosis, OR
  - A particular AED is being considered for the member and the AED is contraindicated for individuals with mutations in that gene by ONE of the following:
    - A neurology therapy FDA label requires results from the genetic test to effectively or safely use or avoidance the therapy for the member’s epilepsy type and the member has not previously had a trial of the therapy, or
    - An American neurological society specifically recommends the testing for the safe and effective use or avoidance of a therapy and the member has not previously had a trial of the therapy, OR
  - The member meets all criteria in a test-specific guideline, if available, (see Table 1 for a list of genes, associated conditions, and applicable policy), AND
  - The member does not have a known underlying cause for their seizures (e.g. tumor, head trauma, known genetic condition), AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

If the member meets the following criteria, the entire panel will be approved. However, the laboratory will be redirected to use a panel CPT code for billing purposes (e.g. 81479):

- The member has a diagnosis of early infantile epileptic encephalopathy, OR
- The member has a diagnosis of infantile spasms, OR
- The member has a diagnosis of intractable, neonatal seizures, OR
- The member has a diagnosis of febrile seizures with at least one episode of status epilepticus, OR
- The member has a progressive neurological disease defined by the following:
  - Member has epilepsy with persistent loss of developmental milestones, and
  - Member’s seizures are worsening in severity and/or frequency despite treatment, OR
- A particular AED is being considered for the member and there are 2 or more genes on the panel for which the AED is contraindicated for individuals with mutations in that gene by ONE of the following:
  - A neurology therapy FDA label requires results from the genetic test to effectively or safely use or avoidance the therapy for the member’s epilepsy type and the member has not previously had a trial of the therapy, or
  - An American neurological society specifically recommends the testing for the safe and effective use or avoidance of a therapy and the member has not previously had a trial of the therapy, AND
- The member does not display clinical features of a specific condition for which testing is available (e.g. Tuberous Sclerosis, Angelman Syndrome, Rett Syndrome, etc.), AND
- The member does not have a known underlying cause for their seizures (e.g. tumor, head trauma, known genetic condition), AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

When a multi-gene panel is being requested and will be billed with a single panel CPT code (e.g. 81479), the panel will be considered medically necessary when the following criteria are met:

- The member has a diagnosis of early infantile epileptic encephalopathy, OR
- The member has a diagnosis of infantile spasms, OR
- The member has a diagnosis of intractable, neonatal seizures, OR
- The member has a diagnosis of febrile seizures with at least one episode of status epilepticus, OR
• The member has a progressive neurological disease defined by the following:
  o Member has epilepsy with persistent loss of developmental milestones, and
  o Member’s seizures are worsening in severity and/or frequency despite treatment, OR

• A particular AED is being considered for the member and there are 2 or more genes on the panel for which the AED is contraindicated for individuals with mutations in that gene by ONE of the following:
  o A neurology therapy FDA label requires results from the genetic test to effectively or safely use or avoidance the therapy for the member’s epilepsy type and the member has not previously had a trial of the therapy, or
  o An American neurological society specifically recommends the testing for the safe and effective use or avoidance of a therapy and the member has not previously had a trial of the therapy, AND

• The member does not display clinical features of a specific condition for which testing is available (e.g. Tuberous Sclerosis, Angelman Syndrome, Rett Syndrome, etc.), AND

• The member does not have a known underlying cause for their seizures (e.g. tumor, head trauma, known genetic condition), AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Billing and reimbursement considerations

• The billed amount should not exceed the list price of the test.

• Broad epilepsy panels may not be medically necessary when a narrower panel is available and more appropriate based on the clinical findings.

• Genetic testing is only necessary once per lifetime. Therefore, a single gene included in a panel or a multi-gene panel may not be reimbursed if testing has been performed previously. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

• This guideline may not apply to genetic testing for indications that are addressed in test-specific guidelines. Please see the test-specific list of guidelines for a complete list of test-specific panel guidelines.

• If a panel was previously performed and an updated, larger panel is being requested, only testing for the medically necessary, previously untested genes will be reimbursable. Therefore, only the most appropriate procedure codes for those additional genes will be considered for reimbursement.
• If the laboratory will not accept redirection to a single code, the medical necessity of each billed component procedure will be assessed independently using the criteria above for single gene testing. Only the individual panel components that meet medical necessity criteria as a first tier of testing will be reimbursed. The remaining individual components will not be reimbursable.

Table 1: Common epilepsy genes, associate conditions and applicable guidelines

This list is not all inclusive:

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**Note** *90% of Unverricht-Lundborg syndrome is due to a repeat expansion in CSTB that may not be detected on next-generation sequencing.

ADNFLE = Autosomal Dominant Frontal Lobe Epilepsy; BFIS = Benign Familial Infantile Seizures; BFNS = Benign Familial Neonatal Seizures; EOEE = Early-Onset Epileptic Encephalopathy; GEFS+ = Generalized Epilepsy with Febrile Seizures Plus; PME = Progressive Myoclonic Epilepsy

**References**


Genetic Testing for Nonsyndromic Hearing Loss and Deafness

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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What is nonsyndromic hearing loss and deafness

Definition

Prelingual hearing loss affects about 1 out of every 500 individuals.\(^1\) Approximately 20% of cases are attributed to environmental causes, including viral (cytomegalovirus) or bacterial (meningitis) infection, trauma, prenatal exposure to certain drugs, and other environmental factors.\(^1\) The remaining 80% of cases are thought to be genetic, either as part of a recognized genetic syndrome, or as isolated, nonsyndromic hearing loss (NSHL).\(^1\)

- 95% of congenital hearing loss is detected by newborn screening. Diagnosis of hearing loss may involve physiologic testing (including auditory brainstem response or ABR/BAER) and/or audiometry.\(^1\)

- 70-80% of genetic hearing loss is nonsyndromic, with no related systemic findings.\(^1,2\) Some syndromic forms of hearing loss and deafness may masquerade as nonsyndromic in infancy and early childhood, before additional symptoms emerge. For example, goiter does not develop until puberty or adulthood in Pendred syndrome; retinitis pigmentosa emerges in adolescence in Usher syndrome; and males with Deafness-Dystonia-Optic Neuronopathy (Mohr-Tranebjaerg) Syndrome begin having progressive neurological symptoms in their teens.\(^3\)

- Many inheritance patterns are possible in NSHL; 80% is autosomal recessive, 15-19% is autosomal dominant, and ~1% is mitochondrial or X-linked.\(^1,2\)

### Procedures addressed by this guideline

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• A study of 440 individuals with genetic hearing loss found mutations in ~40% of
cases tested with a multigene panel (see Table for specific gene frequencies). The
only feature with an adverse effect on test yield was unilateral hearing loss, for
which the panel only identified mutations in 1% of cases. In another study, the
mutation detection rate was ~60% via multigene panel; multigene panel testing was
noted to be more cost-effective than single gene testing.

• While the most common cause of severe-to-profound autosomal recessive NSHL in
most populations is mutation of GJB2 (DFNB1 locus), there is ethnic variability. Approximately 1% of
DFNB1 is due to compound heterozygous mutations in GJB2 and GJB6. The most common cause of mild-to-moderate autosomal recessive
hearing loss is mutations of STRC.

• Mitochondrial NSHL is caused by mutations in MT-RNR1 (~71%), and less often,
MT-TS1 (~29%). MT-RNR1 pathogenic variants, particularly the m.1555A>G
allele, are associated with a predisposition to aminoglycoside ototoxicity, with
~100% penetrance after exposure to aminoglycosides. Without aminoglycoside
exposure, penetrance is ~80% by age 65 years.

• Management of congenital hearing loss or deafness may include hearing aids,
cochlear implants, and appropriate educational interventions. Uncovering the
genetic etiology of the hearing loss may also identify (or allay concerns about)
comorbidities that may require referral for specialty care.

Test information
• There are various methods used to test for mutations in genes which can cause
hearing loss and deafness.
  o Single gene analysis
  o Panel testing using next generation sequencing

• Until recently, most sequencing tests used the Sanger sequencing methodology
that was originally developed in the 1970s. Sanger sequencing is labor intensive
and did not lend itself to high-throughput applications.

• Next generation sequencing (NGS), which is also sometimes called massively
parallel sequencing, has been developing since about 2005 to allow larger scale
and more efficient gene sequencing. NGS relies on sequencing many copies of
small pieces of DNA simultaneously and using bioinformatics to assemble the
sequence.

• The efficiency of NGS has led to an increasing number of large, multi-gene testing
panels. NGS panels that test several genes at once are particularly well-suited to
conditions caused by more than one gene or where there is considerable clinical
overlap between conditions, making it difficult to reliably narrow down likely causes.
As a result, several laboratories have begun to combine genes involved in certain
conditions, which often have both of those characteristics. However, NGS may not
perform as well as Sanger sequencing in some applications. Results may also be
obtained that cannot be adequately interpreted based on the current knowledgebase. When a sequence variation is identified that has not been previously characterized or shown to cause the disorder in question, it is called a variant of uncertain significance (VUS). VUSs are relatively common findings when sequencing large amounts of DNA with NGS.

- Nonsyndromic hearing loss and deafness multi-gene panels include a wide variety of genes associated with nonsyndromic hearing loss and deafness. Multi-gene nonsyndromic hearing loss and deafness panels may also include genes for syndromes that mimic nonsyndromic hearing loss (e.g. Usher syndrome, Pendred syndrome, Jervell and Lange-Nielsen syndrome, etc.).

- Under certain circumstances, technologies used in multi-gene testing may fail to identify mutations that might be identifiable through single-gene testing. If high clinical suspicion remains for a particular syndrome after negative multi-gene test results, consultation with the testing lab and/or additional targeted genetic testing may be warranted.

- Multi-gene tests vary in technical specifications (e.g., depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis).

- Since genes can be easily added or removed from multi-gene tests over time by a given lab, medical records must document which genes were included in the specific multi-gene test used from each patient, and in which labs they were performed.

- Additionally, tests should be chosen that maximize the likelihood of identifying mutations in the genes of interest and that will alter patient management.

Guidelines and evidence

- In 2016, the International Pediatric Otolaryngology Group (IPOG) stated: 7
  - “In the setting of unilateral hearing loss, genetic testing has a limited role unless syndromic hearing loss is suspected.”
  - “After an audiogram and physical exam, comprehensive genetic testing (CGT) that relies on next generation sequencing (NGS) methodologies should guide subsequent workup in children with bilateral sensorineural hearing loss.”
  - “Diagnostic rates for single gene testing for GJB2/GJB6 vary significantly based on the patient’s ethnicity, and do not outperform the diagnostic rates for comprehensive genetic testing. In cases where CGT is unavailable, single gene testing can be directed by the audiometric phenotype and ethnicity.”
  - The general consensus of the authors was that temporal bone imaging “should not be a routine part of the diagnostic algorithm for bilateral symmetric sensorineural hearing loss.”

- In 2014, the American College of Medical Genetics and Genomics (ACMG) made the following recommendations: 2
A genetic evaluation is recommended for all cases of congenital deafness or hearing loss with onset in childhood or early adulthood. While the usefulness of ancillary testing (e.g. electrocardiogram, renal ultrasound, temporal bone imaging and ophthalmology examination) was mentioned, it was acknowledged that genetic testing via NGS panels would soon become more cost-effective. Cytomegalovirus (CMV) testing is important for cases of congenital hearing loss, but only accurate in the first 6 weeks of life.

Genetic testing to confirm a diagnosis of suspected syndromic hearing loss is recommended based on clinical findings. For apparently nonsyndromic hearing loss, a tiered approach was recommended: If the personal and family history is suggestive of a particular gene, single gene testing should be performed first. For simplex cases and cases with apparent autosomal recessive inheritance, the next step should be testing of GJB2 and GJB6. If single-gene testing is not diagnostic, testing via NGS panels, whole exome sequencing, or whole genome sequencing should be considered.

The statement stopped short of endorsing the use of NGS panels as a first-tier test, but noted they are “rapidly replacing” sequencing of the GJB2 and GJB6 loci and would soon be a more cost-effective alternative.

An expert-authored review of nonsyndromic hearing loss states:3

- “A comprehensive deafness-specific genetic panel that includes all genes implicated in nonsyndromic hearing loss and nonsyndromic hearing loss mimics is recommended as the initial genetic test.”
- “Performing sequence analysis of GJB2 alone is not cost-effective unless it is limited to persons with severe-to-profound congenital nonsyndromic hearing loss. Offering single-gene testing of GJB2 reflexively to everyone with congenital hearing loss without regard to the degree of hearing loss is not evidence based and not cost effective.”

An expert-authored review on hereditary hearing loss and deafness1 likewise states that a multi-gene test is recommended for apparent nonsyndromic hearing loss, while individuals with features of syndromic hearing loss should be diagnosed with targeted genetic testing. Ancillary cardiac, ophthalmologic and renal evaluations are only recommended on the basis of genetic test results or clinical findings.

An expert-authored review on mitochondrial NSHL6 states that the diagnosis should be suspected in individuals with moderate-to-profound hearing loss and a family history suggestive of maternal inheritance (e.g. no transmission through a male), or onset of hearing loss after exposure to an aminoglycoside antibiotic.

To confirm the diagnosis: MT-RNR1 testing, beginning with targeted analysis for the m.1555A>G mutation, is recommended following aminoglycoside exposure. In the absence of aminoglycoside exposure, testing of both MT-RNR1 and MT-TS1 is recommended. If these tests fail to confirm a diagnosis, mitochondrial genome sequencing can be considered.
An alternative strategy is to perform a multi-gene panel that includes both MT-RNR1 and MT-TS1, plus other genes of interest.

Criteria

Known Familial Mutation Analysis

- Previous testing:
  - Member has not previously had testing for the requested mutation(s), AND
  - Member has a 1st, 2nd, or 3rd degree biologic relative with a pathogenic mutation(s) in a gene associated with nonsyndromic hereditary hearing loss or deafness, AND
  - Member is at risk of inheriting the pathogenic mutation based on the family history and the inheritance pattern associated with the mutation, AND

- Diagnostic testing:
  - Member has nonsyndromic hearing loss or deafness that is consistent with the mutation in the family, OR

- Carrier screening:
  - Member is of reproductive age, and
  - Member has ability and intention to reproduce, or
  - Member is currently pregnant.

GJB2 Sequencing

- Previous testing:
  - Member has not previously had GJB2 sequencing, and
  - No known pathogenic hearing loss/deafness gene variants in a biologic relative, AND

- Diagnostic Testing:
  - Member has a diagnosis of bilateral sensorineural hearing loss, and
  - Prelingual onset of hearing loss (prior to speech development), and
  - No known cause for the member’s hearing loss (e.g., prenatal exposure to ototoxic medication or TORCH infection, known genetic disorder), and
  - Absence of significant dysmorphism, congenital anomalies or other signs of syndromic hearing loss, and
o Member’s family history is consistent with autosomal recessive inheritance (including simplex cases), OR

• Carrier screening
  o Member is of reproductive age, and
  o Has potential and intention to reproduce, and
  o Has a reproductive partner who is a carrier of a GJB2/GJB6 mutation, or
  o Has a reproductive partner with GJB2/GJB6-related deafness.

GJB6 Common Variant Analysis for 309kb and 232kb Deletions

• Previous testing:
  o Member has not previously had GJB6 common variant analysis or deletion/duplication analysis, AND

• Diagnostic Testing:
  o Member meets criteria for GJB2 sequencing, and
  o No mutation or only one mutation identified on GJB2 sequencing, OR

• Carrier screening
  o Member is of reproductive age, and
  o Has potential and intention to reproduce, and
  o Has a 1st, 2nd, or 3rd-degree biologic relative with a GJB6 variant, or
  o Member meets criteria for GJB2 sequencing, and
  o No mutation identified on GJB2 sequencing.

MT-RNR1 Targeted Mutation Analysis for m.1555A>G Mutation

• Previous testing:
  o Member has not previously had MT-RNR1 targeted mutation analysis, and
  o No known pathogenic hearing loss/deafness gene variants in a biologic relative, AND

• Diagnostic Testing:
  o Member has a diagnosis of bilateral sensorineural hearing loss, and
  o No known cause for the member’s hearing loss (e.g., prenatal exposure to ototoxic medication or TORCH infection, known genetic disorder), and
Absence of significant dysmorphism, congenital anomalies or other signs of syndromic hearing loss, and

Member has one of the following risk factors for MT-RNR1 related deafness:

- History of aminoglycoside antibiotic exposure (gentamycin, tobramycin, amikacin, kanamycin, or streptomycin), or
- Member’s family history is strongly suggestive of mitochondrial inheritance (no transmission through a male).

**MT-RNR1 Sequencing**

- **Previous testing:**
  - Member has not previously had MT-RNR1 sequencing, and
  - No mutations detected in any previous MT-RNR1 testing (targeted m.1555A>G mutation analysis), and
  - No known pathogenic hearing loss/deafness gene variants in a biologic relative, **AND**

- **Diagnostic Testing:**
  - Member has a diagnosis of bilateral sensorineural hearing loss, and
  - No known cause for the member’s hearing loss (e.g., prenatal exposure to ototoxic medication or TORCH infection, known genetic disorder), and
  - Absence of significant dysmorphism, congenital anomalies or other signs of syndromic hearing loss, and
  - Member has one of the following risk factors for MT-RNR1 related deafness:
    - Aminoglycoside antibiotic exposure (gentamycin, tobramycin, amikacin, kanamycin, or streptomycin) prior to hearing loss onset, or
    - Member’s family history is strongly suggestive of mitochondrial inheritance (no transmission through a male).

**MT-TS1 Sequencing**

- **Previous testing:**
  - Member has not previously had MT-TS1 analysis, and
  - No mutations detected in any previous MT-TS1 testing (targeted variant analysis), and
• Diagnostic Testing:
  o Member has a formal diagnosis of bilateral sensorineural hearing loss, and
  o No known cause for the member’s hearing loss (e.g., prenatal exposure to ototoxic medication or TORCH infection, known genetic disorder), and
  o Absence of significant dysmorphism, congenital anomalies, or other signs of syndromic hearing loss, and
  o Member’s family history is strongly suggestive of mitochondrial inheritance (no transmission through a male).

Hearing Loss and Deafness Multigene Panel Testing

When a multi-gene panel is being requested and will be billed with a panel CPT code (e.g. 81430, 81431, 81479), the panel will be considered medically necessary when the following criteria are met:

• Previous testing:
  o Member has not previously had a hearing loss panel, and
  o No known pathogenic hearing loss/deafness gene variants in a biologic relative, AND

• Diagnostic Testing:
  o Member has a diagnosis of bilateral sensorineural hearing loss, and
  o No known cause for the member’s hearing loss (e.g., prenatal exposure to ototoxic medication or TORCH infection, known genetic disorder), and
  o Absence of significant dysmorphism, congenital anomalies or other signs of syndromic hearing loss.

When separate procedure codes will be billed for individual hearing loss genes (e.g., Tier 1 MoPath codes 81200-81355 or Tier 2 MoPath codes 81400-81408), the entire panel will be approved if the above criteria are met. However, the laboratory will be redirected to use an appropriate panel CPT code for billing purposes (e.g. 81430, 81431, 81479).

Billing and reimbursement considerations

• The billed amount should not exceed the list price of the test.
• Broad hearing loss and deafness panels may not be medically necessary when a narrower panel is available and more appropriate based on the clinical findings.
• Genetic testing is only necessary once per lifetime. Therefore, a single gene included in a panel or a multi-gene panel may not be reimbursed if testing has been performed previously. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

• If a panel was previously performed and an updated, larger panel is being requested, only testing for the medically necessary, previously untested genes will be reimbursable. Therefore, only the most appropriate procedure codes for those additional genes will be considered for reimbursement.

• If the laboratory will not accept redirection to a single code, the medical necessity of each billed component procedure will be assessed independently, and only the individual panel components that meet medical necessity criteria as a first tier of testing will be reimbursed. The remaining individual components will not be reimbursable.
  
  o If appropriate first-tier tests cannot be determined on the basis of clinical and family histories, only the following genes may be considered for reimbursement: GJB2, STRC, SLC26A4, TECTA, MYO15A, MYO7A.

• If a single hearing loss/deafness gene test is billed simultaneously with a panel code (e.g. 81430), only the billed procedure that meets medical necessity criteria as a first tier of testing will be reimbursed.
  
  o Panel testing will generally be the most appropriate first-tier test, except when the history is strongly suggestive of the individual genetic disorder requested (e.g. congenital, severe-to-profound deafness for GJB2 analysis or history of aminoglycoside exposure for MT-RNR1 analysis).

References


Genitourinary Conditions Molecular Testing

Introduction

Molecular testing for genitourinary conditions is addressed by this guideline.

Procedures addressed

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<td>Human Papillomavirus (HPV), low-risk types (for example, 6, 11, 42, 43, 44)</td>
<td>87623</td>
</tr>
<tr>
<td>Human Papillomavirus (HPV), high-risk types (for example, 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 68)</td>
<td>87624, G0476</td>
</tr>
<tr>
<td>Human Papillomavirus (HPV), types 16 and 18 only, includes type 45, if performed</td>
<td>87625</td>
</tr>
</tbody>
</table>
Procedures addressed by this guideline | Procedure codes
---|---
Herpes simplex virus | 87528
| 87529
| 87530
Infectious agent detection by nucleic acid (DNA or RNA), not otherwise specified; direct probe technique, each organism | 87797
Infectious agent detection by nucleic acid (DNA or RNA), not otherwise specified; amplified probe technique, each organism | 87798
Infectious agent detection by nucleic acid (DNA or RNA), not otherwise specified; quantification, each organism | 87799
Infectious agent detection by nucleic acid (DNA or RNA), human papillomavirus (HPV) for five or more separately reported highrisk HPV types (eg, 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 59, 68) (ie, genotyping) | 0500T

**Test information**

**Introduction**

Molecular testing for genitourinary conditions may include nucleic acid testing, flow cytometry, immunohistochemistry, or other specialized molecular studies.

**Criteria**

**Introduction**

Requests for molecular testing for genitourinary conditions are reviewed using these criteria.

**Chlamydia trachomatis**

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chlamydia Trachomatis Detection, Direct Method</td>
<td>87490</td>
<td>Procedure code is eligible for reimbursement when criteria are met</td>
</tr>
<tr>
<td>Chlamydia Trachomatis</td>
<td>87491</td>
<td>Procedure code is eligible</td>
</tr>
<tr>
<td>Test name</td>
<td>Procedure code</td>
<td>Reimbursement</td>
</tr>
<tr>
<td>-----------------------------------------------</td>
<td>----------------</td>
<td>---------------------------------------------------------</td>
</tr>
<tr>
<td>Detection, Amplified Method</td>
<td></td>
<td>for reimbursement when criteria are met</td>
</tr>
<tr>
<td>Chlamydia Trachomatis Detection, Quantification Method</td>
<td>87492</td>
<td>Procedure code is not eligible for reimbursement under any circumstances</td>
</tr>
</tbody>
</table>

**Direct or amplified Chlamydia trachomatis detection**

**Medical necessity requirements:**

Nucleic acid amplification testing (NAAT) for Chlamydia trachomatis through either direct (CPT 87490) or amplified (CPT 87491) probe studies is considered medically necessary for individuals with clinical indications as outlined here.

Indications for testing in asymptomatic individuals:

- Routine annual screening of all sexually active women aged less than or equal to 25 years
- Screening of sexually active women greater than 25 years of age with risk factors (e.g., those who have a new sex partner or multiple sex partners)
- Routine screening for all pregnant women during one of the first prenatal visits
- Retesting of all pregnant women aged less than or equal to 25 years performed during the third trimester
- Retesting of all pregnant women over age 25 during the third trimester when at increased risk for Chlamydia (e.g., women who have a new or multiple sex partners, women with a history of a previous STI, high risk behavior such as inconsistent condom use, sex work)
- Screening of sexually active men with risk factors (e.g., men in correctional facilities, presenting to STI clinics, or who have infected partner)

Indications for testing in symptomatic individuals:

- Cervicitis
- Urethritis

**Test frequency:**

Repeat testing to document eradication of infection after completing an appropriate treatment regimen is recommended only in the following settings: patient is pregnant, symptoms persist, re-infection is suspected, or compliance with therapy is in question. Routine test of cure is not recommended.

- Non-pregnant recently infected women should be retested 3 to 12 months after treatment.
Based on guidelines for initial and repeat testing, no more than five screenings in a year should be necessary regardless of pregnancy or other risk factors.

Billing and reimbursement:
When testing is medically necessary, the following limitations apply:

- NAAT may be performed on urine, vaginal, or cervical samples. It is usually sufficient to test one site. When necessary to test more than one site:
  - Additional units must be billed with modifier 59.
  - No more than 3 units of 87490 or 87491 for chlamydia trachomatis molecular testing may be billed for the same date of service.

- More than one type of molecular test for the same organism will not be reimbursed for the same date of service (e.g., 87490 and 87491 may not be billed together).

- Based on guidelines for initial and repeat testing, no more than five screenings in a year should be necessary regardless of pregnancy or other risk factors.

Quantitative Chlamydia trachomatis testing

Medical necessity requirements:
Medical necessity of quantitative testing for chlamydia trachomatis (CPT 87492) has not been demonstrated, and is therefore determined to be investigational and experimental. This procedure code is not eligible for reimbursement for any clinical indications.

Billing and reimbursement:
If the laboratory’s testing platform consists of direct or amplified and quantitative testing methodologies, yet only direct or amplified testing is considered medically necessary based on these criteria, the lab may request reimbursement for only the medically necessary components of the test by using a procedure code that does not represent all testing methodologies performed.

Neisseria gonorrhoeae

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Neisseria Gonorrhoeae, Direct Method</td>
<td>87590</td>
<td>Procedure code is eligible for reimbursement when criteria are met</td>
</tr>
<tr>
<td>Neisseria Gonorrhoeae, Amplified Method</td>
<td>87591</td>
<td>Procedure code is eligible for reimbursement when criteria are met</td>
</tr>
</tbody>
</table>
**Direct or amplified Neisseria gonorrhoeae detection**

**Medical necessity requirements:**

Nucleic acid amplification testing (NAAT) for Neisseria gonorrhoeae through either direct (CPT 87590) or amplified (CPT 87591) probe studies is considered medically necessary for individuals with clinical indications as outlined here.

Indications for testing in asymptomatic individuals:

- Annual screening of all sexually active women aged less than or equal to 25 years.
- Screening of women greater than 25 years who are at increased risk for infection (e.g., women with previous gonorrhea infection, other STIs, new or multiple sex partners, and inconsistent condom use, sex workers, or women living in communities with a high prevalence of disease).
- All pregnant women at increased risk for gonorrhea (as defined in the above criteria) should be screened at the first prenatal visit for *N. gonorrhoeae*.
- Uninfected pregnant women who remain at high risk for gonococcal infection also should be retested during the third trimester.
- Screening of sexually active individuals who have an infected partner.

Indications for testing in symptomatic individuals:

- Cervicitis
- Urethritis

**Test frequency:**

When indicated, repeat testing to document eradication should not be performed until 3–4 weeks after the positive result. Pregnant women diagnosed with gonococcal infection during the first trimester should be retested within approximately 3–6 months, preferably in the third trimester. Recently infected women should be retested 3 to 12 months after treatment. When repeat testing is indicated, the following limitations apply:

- Repeat testing will not be reimbursed if performed within three weeks (less than 21 days) from a previous test.
Based on guidelines for initial and repeat testing, no more than five screenings in a year should be necessary regardless of pregnancy or other risk factors.

**Billing and reimbursement**

When testing is medically necessary, the following limitations apply:

- Nucleic acid amplification test (NAAT) may be performed on urine, vaginal, or cervical samples. It is usually sufficient to test one site. When necessary to test more than one site:
  - Additional units must be billed with modifier 59.
  - No more than 3 units of 87590 or 87591 for Neisseria gonorrhoeae molecular testing may be billed for the same date of service.

- More than one type of molecular test for the same organism will not be reimbursed for the same date of service (e.g., 87590 and 87591 may not be billed together).

**Quantitative Neisseria gonorrhoeae testing**

**Medical necessity requirements:**

Medical necessity of quantitative testing for Neisseria gonorrhoeae (CPT 87592) has not been demonstrated, and is therefore determined to be investigational and experimental. This procedure code is not eligible for reimbursement for any clinical indications.

**Billing and reimbursement:**

If the laboratory’s testing platform consists of direct or amplified and quantitative testing methodologies, yet only direct or amplified testing is considered medically necessary based on these criteria, the lab may request reimbursement for only the medically necessary components of the test by using a procedure code that does not represent all testing methodologies performed.

**Trichomonas vaginalis**

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trichomonas Vaginalis Detection, Direct Method</td>
<td>87660</td>
<td>Procedure code is eligible for reimbursement when criteria are met</td>
</tr>
<tr>
<td>Trichomonas Vaginalis Detection, Amplified Method</td>
<td>87661</td>
<td>Procedure code is eligible for reimbursement when criteria are met</td>
</tr>
</tbody>
</table>
Direct or amplified Trichomonas vaginalis detection

Medical necessity requirements:

Nucleic acid amplification testing (NAAT) for Trichomonas vaginalis through either direct (CPT 87660) or amplified (CPT 87661) probe studies is considered medically necessary for individuals with clinical indications as outlined here.

Indications for testing in asymptomatic individuals:¹

- Evidence does not support routine screening for Trichomonas vaginalis in asymptomatic women (pregnant or non-pregnant) or men who are not at high risk for infection.
- Screening can be considered in those at increased risk for Trichomonas vaginalis infection for reasons such as new or multiple sex partners, history of STIs, sex work, or drug use.
- Screening should also be performed in sexually active women who are HIV-positive at entry into care and then at least annually.

Indications for testing in symptomatic individuals:¹

- Vaginitis, abnormal vaginal discharge, cervicitis, nongonococcal urethritis, vulvar pruritis, or pelvic inflammatory disease.
- Sexually active women with trichomoniasis may be rescreened for Trichomonas vaginalis at 3 months following initial infection.
- Screening of sexually active individuals who have an infected partner.

Test frequency:

- Repeat testing should not be necessary more frequently than every three months.
- Based on guidelines for initial and repeat testing, no more than five screenings in a year should be necessary regardless of pregnancy or other risk factors.

Billing and reimbursement:

The medical necessity of testing will be determined based on the following claims data:

- When testing asymptomatic individuals, an ICD code that supports increased risk, infected partner, or positive HIV status must be submitted on the claim. For guidance, see Tables: ICD Codes Indicating High Risk Indications, ICD Codes Indicating Infected Partner, and ICD Codes Indicating HIV Positive Status.
- When testing symptomatic individuals, an ICD code that describes the common symptoms, as defined in Table: *ICD Codes Indicating Symptoms of Genitourinary Conditions*, must be submitted on the claim.

When testing is medically necessary, the following limitations apply:

- Nucleic acid amplification test (NAAT) may be performed on urine, vaginal, or cervical samples.\(^1\) It is usually sufficient to test one site. When necessary to test more than one site:
  - Additional units must be billed with modifier 59.
  - No more than 3 units of 87660 or 87661 for Trichomonas vaginalis molecular testing may be billed for the same date of service.

- More than one type of molecular test for the same organism will not be reimbursed for the same date of service (e.g., 87660 and 87661 may not be billed together).

### Candida species

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Candida Detection, Direct Method</td>
<td>87480</td>
<td>Procedure code is eligible for reimbursement when criteria are met.</td>
</tr>
<tr>
<td>Candida Detection, Amplified Method</td>
<td>87481</td>
<td>Procedure code is eligible for reimbursement when criteria are met.</td>
</tr>
<tr>
<td>Candida Detection, Quantification Method</td>
<td>87482</td>
<td>Procedure code is not eligible for reimbursement under any circumstances.</td>
</tr>
</tbody>
</table>

**Direct or amplified Candida detection**

**Medical necessity requirements:**

Nucleic acid amplification testing (NAAT) for Candida albicans through either direct (CPT 87480) or amplified (CPT 87481) probe studies is considered medically necessary for individuals with clinical indications as outlined here.

Indications for asymptomatic individuals:

- Evidence does not support routine screening for Candida species in asymptomatic pregnant women, non-pregnant women, or men unless HIV status is positive.\(^1\)

Indications for symptomatic individuals:\(^1\)
• Candida testing is generally diagnosed by non-molecular methods (clinical criteria, microscopy, culture, etc.). Molecular testing for Candida should rarely be necessary.

• However, guidelines do support molecular testing for Candida in symptomatic females when microscopy is not available.
  • Post-service medical necessity review may be employed to ensure appropriate non-molecular methods have been utilized or were unavailable.

Billing and reimbursement:

The medical necessity of testing will be determined based on the following claims data:

• When testing asymptomatic males or females, an ICD code that supports positive HIV status must be submitted on the claim (see Table: ICD Codes Indicating HIV Positive Status). Note that testing for males is only indicated when HIV positive (i.e., no symptomatic or other testing indications).

• When testing symptomatic females, an ICD code that describes the common symptoms must be submitted on the claim (see Table: ICD Codes Indicating Symptoms of Genitourinary Conditions).

• When testing is medically necessary, the following limitations apply:
  • It should only be necessary to test one site. Therefore, only one unit per date of service is reimbursable.
  • Subtyping for Candida glabrata and other nonalbicans Candida species is not routinely medically necessary, so only one unit will be routinely reimbursed. Exceptions may be considered if complicated vulvovaginal candidiasis (VVC) is diagnosed. Complicated VVC may include:
    • Recurrent VVC (defined as 4 or more episodes of symptomatic VVC within 1 year), or
    • Severe VVC (i.e. extensive vulvar erythema, edema, excoriation, and fissure formation)
  • More than one type of molecular test for the same organism will not be reimbursed for the same date of service (e.g., 87480 and 87481 may not be billed together).

Quantitative Candida testing

Medical necessity requirements:

Medical necessity of quantitative testing for Candida albicans (CPT 87482) has not been demonstrated, and is therefore determined to be investigational and
This procedure code is not eligible for reimbursement for any clinical indications.

Billing and reimbursement:

If the laboratory’s testing platform consists of direct or amplified and quantitative testing methodologies, yet only direct or amplified testing is considered medically necessary based on these criteria, the lab may request reimbursement for only the medically necessary components of the test by using a procedure code that does not represent all testing methodologies performed.

**Gardnerella vaginalis**

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gardnerella Vaginalis Detection, Direct Method</td>
<td>87510</td>
<td>Procedure code is eligible for reimbursement when criteria are met.</td>
</tr>
<tr>
<td>Gardnerella Vaginalis Detection, Amplified Method</td>
<td>87511</td>
<td>Procedure code is eligible for reimbursement when criteria are met.</td>
</tr>
<tr>
<td>Gardnerella Vaginalis Detection, Quantification</td>
<td>87512</td>
<td>Procedure code is not eligible for reimbursement under any circumstances</td>
</tr>
</tbody>
</table>

**Direct or amplified Gardnerella vaginalis detection**

**Medical necessity requirements:**

Nucleic acid amplification testing (NAAT) for Gardnerella vaginalis through either direct (CPT 87510) or amplified (CPT 87511) probe studies is considered medically necessary for individuals with clinical indications as outlined here.

**Indications for asymptomatic individuals:**

- Evidence does not support routine screening for Gardnerella vaginalis in asymptomatic pregnant women, non-pregnant women, or men for any indications.

**Indications for symptomatic individuals:**

- Gardnerella vaginalis testing is generally diagnosed by non-molecular methods (clinical criteria and microscopy.). Molecular testing for Gardnerella vaginalis should rarely be necessary.
However, guidelines do support molecular testing for Gardnerella vaginalis in symptomatic females only when microscopy is not available.

- Post-service medical necessity review may be employed to ensure appropriate non-molecular methods have been utilized or were unavailable.

**Billing and reimbursement:**

The medical necessity of testing will be determined based on the following claims data:

- When testing symptomatic females, an ICD code that describes the common symptoms must be submitted on the claim (see Table Symptoms of STIs).
- Note that there are no medically necessary indications for testing in males.
- When testing is medically necessary, the following limitations apply:
  - Medical necessity of quantitative testing for Gardnerella vaginalis (CPT 87512) has not been demonstrated for any indication, and is therefore determined to be investigational and experimental.
  - It should only be necessary to test one site. Therefore, only one unit per date of service is reimbursable.
  - More than one type of molecular test for the same organism will not be reimbursed for the same date of service (e.g., 87510 and 87511 may not be billed together).

**Quantitative Gardnerella vaginalis testing**

**Medical necessity requirements:**

Medical necessity of quantitative testing for Gardnerella vaginalis (CPT 87512) has not been demonstrated, and is therefore determined to be investigational and experimental. This procedure code is not eligible for reimbursement for any clinical indications.

**Billing and reimbursement:**

If the laboratory’s testing platform consists of direct or amplified and quantitative testing methodologies, yet only direct or amplified testing is considered medically necessary based on these criteria, the lab may request reimbursement for only the medically necessary components of the test by using a procedure code that does not represent all testing methodologies performed.

Herpes simplex virus (HSV)

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Herpes Simplex Virus</td>
<td>87528</td>
<td>Procedure code is eligible</td>
</tr>
<tr>
<td>Test name</td>
<td>Procedure code</td>
<td>Reimbursement</td>
</tr>
<tr>
<td>-----------------------------------------------</td>
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<td>----------------------------------------------------</td>
</tr>
<tr>
<td>Detection, Direct Method</td>
<td></td>
<td>for reimbursement when criteria are met.</td>
</tr>
<tr>
<td>Herpes Simplex Virus Detection, Amplified</td>
<td>87529</td>
<td>Procedure code is eligible for reimbursement when</td>
</tr>
<tr>
<td>Method</td>
<td></td>
<td>criteria are met.</td>
</tr>
<tr>
<td>Herpes Simplex Virus Detection, Quantification</td>
<td>87530</td>
<td>Procedure code is eligible for reimbursement when</td>
</tr>
<tr>
<td>Method</td>
<td></td>
<td>criteria are met.</td>
</tr>
</tbody>
</table>

Direct or amplified Herpes simplex virus (HSV) detection

Medical necessity requirements:

Nucleic acid amplification testing (NAAT) for Herpes simplex virus (HSV) through either direct (CPT 87528) or amplified (CPT 87529) probe studies is considered medically necessary for individuals with clinical indications as outlined here.

Indications for testing in asymptomatic Individuals: 1,5,6,7

- Current guidelines explicitly recommend against testing asymptomatic individuals for HSV.

Indications for testing in symptomatic Individuals: 1,5,6

- New or recurrent vesicular and/or ulcerative lesions, vesicles or ulcers on or around the genitals, rectum, buttocks, thighs
- Recurrent genital symptoms or atypical symptoms and negative HSV cultures

Billing and reimbursement:

When testing is medically necessary, the following limitations apply:

- It should only be necessary to test one site. Therefore, only one unit per date of service is reimbursable.
- More than one type of molecular test for the same organism will not be reimbursed for the same date of service (e.g., 87528 and 87529 may not be billed together).

Quantitative Herpes simplex virus testing

Medical necessity requirements:

Quantitative testing for Herpes simplex virus (HSV) (CPT 87530) may be reasonable for monitoring disease in some circumstances.
Therefore, quantitative HSV testing will be reimbursable when a diagnosis has been established and the need for monitoring is documented in the medical record. Quantitative HSV testing should not be used for the primary diagnosis of HSV.

Billing and reimbursement:

If the laboratory’s testing platform consists of direct or amplified and quantitative testing methodologies, yet only direct or amplified testing is considered medically necessary based on these criteria, the lab may request reimbursement for only the medically necessary components of the test by using a procedure code that does not represent all testing methodologies performed.

### Human Papillomavirus (HPV)

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Human Papillomavirus (HPV) Detection, Low-Risk Types</td>
<td>87623</td>
<td>Procedure code is not eligible for reimbursement under any circumstances.</td>
</tr>
<tr>
<td>Human Papillomavirus (HPV) Detection, High-Risk Types</td>
<td>87624</td>
<td>Procedure code is eligible for reimbursement when criteria are met.</td>
</tr>
<tr>
<td>Human Papillomavirus (HPV) Detection, Types 16 and 18</td>
<td>87625</td>
<td>Procedure code is eligible for reimbursement when criteria are met.</td>
</tr>
<tr>
<td>Human Papillomavirus (HPV) Detection, High-Risk Types</td>
<td>G0476</td>
<td>Procedure code is eligible for reimbursement when criteria are met.</td>
</tr>
</tbody>
</table>

**Low-risk genotyping for human papillomavirus (HPV)**

**Medical necessity requirements:**

Medical necessity of testing for low-risk (non-oncogenic) types of HPV (CPT 87623) has not been demonstrated, and is therefore determined to be investigational and experimental. This procedure code is not eligible for reimbursement for any clinical indications.

- Per American College of Obstetrics and Gynecology cervical cancer clinical management guidelines, HPV testing should be performed only to detect high-risk (oncogenic) types of HPV.\(^{10}\)
- As stated in American Society for Colposcopy and Cervical Pathology (ASCCP) guidelines, “testing for low-risk (nononcogenic) HPV types has no role in the evaluation of women with abnormal cervical cytologic results.”\(^{11}\)
- Per Centers for Disease Control and Prevention (CDC) Sexually Transmitted Diseases guidelines, “HPV testing is not recommended for anogenital wart diagnosis, because test results are not confirmatory and do not guide genital wart management.”

**High-risk genotyping for human papillomavirus (HPV)**

**Medical necessity requirements:**

Nucleic acid amplification testing (NAAT) for human papillomavirus high-risk genotypes (87624, 87625, G0476) is considered medically necessary for individuals with clinical indications as outlined here.

**Indications for testing in asymptomatic individuals:**

- Among women age 30-65, HPV testing may be performed every 5 years in combination with pap smear (co-testing) for routine screening.
- Women aged 30 years and older who are HPV positive but cytology negative may:
  - Test again by co-testing in one year, or
  - Be tested by HPV high risk oncogenic subtype genotyping
- Women aged 30 years and older with cytology reported as negative and with absent or insufficient endocervical/transformation zone (EC/TZ) component and no or unknown HPV test result

**Indications for testing in symptomatic individuals:**

- Reflex to HPV testing for management of women with atypical squamous cells of undetermined significance (ASC-US) cervical cytology results starting at age 21.
- Co-testing at 1 year post cervical intraepithelial neoplasia grade 1 (CIN1) or no lesion preceded by HPV-16 or HPV-18 positivity, persistent untyped oncogenic HPV, ASC-US, and low grade squamous intraepithelial lesion (LSIL) starting at age 25.
- For women treated for cervical intraepithelial neoplasia (CIN 2, CIN 3, or CIN 2, 3), co-testing at 12 months and 24 months is recommended.
- Post-menopausal women with LSIL.

**Exclusions:**

Medical necessity of the following methods for HPV detection has not been demonstrated and is therefore determined to be investigational and experimental.

- Flow cytometry (e.g., HPV OncoTect) (CPT 88184, 88185, and/or 88187).
Billing and reimbursement:
Based on medical necessity criteria, HPV testing will not be reimbursed when billed for individuals with the following age restrictions:

- Testing in asymptomatic individuals is not medically necessary before age 30.
- Testing is not medically necessary before age 21 for any indication (asymptomatic or symptomatic).

When testing is medically necessary, the following limitations apply:

- Nucleic acid amplification test (NAAT) may be performed on endocervical samples.\(^9,11\) It is usually sufficient to test one specimen.
- Therefore, no more than 1 unit of CPT 87624 or 87625 for human papillomavirus molecular testing may be billed for the same date of service.
- More than one type of molecular test for the same organism will not be reimbursed for the same date of service. For example, nucleic acid detection of high risk subtypes HPV-16 and HPV-18 by two methodologies (CPT 87624 and 87625) cannot be billed together, and nucleic acid detection by either of these methodologies cannot be billed with a test using another molecular methodology (e.g., in situ hybridization, CPT 88365).

Infectious agent, not otherwise specified

<table>
<thead>
<tr>
<th>Test name</th>
<th>Procedure code</th>
<th>Reimbursement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Miscellaneous Infectious Agent Detection, Direct Molecular Method</td>
<td>87797</td>
<td>Procedure code is not eligible for reimbursement for indications addressed by this policy.</td>
</tr>
<tr>
<td>Miscellaneous Infectious Agent Detection, Amplified Molecular Method</td>
<td>87798</td>
<td>Procedure code is not eligible for reimbursement for indications addressed by this policy.</td>
</tr>
<tr>
<td>Miscellaneous Infectious Agent Detection, Quantitative Molecular Method</td>
<td>87799</td>
<td>Procedure code is not eligible for reimbursement for indications addressed by this policy.</td>
</tr>
</tbody>
</table>

Miscellaneous infectious agent detection

Medical necessity requirements:

- Molecular testing to detect a variety of organisms that do not have organism-specific procedure codes may be billed under the infectious agents not otherwise specified (NOS) codes (CPT 87797, 87798, 87799). This guideline
only addresses some organisms and clinical settings. It does not apply to all testing performed under these codes.

- The genitourinary organisms for which molecular testing is supported by guidelines are represented by organism-specific CPT codes. There are no clinical indications for any infectious agents billed under not otherwise (NOS) specified procedure codes that are supported by current evidence for the evaluation or management of genitourinary conditions, including bacterial vaginosis. Therefore, testing for organisms NOS is considered investigational and experimental in the setting of screening for genitourinary conditions, including but not limited to the following tests:
  - NuSwab®, Vaginitis Plus (VG+) (A vaginae, BVAB-2, Megasphaera Type 1)\(^{12}\)
  - OneSwab® (A vaginae, Megasphaera Type 1 and 2, BVAB-2)\(^{13}\)
  - SureSwab® Vaginosis, Vaginitis Plus (G vaginalis, A vaginae, Megasphaera species)\(^{14}\)

**Billing and reimbursement:**

The following criteria are used to determine if testing for infectious agents NOS is being performed in the setting of genitourinary condition detection or management, including bacterial vaginosis:

- When billed with any ICD code included in Table: *ICD Codes Indicating Testing Done for Genitourinary Conditions* below.
- When billed on the same date of service with any other organism-specific CPT code referenced in this policy.

**ICD10 Codes**

- ICD10 codes in this section may be used to support medical necessity as described in the above guidelines.

**ICD Codes Indicating High Risk Indications**

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>F10.X</td>
<td>Alcohol related disorders</td>
</tr>
<tr>
<td>F11.X</td>
<td>Opioid related disorders</td>
</tr>
<tr>
<td>F12.X</td>
<td>Cannabis related disorders</td>
</tr>
<tr>
<td>F13.X</td>
<td>Sedative, hypnotic, or anxiolytic related disorders</td>
</tr>
<tr>
<td>F14.X</td>
<td>Cocaine related disorders</td>
</tr>
<tr>
<td>F15.X</td>
<td>Other stimulant related disorders</td>
</tr>
</tbody>
</table>
### ICD10 Code or Range

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>F16.X</td>
<td>Hallucinogen related disorders</td>
</tr>
<tr>
<td>F18.X</td>
<td>Inhalant related disorders</td>
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<tr>
<td>F19.X</td>
<td>Other psychoactive substance related disorders</td>
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<tr>
<td>O99.32X</td>
<td>Drug use complicating pregnancy, childbirth, and the puerperium</td>
</tr>
<tr>
<td>Z72.5X</td>
<td>High risk sexual behavior</td>
</tr>
<tr>
<td>Z77.9</td>
<td>Other contact with and (suspected) exposures hazardous to health</td>
</tr>
</tbody>
</table>

### ICD Codes Indicating Infected Partner

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
</tr>
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<tbody>
<tr>
<td>Z20.2</td>
<td>Contact with and (suspected) exposure to infections with a predominantly sexual mode of transmission</td>
</tr>
<tr>
<td>Z20.6</td>
<td>Contact with and (suspected) exposure to human immunodeficiency virus [HIV]</td>
</tr>
<tr>
<td>Z20.8X</td>
<td>Contact with and (suspected) exposure to other communicable diseases</td>
</tr>
<tr>
<td>Z20.9</td>
<td>Contact with and (suspected) exposure to unspecified communicable disease</td>
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### ICD Codes Indicating HIV Positive Status

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
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<tbody>
<tr>
<td>B20</td>
<td>Human immunodeficiency virus [HIV] disease</td>
</tr>
<tr>
<td>B97.35</td>
<td>Human immunodeficiency virus, type 2 [HIV-2]</td>
</tr>
<tr>
<td>O98.7X</td>
<td>Human immunodeficiency virus [HIV] disease complicating pregnancy, childbirth and the puerperium</td>
</tr>
<tr>
<td>R75</td>
<td>Inconclusive laboratory evidence of human immunodeficiency virus [HIV]</td>
</tr>
<tr>
<td>Z21</td>
<td>Asymptomatic human immunodeficiency virus [HIV] infection status</td>
</tr>
</tbody>
</table>
### ICD Codes Indicating Symptoms of Genitourinary Conditions

<table>
<thead>
<tr>
<th>ICD10 Code or Range</th>
<th>Description</th>
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<tbody>
<tr>
<td>A56.X</td>
<td>Other sexually transmitted chlamydial diseases</td>
</tr>
<tr>
<td>A59.X</td>
<td>Urogenital trichomoniasian</td>
</tr>
<tr>
<td>B37.3</td>
<td>Candidiasis of vulva and vagina</td>
</tr>
<tr>
<td>B37.4X</td>
<td>Candidiasis of other urogenital sites</td>
</tr>
<tr>
<td>L29.X</td>
<td>Pruritus</td>
</tr>
<tr>
<td>N34.X</td>
<td>Urethritis and urethral syndrome</td>
</tr>
<tr>
<td>N35.1X</td>
<td>Postinfective urethral stricture, not elsewhere classified</td>
</tr>
<tr>
<td>N37</td>
<td>Urethral disorders in diseases classified elsewhere</td>
</tr>
<tr>
<td>N72</td>
<td>Inflammatory disease of cervix uteri</td>
</tr>
<tr>
<td>N73.X</td>
<td>Other female pelvic inflammatory diseases</td>
</tr>
<tr>
<td>N75.X</td>
<td>Diseases of Bartholin's gland</td>
</tr>
<tr>
<td>N76.X</td>
<td>Other inflammation of vagina and vulva</td>
</tr>
<tr>
<td>N77.X</td>
<td>Vulvovaginal ulceration and inflammation in diseases classified elsewhere</td>
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<tr>
<td>N89.8</td>
<td>Other specified noninflammatory disorders of vagina</td>
</tr>
<tr>
<td>N89.9</td>
<td>Noninflammatory disorder of vagina, unspecified</td>
</tr>
<tr>
<td>N94.1</td>
<td>Dyspareunia</td>
</tr>
<tr>
<td>N95.2</td>
<td>Postmenopausal atrophic vaginitis</td>
</tr>
<tr>
<td>O23.X</td>
<td>Infections of genitourinary tract in pregnancy</td>
</tr>
<tr>
<td>O86.X</td>
<td>Other puerperal infections</td>
</tr>
<tr>
<td>R10.2</td>
<td>Pelvic and perineal pain</td>
</tr>
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### ICD Codes Indicating Testing Done for Genitourinary Conditions

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<tr>
<td>A51.X</td>
<td>Early syphilis</td>
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<tr>
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<td>Description</td>
</tr>
<tr>
<td>--------------------</td>
<td>-----------------------------------------------------------</td>
</tr>
<tr>
<td>A52.X</td>
<td>Late syphilis</td>
</tr>
<tr>
<td>A53.X</td>
<td>Other and unspecified syphilis</td>
</tr>
<tr>
<td>A54.X</td>
<td>Gonococcal infection</td>
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<tr>
<td>A55</td>
<td>Chlamydial lymphogranuloma (venereum)</td>
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<tr>
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<td>Other sexually transmitted chlamydial diseases</td>
</tr>
<tr>
<td>A57</td>
<td>Chancroid</td>
</tr>
<tr>
<td>A58</td>
<td>Granuloma inguinale</td>
</tr>
<tr>
<td>A59.X</td>
<td>Trichomoniasis</td>
</tr>
<tr>
<td>A60.X</td>
<td>Anogenital herpesviral [herpes simplex] infections</td>
</tr>
<tr>
<td>A63.X</td>
<td>Other predominantly sexually transmitted diseases, not elsewhere classified</td>
</tr>
<tr>
<td>A64</td>
<td>Unspecified sexually transmitted disease</td>
</tr>
<tr>
<td>A74.89</td>
<td>Other chlamydial diseases</td>
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<tr>
<td>A74.9</td>
<td>Chlamydial infection, unspecified (includes childbirth and postpartum)</td>
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<td>B37.3</td>
<td>Candidiasis of vulva and vagina</td>
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<tr>
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<td>Candidiasis of other urogenital sites</td>
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<td>Papillomavirus as the cause of diseases classified elsewhere</td>
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<td>L29.X</td>
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<td>Reiter's disease, unspecified site</td>
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<td>N35.X</td>
<td>Urethral stricture</td>
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<tr>
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<td>Urethral disorders in diseases classified elsewhere</td>
</tr>
<tr>
<td>N39.0</td>
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<td>Disorder of urinary system, unspecified</td>
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<td>N70.X</td>
<td>Salpingitis and oophoritis</td>
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<tr>
<td>N71.X</td>
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<tr>
<td>N72</td>
<td>Inflammatory disease of cervix uteri</td>
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<tr>
<td>N73.X</td>
<td>Other female pelvic inflammatory</td>
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<tr>
<td>ICD10 Code or Range</td>
<td>Description</td>
</tr>
<tr>
<td>-------------------</td>
<td>-----------------------------------------------------------------------------</td>
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<tr>
<td>N74</td>
<td>Female pelvic inflammatory disorders in diseases classified elsewhere</td>
</tr>
<tr>
<td>N75.X</td>
<td>Diseases of Bartholin's gland</td>
</tr>
<tr>
<td>N76.X</td>
<td>Other inflammation of vagina and vulva</td>
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<tr>
<td>N77.X</td>
<td>Vulvovaginal ulceration and inflammation in diseases classified elsewhere</td>
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<td>Dyspareunia</td>
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<tr>
<td>O86.X</td>
<td>Other puerperal infections</td>
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<td>R87.5</td>
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<tr>
<td>R87.6X</td>
<td>Abnormal cytological findings in specimens from female genital organs</td>
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<tr>
<td>R87.8X</td>
<td>Other abnormal findings in specimens from female genital organs</td>
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<td>Z00.00</td>
<td>Encounter for general adult medical examination without abnormal findings</td>
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<td>Z11.9</td>
<td>Encounter for screening for infectious and parasitic diseases, unspecified</td>
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<td>Z12.4</td>
<td>Encounter for screening for malignant neoplasm of cervix</td>
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<td>ICD10 Code or Range</td>
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<td>---------------------</td>
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</tr>
<tr>
<td>Z20.2</td>
<td>Contact with and (suspected) exposure to infections with a predominantly sexual mode of transmission</td>
</tr>
<tr>
<td>Z20.6</td>
<td>Contact with and (suspected) exposure to human immunodeficiency virus [HIV]</td>
</tr>
<tr>
<td>Z20.8X</td>
<td>Contact with and (suspected) exposure to communicable diseases</td>
</tr>
<tr>
<td>Z20.9</td>
<td>Contact with and (suspected) exposure to unspecified communicable disease</td>
</tr>
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<td>Z30.X</td>
<td>Encounter for contraceptive management</td>
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<tr>
<td>Z31.X</td>
<td>Encounter for procreative management</td>
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<tr>
<td>Z32.X</td>
<td>Encounter for pregnancy test and childbirth and childcare instruction</td>
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<td>Z33.X</td>
<td>Pregnant state</td>
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<td>Z34.X</td>
<td>Encounter for supervision of normal pregnancy</td>
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<td>Z36</td>
<td>Encounter for antenatal screening of mother</td>
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<tr>
<td>Z39.X</td>
<td>Encounter for maternal postpartum care and examination</td>
</tr>
<tr>
<td>Z64.0</td>
<td>Problems related to unwanted pregnancy</td>
</tr>
<tr>
<td>Z64.1</td>
<td>Problems related to multiparity</td>
</tr>
<tr>
<td>Z71.7</td>
<td>Human immunodeficiency virus [HIV] counseling</td>
</tr>
<tr>
<td>Z72.5X</td>
<td>High risk sexual behavior</td>
</tr>
<tr>
<td>Z77.9</td>
<td>Other contact with and (suspected) exposures hazardous to health</td>
</tr>
<tr>
<td>Z97.5</td>
<td>Presence of (intrauterine) contraceptive device</td>
</tr>
</tbody>
</table>

### References

**Introduction**

These references are cited in this guideline.


13. OneSwab One Vial...Multiple Pathogens. www.mdlab.com Available at: https://www.mdlab.com/forms/Brochures/OSUS.pdf
# Hereditary Cancer Syndrome Multigene Panels

## Introduction

Hereditary cancer syndrome multigene panel testing is addressed by this guideline.

## Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan’s procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>APC Sequencing</td>
<td>81201</td>
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<tr>
<td>APC Deletion/Duplication Analysis</td>
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<td>ATM Sequencing</td>
<td>81408</td>
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<td>BRCA1/2 Sequencing</td>
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<td>BRCA1/2 Deletion/Duplication Analysis</td>
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</tr>
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<tr>
<td>BRCA2 Sequencing</td>
<td>81216</td>
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<td>Chromosomal Microarray [BAC], Constitutional</td>
<td>81228</td>
</tr>
<tr>
<td>Chromosomal Microarray [SNP], Constitutional</td>
<td>81229</td>
</tr>
<tr>
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</tr>
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<tr>
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</tr>
<tr>
<td>MSH6 Sequencing</td>
<td>81298</td>
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<td>MSH6 Deletion/Duplication Analysis</td>
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<td>81317</td>
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<td>PMS2 Deletion/Duplication Analysis</td>
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</tr>
<tr>
<td>PTEN Sequencing</td>
<td>81321</td>
</tr>
<tr>
<td>PTEN Deletion/Duplication Analysis</td>
<td>81323</td>
</tr>
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<td>Procedures addressed by this guideline</td>
<td>Procedure codes</td>
</tr>
<tr>
<td>----------------------------------------</td>
<td>----------------</td>
</tr>
<tr>
<td>Hereditary breast cancer-related disorders (eg, hereditary breast cancer, hereditary ovarian cancer, hereditary endometrial cancer); genomic sequence analysis panel, must include sequencing of at least 10 genes, always including BRCA1, BRCA2, CDH1, MLH1, MSH2, MSH6, PALB2, PTEN, STK11, and TP53</td>
<td>81432</td>
</tr>
<tr>
<td>Hereditary colon cancer disorders (eg, Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis); genomic sequence analysis panel, must include sequencing of at least 10 genes, including APC, BMPR1A, CDH1, MLH1, MSH2, MSH6, MUTYH, PTEN, SMAD4, and STK11</td>
<td>81435</td>
</tr>
<tr>
<td>Hereditary colon cancer disorders (eg, Lynch syndrome, PTEN hamartoma syndrome, Cowden syndrome, familial adenomatosis polyposis); duplication/deletion analysis panel, must include analysis of at least 5 genes, including MLH1, MSH2, EPCAM, SMAD4, and STK11</td>
<td>81436</td>
</tr>
<tr>
<td>Hereditary neuroendocrine tumor disorders (eg, medullary thyroid carcinoma, parathyroid carcinoma, malignant pheochromocytoma or paraganglioma); genomic sequence analysis panel, must include sequencing of at least 6 genes, including MAX, SDHB, SDHC, SDHD, TMEM127, and VHL</td>
<td>81437</td>
</tr>
<tr>
<td>Hereditary neuroendocrine tumor disorders (eg, medullary thyroid carcinoma, parathyroid carcinoma,</td>
<td>81438</td>
</tr>
</tbody>
</table>
Procedures addressed by this guideline | Procedure codes
--- | ---
malignant pheochromocytoma or paraganglioma); duplication/deletion analysis panel, must include analyses for SDHB, SDHC, SDHD, and VHL | 
Miscellaneous hereditary cancer syndrome gene tests | 81400
Miscellaneous hereditary cancer syndrome gene tests | 81401
Miscellaneous hereditary cancer syndrome gene tests | 81402
Miscellaneous hereditary cancer syndrome gene tests | 81403
Miscellaneous hereditary cancer syndrome gene tests | 81404
Miscellaneous hereditary cancer syndrome gene tests | 81405
Miscellaneous hereditary cancer syndrome gene tests | 81406
Miscellaneous hereditary cancer syndrome gene tests | 81407
Miscellaneous hereditary cancer syndrome gene tests | 81408
Miscellaneous hereditary cancer syndrome gene tests | 81479

What are hereditary cancer syndromes

Definition

A hereditary cancer syndrome is when a mutation in a single gene causes a significantly increased risk for certain cancers. Hereditary cancer syndromes are usually characterized by a pattern of specific cancer types occurring together in the same family, younger ages of cancer diagnosis than usual, or other co-existing non-cancer conditions.

Prevalence

Most cancer is sporadic and believed to be caused by a mix of behavioral or lifestyle, environmental, and inherited risk factors. However, about 5-10% of cancers are believed to have a major inherited component.1
Hereditary cancer syndromes

There are at least 50 hereditary cancer syndromes. This table lists some of the most common along with associated cancers.

<table>
<thead>
<tr>
<th>Syndrome</th>
<th>Associated cancers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Hereditary breast and ovarian cancer syndrome (HBOC)</td>
<td>breast, ovarian, fallopian tube, or primary peritoneal cancer, pancreatic, prostate</td>
</tr>
<tr>
<td>Lynch syndrome</td>
<td>colorectal, endometrial, small bowel, stomach, ovarian, pancreatic, ureteral and renal pelvis, biliary tract, brain, sebaceous adenoma, keratoacanthoma tumors</td>
</tr>
<tr>
<td>Familial adenomatous polyposis</td>
<td>colorectal and other gastrointestinal cancers, gastrointestinal tract polyps such as adenomas and fundic gland, osteomas, desmoids, thyroid cancer, hepatoblastoma</td>
</tr>
<tr>
<td>MUTYH-associated polyposis</td>
<td>colorectal and other gastrointestinal cancers</td>
</tr>
</tbody>
</table>

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<table>
<thead>
<tr>
<th>Syndrome</th>
<th>Associated cancers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cowden syndrome</td>
<td>• benign and malignant tumors of the breast, endometrium, and thyroid</td>
</tr>
<tr>
<td></td>
<td>• cancer and polyps (hamartomas) in the colon and rectum</td>
</tr>
<tr>
<td>Li Fraumeni syndrome</td>
<td>• soft tissue sarcoma</td>
</tr>
<tr>
<td></td>
<td>• osteosarcoma</td>
</tr>
<tr>
<td></td>
<td>• leukemia</td>
</tr>
<tr>
<td></td>
<td>• melanoma</td>
</tr>
<tr>
<td></td>
<td>• breast</td>
</tr>
<tr>
<td></td>
<td>• pancreas</td>
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<tr>
<td></td>
<td>• colon</td>
</tr>
<tr>
<td></td>
<td>• adrenal cortex</td>
</tr>
<tr>
<td></td>
<td>• stomach</td>
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<tr>
<td></td>
<td>• esophagus</td>
</tr>
<tr>
<td></td>
<td>• brain</td>
</tr>
<tr>
<td>Peutz-Jeghers syndrome</td>
<td>• polyps (hamartomas) in the stomach</td>
</tr>
<tr>
<td></td>
<td>• small intestine and colon</td>
</tr>
<tr>
<td></td>
<td>• pancreas</td>
</tr>
<tr>
<td></td>
<td>• lung</td>
</tr>
<tr>
<td></td>
<td>• breast</td>
</tr>
<tr>
<td></td>
<td>• uterine</td>
</tr>
<tr>
<td></td>
<td>• ovarian</td>
</tr>
</tbody>
</table>

**Overlapping clinical findings**

Many hereditary cancer syndromes can include the same types of cancer and therefore have overlapping clinical findings. For example, breast cancer is a feature of HBOC caused by BRCA mutations, Li Fraumeni syndrome, Cowden syndrome, and others. Sometimes, the pattern of cancers in the family or pathognomonic features makes the underlying syndrome clear. However, in many cases it can be
difficult to reliably diagnose hereditary cancer syndromes based on clinical and family history alone.

Test information

Introduction

Testing for hereditary cancer syndromes may include multigene panel testing.

Sanger Sequencing

Until recently, most sequencing tests used the Sanger sequencing methodology that was originally developed in the 1970s. Sanger sequencing is labor intensive and did not lend itself to high-throughput applications.

Next-generation sequencing (NGS)

NGS, which is also sometimes called massively parallel sequencing, has been developing since about 2005 to allow larger scale and more efficient gene sequencing. NGS relies on sequencing many copies of small pieces of DNA simultaneously and using bioinformatics to assemble the sequence.

The efficiency of NGS has led to an increasing number of large, multi-gene testing panels. NGS panels that test several genes at once are particularly well-suited to conditions caused by more than one gene or where there is considerable clinical overlap between conditions making it difficult to reliably narrow down likely causes. As a result, several laboratories have begun to combine genes involved in causing various hereditary cancer syndromes, which often have both of those characteristics.

Detection rate of NGS

NGS may not perform as well as Sanger sequencing in some applications. Results may also be obtained that cannot be adequately interpreted based on the current knowledgebase.

- When a sequence variation is identified that has not been previously characterized or shown to cause the disorder in question, it is called a variant of uncertain significance (VUS). VUSs are relatively common findings when sequencing large amounts of DNA with NGS.
Under certain circumstances, technologies used in multi-gene testing may fail to identify mutations that might be identifiable through single-gene testing. If high clinical suspicion remains for a particular syndrome after negative multi-gene test results, consultation with the testing lab, additional targeted genetic testing, or both may be warranted.

**Hereditary cancer syndrome multi-gene panels**

Hereditary cancer syndrome multi-gene panels include a wide variety of genes and may be focused on the genetic causes of a particular cancer type or broad detection of common hereditary cancer syndromes.

Multi-gene tests vary in technical specifications. For example, different labs may have different depth of coverage, extent of Intron/Exon Boundary analysis, or methodology of large Deletion/Duplication Analysis.

Because genes can be easily added or removed from multi-gene tests over time by a given lab, medical records must document:

- the genes included in the specific multi-gene test used from each patient, and
- the labs that performed the panels.

**Moderate risk genes**

Panels may also include genes believed to be associated with cancer, but with a more modest impact on risk than recognized hereditary cancer syndromes. Results for such genes are of less clear value because there often are not clear management recommendation for mutation-positive individuals.

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to hereditary cancer syndrome panel testing.

**National Comprehensive Cancer Network (NCCN)**

The National Comprehensive Cancer Network (NCCN) makes the following general recommendations for using multi-gene panels in evaluating risk for breast and ovarian cancer and now includes this option in some management algorithms:

"Because of their complexity, multi-gene testing is ideally offered in the context of professional genetic expertise for pre- and post-test counseling."

"Testing of an individual without a cancer diagnosis should only be considered when an appropriate affected family member is unavailable for testing."
“When more than one gene can explain an inherited cancer syndrome, then multi-gene testing may be more efficient and/or cost effective. As commercially available tests differ in the specific genes analyzed (as well as classification of variants and many other factors), choosing the specific laboratory and test panel is important. Multi-gene testing can include “intermediate” penetrant (moderate-risk) genes. For many of these genes, there is limited data on the degree of cancer risk and there are no clear guidelines on risk management for carriers of mutations. Not all genes included on available multi-gene tests are necessarily clinically actionable.” If a moderate risk gene mutation is identified, “gene carriers should be encouraged to participate in clinical trials or genetic registries.”

“Mutations in many breast cancer susceptibility genes involved in DNA repair may be associated with the rare autosomal recessive condition, Fanconi anemia.” Therefore, multi-gene testing may unexpectedly reveal that an individual and their family are at an increased risk for this condition.

“There is an increased likelihood of finding variants of unknown significance when testing for mutations in multiple genes.”

American College of Medical Genetics

The American College of Medical Genetics has a policy statement that offers general guidance on the clinical application of large-scale sequencing focusing primarily on whole exome and whole genome testing. However, some of the recommendations regarding counseling around unexpected results, variants of unknown significance, and minimum requirements for reporting apply to many NGS applications.

Criteria

Introduction

Requests for hereditary cancer syndrome panel testing are reviewed using these criteria.

Criteria

This guideline applies to all hereditary cancer syndrome panels, which are defined as assays that simultaneously test for more than one hereditary cancer syndrome. This guideline does not apply when testing more than one gene related to the same hereditary cancer syndrome (e.g., Lynch syndrome).

Medical necessity coverage generally relies on criteria established for testing individual hereditary cancer syndromes. See the Coverage Guidance table for examples of genes known to be included in currently available hereditary cancer syndrome multi-gene panels with coverage guidance. This is not intended to be a complete list of available genes as these panels are evolving rapidly.
However, this guideline takes into account the efficiency gains from simultaneously testing multiple candidate genes. Therefore, coverage requirements rely to some degree on how the panel will be billed. Panels may be billed in a variety of ways:

- **Gene sequencing portion:**
  - A separate CPT code for sequencing each gene studied or a subset (e.g., 81201, 81294, 81297, etc.)
  - A single CPT code developed specifically for a particular type of panel (e.g., 81432, 81435, 81437)
  - A single unlisted CPT code (e.g., 81479)

- **Deletion/duplication analysis portion:**
  - A separate CPT code for deletion/duplication analysis of each gene studied or a subset (e.g., 81203, 81292, 81294, 81404, 81479, etc.)
  - A single CPT code developed specifically for a particular type of panel (e.g., 81433, 81436, 81438)
  - Microarray analysis (e.g., 81228 or 81229)
  - Part of a single unlisted CPT code for the sequencing and deletion/duplication portions of the panel (e.g., 81479)

Hereditary cancer syndrome multi-gene panels will be reimbursed when the following criteria are met:

- Panel will be billed with separate procedure codes for each gene analyzed (however, please note that the billed amount should not exceed the list price of the test).
  - The medical necessity of each billed procedure will be assessed independently. See the Coverage Guidance table for gene-specific policy guidance.
    - When a patient meets medical necessity criteria for any hereditary cancer syndrome gene(s) included in a multi-gene panel, genetic testing for the clinically indicated gene(s) will be reimbursed. This includes the sequencing and deletion/duplication components.
Any genes that are included in a multi-gene panel but do NOT meet medical necessity criteria will NOT be reimbursed. It will be at the laboratory, provider, and patient’s discretion to determine if a multi-gene panel remains the preferred testing option.

- Sequencing and/or deletion/duplication analysis† of any hereditary cancer syndrome gene(s) should only be performed once per lifetime and will therefore only be reimbursed once per lifetime. If gene testing was previously performed, and is now being included in a panel, such testing will not be separately reimbursable regardless of whether clinical coverage criteria are met, OR

- Panel will be billed with a single procedure code to represent all genes being sequenced, with or without another single procedure code representing the deletion/duplication analysis† portion. Code(s) may be specific to that panel or an unlisted code, such as 81479.

- No previous hereditary cancer syndrome testing has been performed

  - Medical necessity must be established for at least two conditions included in the panel (e.g., hereditary breast and ovarian cancer and Li Fraumeni syndrome). Note that this is two conditions and not two genes (i.e., meeting criteria for only Lynch syndrome, which is caused by mutations in at least 5 genes, would not fulfill criteria alone).

    Although not a complete list, the following are considered separate conditions:
    - Hereditary breast cancer - this includes both BRCA1/2 and PALB2.
    - Lynch syndrome
    - Li-Fraumeni
    - Familial adenomatous polyposis
    - Cowden syndrome

- Testing for one condition was performed and billed separately. A multi-gene panel is now being considered as a reflex and will be billed at a rate comparable to single syndrome pricing (e.g., myRisk update).

  - Medical necessity must be established for at least one condition included in the panel in addition to the already tested condition (e.g., hereditary breast and ovarian cancer was already performed, but Lynch syndrome criteria are also met).

  - Note that if BRCA1/2 testing was already performed and PALB2 criteria are now met, PALB2 testing alone would be reimbursable and not a reflex panel test (e.g. myRisk Update).

† When deletion/duplication testing is not part of a single panel CPT code being billed, deletion/duplication testing should be billed in only one of the following ways:
• A separate CPT code for deletion/duplication analysis of each individual gene (may include non-specific molecular pathology tier 2 codes or unlisted code 81479), or
• A single CPT code specific to the performed deletion/duplication analysis panel, or
• A single microarray procedure

Procedure codes representing multiple methods for deletion/duplication testing will not be reimbursable for the same panel (e.g., test-specific deletion/duplication procedure codes and microarray will not both be reimbursable for the same panel).

Coverage guidance

The following table describes coverage guidance for genes associated with hereditary cancer syndromes.

Coverage Guidance for Genes Included in Hereditary Cancer Syndrome Multi-Gene Panels

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**Not reimbursed**

Gene testing is not reimbursed strictly for hereditary cancer indications. In general, this category applies to genes that have only a low to moderate impact on cancer risk (compared to high penetrance cancer syndrome-causing genes) and no clear management guidelines associated with identifying a mutation.

**References**

**Introduction**

These references are cited in this guideline.


# Hereditary Connective Tissue Disorder Testing

MOL.TS.268.A  
**v2.0.2018**

## Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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**What are hereditary connective tissue disorders**

**Definition**

Hereditary connective tissue disorders (HCTDs) are a group of disorders that affect the connective tissues that support the skin, bones, joints, heart, blood vessels, eyes, and other organs.\(^1\)

- While specific features vary by type, an unusually large range of joint movement (hypermobility) and cardiovascular disease (such as thoracic aortic aneurysms and dissections, or TAAD) are features that are present in many HCTDs. Medical management may differ based on the underlying genetic etiology.

- In many cases, a careful clinical examination by a specialist familiar with clinical features of these conditions can help to point toward one condition. In these cases, testing for gene(s) associated with a single condition would be most appropriate. However, in some cases, it can be difficult to reliably diagnose an HCTD based on clinical and family history alone.

- More than a half million people in the United States are estimated to have an HCTD.\(^1\)

- There are more than 200 HCTDs.\(^1\) Some of the most common types are summarized below:

  - **Arterial tortuosity syndrome (ATS)** — An autosomal recessive disorder associated with severe and widespread tortuosity of the aorta and middle-sized arteries, with an increased risk of aneurysms and dissections. Other features include stenosis of the aorta and/or pulmonary arteries, characteristic facies with high palate and dental crowding, and soft/doughy skin. Additional connective tissue disorder features that may be present include skeletal findings (scoliosis, pectus anomalies, joint laxity), hernias, hypotonia, and ocular involvement (myopia, keratoconus). SLC2A10 is the only gene known to be associated with ATS. Sequence variants are the most common; exon deletions have been reported in a couple cases.\(^2\)

  - **Congenital contractural arachnodactyly (Beals syndrome)** — An autosomal dominant disorder characterized by a Marfan-like appearance (tall, slender habitus in which arm span exceeds height) and long, slender fingers and toes (arachnodactyly). Most affected individuals have a "crumpled" appearance to their ears and most have contractures of major joints (knees and ankles) at birth. Hip contractures, adducted thumbs, and club foot may occur. The majority of affected individuals have muscular hypoplasia. Kyphosis/scoliosis is present...
in about half of all affected individuals. Dilatation of the aorta is occasionally present. 27-75% of affected individuals have a mutation in the FBN2 gene.³

- **Cutis laxa** — A group of disorders characterized by lax, sagging skin that often hangs in loose folds, causing the face and other parts of the body to have a droopy appearance. Extremely wrinkled skin may be particularly noticeable on the neck and in the armpits and groin. Other features may include arterial aneurysm and dissection, emphysema, and inguinal or umbilical hernia. There are autosomal dominant, autosomal recessive, and X-linked forms. Causative autosomal genes include ELN, FBLN5, ATP6V0A2, and EFEMP2. The X-linked form is due to mutations in ATP7A (see also Occipital Horn Syndrome).⁴

- **Ehlers Danlos syndromes (EDS)** — A heterogeneous group of disorders, the majority of which share the features of joint hypermobility and skin involvement. There are 13 types: classical, classical-like, cardiac-valvular, vascular, hypermobile (includes “joint hypermobility syndrome”), arthrochalasia, dermatosparaxis, kyphoscoliotic, spondylodysplastic, muscular contractural, myopathic, periodontal, and brittle cornea syndrome. Some types have autosomal dominant inheritance, while others are autosomal recessive. Hypermobile type is the most common, but its genetic etiology is currently unknown. Genetic testing is available for the other EDS types (see Table 1 below for a list of genes).⁵,⁶

- **Homocystinuria due to cystathionine beta-synthase deficiency** — An autosomal recessive metabolic disorder in which affected individuals have markedly elevated plasma total homocysteine and methionine. Clinical features include involvement of the eye (ectopia lentis and/or severe myopia), skeletal system (excessive height, long limbs, scoliosis, and pectus excavatum), and vascular system (thromboembolism). Many have developmental delay/intellectual disability. Treatment involves maintenance of normal or near-normal plasma homocysteine concentrations using a specialized diet and vitamin supplementation. The diagnosis can be substantiated by detection of biallelic pathogenic mutations in the CBS gene. Sequence analysis detects 95-98% of mutations, while deletion/duplication analysis detects <5%.⁷

- **Loeys-Dietz syndrome (LDS)** — LDS is an autosomal dominant disorder that affects many parts of the body.⁸ LDS is mostly caused by mutations in either the TGFBR1 gene (20%) or TGFBR2 gene (70%). However, a small percentage of people with LDS may have mutations in the SMAD3 gene (5%), TGFBR2 gene (1%) or TGFBR3. Major manifestations of this condition include “vascular findings (dilatation or dissection of the aorta, other arterial aneurysms or tortuosity), skeletal findings (pectus excavatum or pectus carinatum, scoliosis, joint laxity or contracture, long thin fingers and toes, cervical spine malformation and/or instability), craniofacial findings (widely spaced eyes, bifid uvula/cleft palate, craniosynostosis), and cutaneous findings (translucent skin, easy bruising, dystrophic scars).⁹ Given that there is no clinical diagnostic criteria established for LDS, genetic testing can help with the diagnosis.⁸
o **Marfan syndrome (MFS)** — MFS is an autosomal dominant disorder that affects connective tissue in many parts of the body. MFS is caused by mutations in the FBN1 gene. Approximately 70-93% of people meeting diagnostic criteria for MFS will have a mutation in this gene. Diagnostic criteria, called the Ghent criteria, exists for MFS. Major manifestations of the disease include aortic enlargement and ectopia lentis. Other features include, but are not limited to, bone overgrowth and joint laxity, long arms and legs, scoliosis, sternum deformity (pectus excavatum or carinatum), long thin fingers and toes, dural ectasia (stretching of the dural sac), hernias, stretch marks on the skin, and lung bullae. Symptoms can present in males or females at any age. Symptoms typically worsen over time. Infants who present with symptoms typically have the most severe disease course.  

o **NOTCH1-related aortic valve disease** — NOTCH1 variants can be associated with autosomal dominant congenital heart defects affecting the left ventricular outflow tract (LVOT), most commonly bicuspid aortic valve (BAV). Adult-onset aortic valve calcification is a frequent feature. NOTCH1 variants have also been identified in 4.2% of individuals with sporadic BAV and much less frequently with other LVOT malformations. Mutations in this gene are also associated with Adams-Oliver syndrome, which is characterized by aplasia cutis congenita of the scalp and malformations of the limbs, brain, and cardiovascular system.  

o **Osteogenesis imperfecta (OI)** — A group of disorders associated with a propensity to fractures with little or no trauma. Additional features may include skeletal anomalies, short stature, hearing loss, and blue/gray sclera. The severity is highly variable, ranging from a mild form with few fractures and normal life expectancy, to severe forms with neonatal lethality. OI types I-IV account for the majority of cases, and are caused by heterozygous mutations in the COL1A1 and COL1A2 genes. Inheritance is autosomal dominant. Autosomal recessive forms of OI are rare, and can be associated with mutations in a number of different genes.  

o **Periventricular nodular heterotopia (PVNH)** — An X-linked condition, which is prenatally or neonatally lethal in most males. Therefore, most affected individuals are female. In addition to PVNH, some individuals have connective tissue anomalies such as joint hypermobility, aortic dilation, and other vascular anomalies. 93% of individuals with FLNA-related PVNH have a sequence variant; genomic rearrangements have been reported in a few cases.  

o **Stickler syndrome** — A disorder characterized by ocular findings (myopia, cataract and retinal detachment), hearing loss, craniofacial findings (midfacial underdevelopment and cleft palate), mild spondyloepiphyseal dysplasia and/or early-onset arthritis. Clinical diagnostic criteria are available. >90% of cases are due to mutations in COL2A1 or COL11A1. Mutations in these genes, and COL11A2, are inherited in an autosomal dominant pattern. Mutations in COL9A1, COL9A2, and COL9A3 are rare, and inherited in an autosomal recessive pattern.
Thoracic Aortic Aneurysm and Dissection (TAAD) — Familial TAAD is defined as dilatation and/or dissection of the thoracic aorta, absence of clinical features of MFS, LDS or vascular EDS, and a positive family history of TAAD. Approximately 30% of families with TAAD who do not have a clinical diagnosis of MFS or another syndrome have a causative mutation in one of 13 known TAAD-related genes (see Table 1 below).14

Test information
• Clinical genetic testing is available for many HCTDs. However, hypermobile EDS (hEDS), joint hypermobility syndrome, and nonsyndromic joint hypermobility, including “hypermobility spectrum disorders”, continue to require a clinical diagnosis, since the genetic etiology of these disorders is not yet known.7
• Until recently, most sequencing tests used the Sanger sequencing methodology that was originally developed in the 1970s. Sanger sequencing is labor intensive and did not lend itself to high-throughput applications.
• Next generation sequencing (NGS), which is also sometimes called massively parallel sequencing, has been developing since about 2005 to allow larger scale and more efficient gene sequencing. NGS relies on sequencing many copies of small pieces of DNA simultaneously and using bioinformatics to assemble the sequence.
• The efficiency of NGS has led to an increasing number of large, multi-gene testing panels. NGS panels that test several genes at once are particularly well-suited to conditions caused by more than one gene or where there is considerable clinical overlap between conditions, making it difficult to reliably narrow down likely causes. As a result, several laboratories have begun to combine genes involved in certain conditions, which often have both of those characteristics. However, NGS may not perform as well as Sanger sequencing in some applications. Results may also be obtained that cannot be adequately interpreted based on the current knowledgebase. When a sequence variation is identified that has not been previously characterized or shown to cause the disorder in question, it is called a variant of uncertain significance (VUS). VUSs are relatively common findings when sequencing large amounts of DNA with NGS.
• HCTD multi-gene panels include a wide variety of genes associated with connective tissue disorders. Multi-gene panels may also include genes believed to be associated with HCTDs, but with a lower impact on risk than recognized syndromes. Results for such genes are of less clear value because there often are not clear management recommendation for mutation-positive individuals.
• Under certain circumstances, technologies used in multi-gene testing may fail to identify mutations that might be identifiable through single-gene testing. If high clinical suspicion remains for a particular syndrome after negative multi-gene test results, consultation with the testing lab and/or additional targeted genetic testing may be warranted.
• Multi-gene tests vary in technical specifications (e.g., depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis).
• Since genes can be easily added or removed from multi-gene tests over time by a given lab, medical records must document which genes were included in the specific multi-gene test used from each patient, and in which labs they were performed.

• Additionally, tests should be chosen that maximize the likelihood of identifying mutations in the genes of interest and that will alter patient management.

Guidelines and evidence

• No current U.S guidelines address the use of multi-gene panels in HCTDs.

• An expert-authored review (updated in 2016)\textsuperscript{15} states the following regarding hEDS: “If a patient's personal or family history is suggestive of one of the other types of EDS or another hereditary disorder of connective tissue or arterial fragility syndrome, analysis of an associated gene or multi-gene connective tissue disease panel may be appropriate. Failure to identify a pathogenic variant with such multiple gene testing reduces the likelihood of an arterial fragility syndrome, but does not completely rule it out, especially in the setting of a positive personal or family history of arterial fragility. Negative testing for an arterial fragility syndrome also does not confirm a diagnosis of EDS, hypermobility type. Therefore, such testing is not recommended in the absence of specific suggestive signs, symptoms, or family history.”

• According to the International Consortium on the Ehlers- Danlos Syndromes (2017)\textsuperscript{6}:
  \begin{itemize}
    \item “In view of the vast genetic heterogeneity and phenotypic variability of the EDS subtypes, and the clinical overlap between many of these subtypes, but also with other HCTDs, the definite diagnosis relies for all subtypes, except hEDS, on molecular confirmation with identification of (a) causative variant(s) in the respective gene.”
    \item “Molecular diagnostic strategies should rely on NGS technologies, which offer the potential for parallel sequencing of multiple genes. Targeted resequencing of a panel of genes...is a time- and cost-effective approach for the molecular diagnosis of the genetically heterogeneous EDS. When no mutation (or in case of an autosomal recessive condition only one mutation) is identified, this approach should be complemented with a copy number variant (CNV) detection strategy to identify large deletions or duplications, for example Multiplex Ligation-dependent Probe Amplification (MLPA), qPCR, or targeted array analysis.”
  \end{itemize}
“The diagnosis of hEDS remains clinical as there is yet no reliable or appreciable genetic etiology to test for in the vast majority of patients.”

Criteria

This guideline applies to hereditary connective tissue disorder testing, including single genes as well as multi-gene panels, which are defined as assays that simultaneously test for more than one hereditary connective tissue disorder gene. Medical necessity determination generally relies on criteria established for testing individual genes.

Medical necessity criteria differ based on the type of testing being performed (i.e., individual hereditary connective tissue disorder genes separately chosen versus pre-defined panels of genes) and how that testing will be billed (one or more individual gene procedure codes, specific panel procedure codes, or unlisted procedure codes).

**Hereditary Connective Tissue Disorder single gene tests will be reimbursed when the following criteria are met:**

- The member has a condition that will benefit from information provided by the requested hereditary connective tissue disorder gene testing based on at least one of the following:
  - The member displays clinical features of the condition for which testing is being requested and a genetic diagnosis would result in changes to the member’s medical management, OR
  - The member meets all criteria in a test-specific guideline, if available (see table: Common hereditary connective tissue disorder genes, associated conditions, and applicable guidelines), AND
- The member does not have a known underlying cause for their symptoms (e.g. known genetic condition), AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Hereditary Connective Tissue Disorder multi-gene panels will be reimbursed when the following criteria are met:**

When separate procedure codes will be billed for individual hereditary connective tissue disorder genes (e.g., Tier 1 MoPath codes 81200-81355 or Tier 2 MoPath codes 81400-81408), each individually billed test will be evaluated separately. The following criteria will be applied:

- The member has a condition that will benefit from information provided by the requested hereditary connective tissue disorder gene testing based on at least one of the following:
  - The member displays clinical features of the condition for which testing is being requested and a genetic diagnosis would result in changes to the member’s medical management, OR
The member meets all criteria in a test-specific guideline, if available, (see Common hereditary connective tissue disorder genes, associated conditions, and applicable guidelines table for a list of genes, associated conditions, and applicable guideline), AND

- The member does not have a known underlying cause for their symptoms (e.g. known genetic condition), AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

When a patient meets medical necessity criteria for any hereditary connective tissue disorder gene(s) included in the panel, genetic testing for the clinically indicated gene(s) will be reimbursed. This includes the sequencing and deletion/duplication components. Any genes that are included in a multi-gene panel but do NOT meet medical necessity criteria will NOT be a reimbursable service. It will be at the laboratory, provider, and patient’s discretion to determine if a multi-gene panel remains the preferred testing option.

When a multi-gene panel is being requested and will be billed with a single panel CPT code (e.g. 81410, 81479) to represent all genes being sequenced, with or without another single procedure code representing the deletion/duplication analysis portion, the panel will be considered medically necessary when the following criteria are met:

- Medical necessity must be established for at least TWO conditions included in the panel based on the following:
  - The member displays clinical features of the condition for which testing is being requested and a genetic diagnosis would result in changes to the member’s medical management, OR
  - The member meets all criteria in a test-specific guideline, if available, (see table: Common hereditary connective tissue disorder genes, associated conditions, and applicable guidelines), AND
- The member does not have a known underlying cause for their symptoms (e.g. known genetic condition), AND
- Clinical features are not sufficiently specific to suggest a single causative gene, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

† When deletion/duplication testing is not part of a single panel CPT code being billed, deletion/duplication testing should be billed in only one of the following ways:

- A separate CPT code for deletion/duplication analysis of each individual gene (may include non-specific molecular pathology tier 2 codes and/or unlisted code 81479), or
- A single CPT code specific to the performed deletion/duplication analysis panel (e.g. 81411, 81479), or
• A single microarray procedure (e.g. 81228 or 81229).

Procedure codes representing multiple methods for deletion/duplication testing will not be reimbursable for the same panel (e.g., test-specific deletion/duplication procedure codes and microarray will not both be reimbursable for the same panel).

Exceptions

The following are specifically non-reimbursed indications for Hereditary Connective Tissue Disorder testing:

• Hypermobile EDS or the related clinical entity, “joint hypermobility syndrome”
• Nonsyndromic joint hypermobility, including both asymptomatic and symptomatic forms (e.g., “hypermobility spectrum disorders”)

Billing and reimbursement considerations

• The billed amount should not exceed the list price of the test.
• Broad connective tissue disorder panels may not be medically necessary when a narrower panel is available and more appropriate based on the clinical findings.
• Genetic testing is only necessary once per lifetime. Therefore, a single gene included in a panel or a multi-gene panel may not be reimbursed if testing has been performed previously. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.
• This guideline may not apply to genetic testing for indications that are addressed in test-specific guidelines. Please see the test-specific list of guidelines for a complete list of test-specific panel guidelines.
• If a panel was previously performed and an updated, larger panel is being requested, only testing for the medically necessary, previously untested genes will be reimbursable. Therefore, only the most appropriate procedure codes for those additional genes will be considered for reimbursement.

Common hereditary connective tissue disorder genes, associated conditions, and applicable guidelines

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### Hereditary Connective Tissue Disorder

#### References

1. **What Are Heritable Disorders of Connective Tissue? Fast Facts: An Easy-to-Read Series of Publications for the Public.** (Updated November 2014). Copyright, National Institute of Arthritis and Musculoskeletal and Skin Disease and National Institutes of Health. Available at:  
   https://www.niams.nih.gov/health_info/Connective_Tissue/connective_tissue_ff.asp


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**Note**: Several genes in this table are associated with multiple genetic disorders, including some not listed above. The test should be reviewed for the appropriate condition/indication.


Hereditary Hemochromatosis Testing

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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What is hereditary hemochromatosis

Definition

Hereditary hemochromatosis (HH) is an autosomal recessive genetic disorder that leads to excess iron absorption and storage in the liver, heart, pancreas, and other organs.¹

- Symptoms of hemochromatosis may include:¹,²
  - Hepatomegaly, liver disease, jaundice, cirrhosis, liver cancer
  - Heart disease, arrhythmia, cardiomyopathy
  - Unexplained weakness, chronic fatigue, apathy
  - Arthritis, arthralgia
  - Increased skin pigmentation (bronze color)
  - Weight loss, hair loss
  - Hypothyroidism, hypopituitarism
  - Amenorrhea, early menopause
  - Loss of libido, impotence
  - Adult-onset diabetes

- HH is caused by mutations in the HFE gene.¹ About 1 in 200 to 1 in 300 people in the U.S. are affected with HH.²

- HH is most common in Caucasians, with up to 11% of the population being carriers. The disorder is less common in African Americans and Hispanics, with the carrier prevalence being 2.3% and 3% respectively. HH is very rare in Asians, with less than 1 in 1000 being carriers.¹
HH can be effectively treated in most people. Phlebotomy therapy can alleviate almost all symptoms of iron overload if initiated before organ damage occurs.\(^3\)

When hemochromatosis is suspected, serum iron studies, including serum ferritin and transferrin saturation, are the first step in establishing a diagnosis. Genetic testing of the HFE gene may follow if serum iron studies suggest the presence of iron overload.\(^4\)

Current guidelines support HFE genetic testing in people with:\(^2,4\)

- Serologic evidence of iron overload, considered to be a transferrin saturation >45% and elevated ferritin
- A known family history of hemochromatosis
- A known familial mutation in the HFE gene in a first degree relative

**Test information**

- HFE Mutation Analysis
  - Common changes in the HFE gene associated with HH are C282Y, H63D, and S65C.\(^1\)
  - C282Y and H63D are the most common and account for 87% of hereditary hemochromatosis in European populations.\(^1\) The next most common cause are individually rare mutations.\(^5\) Many labs do not test for S65C because it accounts for <1% of hereditary hemochromatosis.\(^1\) There is controversy over whether the H63D variant causes clinical disease\(^2\). The combination of these mutations determines both the chances of symptoms occurring and their severity.

**Guidelines and evidence**

- The American Association for the Study of Liver Diseases (AASLD) Practice Guidelines (2011):\(^6\)
  - “In a patient with suggestive symptoms, physical findings, or family history, a combination of transferrin saturation (TS) and ferritin should be obtained rather than relying on a single test. (1B) If either is abnormal (TS ≥45% or ferritin above the upper limit of normal), then HFE mutation analysis should be performed. (1B)”
“The guideline developers recommend screening (iron studies and HFE mutation analysis) of first-degree relatives of patients with HFE-related HH to detect early disease and prevent complications”

Screening for Hereditary Hemochromatosis: A Clinical Practice Guideline from the American College of Physicians (2005):  

“Physicians should discuss the risks, benefits, and limitations of genetic testing in patients with a positive family history of hereditary hemochromatosis or those with elevated serum ferritin level or transferrin saturation. Before genetic testing, individuals should be made aware of the benefits and risks of genetic testing. This should include discussing available treatment and its efficacy; costs involved; and social issues, such as impact of disease labeling, insurability and psychological well-being, and the possibility of as-yet-unknown genotypes associated with hereditary hemochromatosis.”

Criteria

HFE known familial mutation testing

• Clinical Consultation:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous genetic testing of the HFE gene, AND
• Presymptomatic/Asymptomatic Genetic Testing:
  o HFE mutation identified in 1st degree biological relative, OR
• Diagnostic Testing:
  o Serologic evidence of iron overload, defined as transferrin saturation greater than or equal to 45% and/or elevated ferritin, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

HFE targeted mutation testing

• Clinical Consultation:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous genetic testing of the HFE gene, AND
• Presymptomatic/Asymptomatic Genetic Testing:
  o Documented family history of first-degree relative with HFE-related HH, OR
• Diagnostic Testing:
  o Serologic evidence of iron overload, defined as transferrin saturation greater than or equal to 45% and/or elevated ferritin, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


HIV Genotype and Phenotype Testing

Introduction

HIV genotype and phenotype testing is addressed by this guideline.

Procedures addressed

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Development of drug resistance to antiretroviral therapy

Human immunodeficiency virus (HIV) replicates rapidly, particularly in response to antiretroviral (ARV) therapies.

These replications are a result of substitutions in the viral protease, reverse transcriptase, or integrase enzymes which are targeted by various ARV drugs that can
lead to drug-resistant mutations (quasi species). These mutations lead to virologic
treatment failure.

Because of the rapidity of these replications, highly active antiretroviral therapy
(HAART) has been designed to use treatments with multiple mechanisms of action in
order to reduce mutations and the development of drug-resistant variants. The drug
included in this regimen include, but are not limited to, nucleoside reverse transcriptase
inhibitors (NRTIs), nonnucleoside reverse transcriptase inhibitors (NNRTIs), protease
inhibitors (PIs), integrase inhibitors, and fusion inhibitors.

**Effectiveness of HAART**

The effectiveness of HAART in suppressing HIV replication for prolonged periods has
allowed HIV to be considered a chronic, rather than a fatal, disease.

However, drug failure continues to occur because HAART regimens do not completely
suppress replication in all cases. Other factors, such as nonadherence, inadequate
potency of treatment, or suboptimal drug levels may play a part in drug failure.

Historically, the initiation of, or a change in, ARV therapy has been based on HIV
ribonucleic acid (RNA) levels and CD4 cell counts, both of which are essential
components of the clinical management of HIV. Because treatment decisions can
irrevocably alter an individual's response to future therapy, clinicians can also utilize
genotypic or phenotypic assays as additional clinical tools for selecting safe and
efficacious treatment regimens.

**Test information**

**Introduction**

HIV phenotype and genotype testing may include genotypic resistance assays,
phenotypic resistance assays, or virtual phenotyping.

**Genotypic resistance assays**

Genotypic resistance assays look for mutations that are present in HIV genes (e.g.,
reverse transcriptase [RT], protease, fusion, or integrase enzymes). Some assays
sequence the entire gene, while others use probes to detect mutations that are known
to produce drug resistance.

Advantages of genotyping over phenotyping are a shorter turnaround time (1-2 weeks
compared to 2-3 weeks) and lower cost.

**Phenotypic resistance assays**

Phenotypic resistance assays measure the ability of the virus to replicate in vitro in the
presence of an ARV drug. The medical literature, the federal government and medical
society guidelines all support the use of HIV genotyping and/or phenotyping to identify
drug-resistant viruses and assist with selecting the most appropriate ARV drugs for an individual.

**Virtual phenotyping**

Virtual phenotyping is a modified genotypic test that looks for mutations in the genetic structure of the HIV virus. When mutations are found, the information is entered into a database that contains data on thousands of HIV samples. If the genotype of the virus being studied matches a genotype in the database, the assumption is that the phenotype will also match.

Virtual phenotyping is not a direct measure; rather, it is a prediction based on genotypic analysis and database matching. Current clinical practice guidelines do not provide specific guidance about the role of virtual phenotyping in HIV resistance testing. However, based on current clinical practice, it appears there is correlation between genotypic interpretation and virtual phenotypic results—particularly when the drug(s) used in HAART have been on the market long enough for the correlative database to garner enough matching genotypes and phenotypes to provide accurate phenotypic results.

As a result, the value of virtual phenotyping is limited when evaluating newer and experimental drugs because there are fewer matching genotypes and phenotypes in the correlative database.

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to HIV genotype and phenotype testing.

**Department of Health and Human Services Panel on Antiretroviral Guidelines for Adults and Adolescents**

A Department of Health and Human Services Panel on Antiretroviral Guidelines for Adults and Adolescents (DHHS, 2016) recommends:

- “HIV drug-resistance testing is recommended in persons with HIV infection at entry into care regardless of whether antiretroviral therapy (ART) will be initiated immediately or deferred.” [Evidence level AII]
  - “If therapy is deferred, repeat testing should be considered at the time of ART initiation.” [Evidence level CIII]
- “Genotypic testing is recommended as the preferred resistance testing to guide therapy in antiretroviral (ARV)-naive patients.” [Evidence level AII]
• “HIV drug-resistance testing should be performed to assist in the selection of active drugs when changing ARV regimens in persons with virologic failure and HIV RNA levels >1,000 copies/mL.” [Evidence level A1]
  o “In persons with HIV RNA levels >500 but <1,000 copies/mL, testing may be unsuccessful but should still be considered.” [Evidence level BII].
• “Drug-resistance testing should also be performed when managing suboptimal viral load reduction.” [Evidence level AII]
• “Genotypic testing is recommended as the preferred resistance testing to guide therapy in patients with suboptimal virologic responses or virologic failure while on first or second regimens.” [Evidence level AII]
• “The addition of phenotypic to genotypic testing is generally preferred for persons with known or suspected complex drug-resistance mutation patterns, particularly to protease inhibitors (PIs).” [Evidence level BIII]
• “Genotypic resistance testing is recommended for all pregnant women before initiation of ART…” [Evidence level AIII]
  o “…and for those entering pregnancy with detectable HIV RNA levels while on therapy.” [Evidence level A1]

Infectious Diseases Society of America

Infectious Diseases Society of America (IDSA, 2013) guidelines agree that HIV resistance testing should be done routinely upon initiation of care and/or at the start of ART therapy initiation as well as in patients experiencing virologic failure:

• “Because drug-resistant virus can be transmitted from one person to another, all patients should be assessed for transmitted drug resistance with an HIV genotype test upon initiation of care.” [Strong recommendation, high quality evidence].
• “If therapy is deferred, repeat testing at the time of antiretroviral therapy (ART) initiation should be considered because of the potential for superinfection.” [Weak recommendation, low quality evidence]
• “Resistance testing is also indicated for patients who are experiencing virologic failure to guide modification of ART.” [Strong recommendation, high quality evidence]

International Antiviral Society–USA Panel

International Antiviral Society–USA Panel (IAS-US, 2012) recommendations contain similar guidance on genotype testing:

• “Baseline genotypic testing for resistance should be performed in all treatment-naive patients…” [Evidence level AlA]
  o “…and in cases of confirmed virologic failure.” [Evidence level AlA]
• “A new regimen should be constructed using resistance testing (both past and present), treatment history and consideration of tolerability and adherence issues.” [Evidence level A1a]

Criteria

Medically Necessary

HIV genotyping is considered medically necessary and, therefore, covered for any of the following:

• For resistance testing at baseline for individuals with acute HIV infection, regardless of whether ARV therapy is initiated or deferred.
• To guide therapy in ARV-naive individuals with chronic HIV infection, regardless of whether ARV is initiated or deferred.
• For all pregnant women prior to initiation of therapy and for those entering pregnancy with detectable HIV RNA levels while on therapy.
• To assist in the selection of active drugs when changing ARV regimens in individuals with virologic failure or suboptimal response on their first or second regimens.
• For individuals with known or suspected complex drug-resistance patterns.

HIV phenotyping is considered medically necessary and, therefore, covered when genotypic results do not allow for drug selection in either of the following instances:

• To assist in the selection of active drugs when changing ARV regimens in individuals with virologic failure or suboptimal response on their third and subsequent regimens, or if the individual had initially acquired a multidrug-resistant virus.
• For individuals with known or suspected complex drug-resistance patterns.

Virtual phenotyping should be used with discretion as a testing option with respect to newer drugs used in antiretroviral therapy (ART) because there are typically fewer matching genotypes and phenotypes in the correlative database.

• Standard phenotyping should be used in these circumstances.

Not Medically Necessary

HIV genotyping and phenotyping performed at the same time is duplicative and is considered not medically necessary and, therefore, not covered.

References

Introduction

These references are cited in this guideline.

2. AIDSinfo. Adult and Adolescent Guidelines: Drug Resistance Testing


14. Deeks SG, Hellmann NS, Grant RM, et al. Novel four-drug salvage treatment regimens after failure of a human immunodeficiency virus type 1 protease inhibitor-


39. Panel on Treatment of HIV-Infected Pregnant Women and Prevention of Perinatal Transmission. Recommendations for use of antiretroviral drugs in pregnant HIV-1-infected women for maternal health and interventions to reduce perinatal HIV-1...


HIV Tropism Testing for Maraviroc Response

Introduction

HIV tropism testing for maraviroc response is addressed by this guideline.

Procedures addressed

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<td>HIV-1 Tropism Genotyping, Other</td>
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</table>

What is HIV tropism testing for Maraviroc response

Definition

HIV tropism testing is used to help determine an individual's response to maraviroc (Selzentry®). Maraviroc is only effective against CCR5-tropic HIV-1.

Human immunodeficiency virus (HIV)

HIV replicates itself in humans by infecting T-cells with CD4 receptors (often called CD4 cells). HIV-1 enters the CD4 cell by binding one of two cell surface co-receptors: CCR5 or CXCR4.\(^1,2\)

Tropism classifications

Tropism is the ability of HIV-1 virus to use one or both of these co-receptors. There are three main tropism classifications:\(^3\)

- **CCR5 tropism (R5-tropic)** — HIV-1 virus that only infects cells with the CCR5 co-receptor.
- **CXCR4 tropism (X4-tropic)** — HIV-1 virus that only infects cells with the CXCR4 co-receptor.
- **Dual or mixed tropism** — HIV-1 virus populations that can use either co-receptor to infect cells.

**Tropism classification changes**

CCR5-tropic virus predominates in early infection and treatment naïve patients.\(^1\)-\(^3\) CXCR4 tropism increases both as the disease progresses and with treatment.\(^1\) In later infection, CXCR4 tropism emerges in about 20% of treatment nave patients.\(^3\) Treatment experienced patients have up to a 50% chance for the presence of CXCR4-tropic virus.\(^1\)

**Treatment**

Maraviroc is an antiretroviral drug that selectively binds to the CCR5 co-receptor. This blocks CCR5-tropic HIV-1 from binding to the co-receptor and entering the cell.\(^4\)

**Contraindication**

Maraviroc is effective only against CCR5-tropic HIV-1. Patients with viruses using both the CXCR4 and CCR5 receptors (dual/mixed tropic) do not respond virologically to Maraviroc.\(^4,5\) Therefore, maraviroc is not indicated for CXCR4-tropic or dual or mixed-trophic HIV-1 infections.\(^4\)

**Clinical resistance**

Virologic failure on Maraviroc can result from outgrowth of undetected CXCR4 virus as a result of Maraviroc treatment.\(^4\)

**Test information**

**Introduction**

HIV tropism testing may include phenotype testing or genotyping assays.

**When to test**

HIV tropism testing should be performed before Maraviroc therapy is initiated. Maraviroc should only be used in adults with CCR5-tropic HIV-1 infections based on those results.\(^2,4\)

Testing may also be considered for patients with treatment failure on maraviroc. Treatment failure is often associated with a switch to CXCR4 tropism.\(^6\)

**Phenotype testing (Trofile\(^\text{®}\))**

Phenotype testing was the first method available and is most widely recommended.\(^2,7\). Phenotyping works by exposing cell lines with CCR5 or CXCR4 co-receptors to virus
made with a patient’s HIV-1 genes that control tropism. The virus’ ability to infect each cell line is assessed based on the expression of a reporter gene.\textsuperscript{2,8} The Trofile website states the assay is “100% sensitive at detecting 0.3% CXR4-using minor variant.”\textsuperscript{8} Patients enrolled in maraviroc clinical trials were screened using the Trofile phenotype assay.\textsuperscript{8,9} A newer, more sensitive version of the assay was subsequently released.\textsuperscript{2}

Genotyping assay

The genotyping assay assesses part of the HIV-1 envelope gene (the third variable loop, V3) that is the primary determinant of tropism. Quest Diagnostics’ website states that sensitivity is 5% at a viral load of 10,000 HIV-1 copies/mL.\textsuperscript{10}

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to HIV tropism testing for maraviroc response.

Department of Health and Human Services Panel

A Department of Health and Human Services Panel on Antiretroviral Guidelines for Adults and Adolescents (2016) recommends:\textsuperscript{2}

- “Coreceptor tropism assay should be performed whenever the use of a CCR5 inhibitor is being considered.” [Evidence level AI]
- “Coreceptor tropism testing is recommended in patients who exhibit virologic failure on a CCR5 inhibitor.” [Evidence level BIII]

Infectious Diseases Society of America

The Infectious Diseases Society of America (IDSA, 2013) guidelines agree that tropism testing should be done before starting any CCR5 antagonist. IDSA also states patients who exhibit virologic failure while taking a CCR5 antagonist may also be considered for tropism testing.\textsuperscript{7}

Maraviroc

Maraviroc (Selzentry ®) has been approved for use in treatment-experienced patients 16 years of age and older with only CCR5-tropic HIV-1 virus and evidence of replication despite the use of several other antiretroviral therapies.\textsuperscript{4} Regarding tropism testing, maraviroc product labeling states that:\textsuperscript{4}

- “Tropism testing must be conducted with a highly sensitive tropism assay that has demonstrated the ability to identify patients appropriate for SELZENTRY use.”
“Use of SELZENTRY is not recommended in subjects with dual/mixed or CXCR4-tropic HIV-1 as efficacy was not demonstrated in a phase 2 study of this patient group.”

Criteria
CCR5 tropism testing is considered medically necessary for the following individuals:

- Individuals with HIV-1 infection considering a CCR5 inhibitor, OR
- Individuals taking a CCR5 inhibitor who experience treatment failure

References
Introduction
These references are cited in this guideline.


HLA-B*1502 Variant Analysis for Carbamazepine Response

Procedures addressed

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<td>HLA-B*1502 Genotyping</td>
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What is HLA-B*1502

Definition

Variation in the HLA-B gene is associated with increased risk for adverse reactions to certain drugs. Testing positive for either one or two HLA-B*1502 alleles increases a person’s risk for a serious adverse skin reaction to carbamazepine.\(^1\,^2\)

- Carbamazepine(Tegretol\(®\), Tegretol XR\(®\), Equetro\(®\), Carbatrol\(®\)) is an antiepileptic agent used in the treatment of seizure disorders, psychiatric disorders, and pain from trigeminal neuralgia.

- A strong association between the risk of developing Stevens-Johnsons syndrome (SJS) and/or toxic epidermal necrolysis (TEN) with carbamazepine treatment and the presence of the inherited variant of the HLA-B gene, HLA-B*1502, has been demonstrated in studies involving patients of Chinese ancestry. For this population, the risk of having a serious reaction is 10 times higher than the risk in Caucasians for which 1 to 6 per 10,000 new users of carbamazepine have a serious reaction to the drug.\(^1\,^6\)

- Across Asian populations, notable variation exists in the prevalence of HLA-B*1502. Greater than 15% of the population is reported positive in Hong Kong, Thailand, Malaysia, and parts of the Philippines, compared to about 10% in Taiwan and 4% in North China. South Asians, including Indians, appear to have intermediate prevalence of HLA-B*1502, averaging 2 to 4%, but higher in some groups. HLA-B*1502 is present in <1% of the population in Japan and Korea. HLA-B*1502 is largely absent in individuals not of Asian origin (e.g., Caucasians, African Americans, Hispanics, and Native Americans).\(^1\)

- Testing for HLA-B*1502 should be performed prior to initiating carbamazepine treatment for most patients of Asian ancestry. Over 90% of carbamazepine treated patients who will experience SJS/TEN have this reaction within the first few months.
of treatment and providers should consider this in determining the need for screening at-risk patients who are currently on therapy.¹

- Having HLA-B*1502 is not abnormal, and there is no other known risk from having it.⁷

Test information

- HLA-B*1502 testing is performed using DNA extracted from whole blood or cheek cells. The test is positive if either one or two HLA-B*1502 alleles are detected and negative if no HLA-B*1502 alleles are detected.²⁷

Guidelines and evidence

- The Clinical Pharmacogenetic Implementation Consortium (2013) published guidelines on the use of HLA-B*1502 testing for patients prescribed carbamazepine:⁸
  - “HLA-B*1502 has a very distinct ethnic and regional distribution that is important to consider when evaluating population risk...The frequency of HLA-B*1502 is highest in Han Chinese...estimates...have been as high as 36%. In general, rates in China range from 1 to 12%. Rates in Singapore and Hong Kong have also been estimated at 10–12%. Rates in Malaysia and Thailand are estimated at 6–8%, whereas in different regions of India, the rates range from 2 to 6%. Korea and Japan have low frequencies of the allele at 0.5 and 0.1%, respectively. The allele is also quite rare in African populations (not observed) and Europeans (0–0.02%).”
  - “HLA-B*1502 is specific for SJS and TEN; there is no evidence that it predisposes to MPEs or hypersensitivity syndrome.”
  - “Much of the evidence linking HLA-B*1502 to SJS/TEN was generated in both children and adults.”
  - “Carbamazepine-induced SJS/TEN usually develops within the first 3 months of therapy; therefore, patients who have been taking carbamazepine for longer than 3 months without developing cutaneous reactions are at low risk (but not zero) of carbamazepine-induced adverse events in the future, regardless of HLA-B*1502 status.”

- A very early study has demonstrated a potential relationship between two other members of the HLA-B75 serotype commonly found in Southeast Asian populations and carbamazepine-induced SJS/TEN. There was a significant association with SJS/TEN found for Southeast Asian individuals with HLA-B*1521 and HLA-B*1511 who were prescribed carbamazepine. It was discovered that all HLA-B75 serotype molecules shared a similar capability to bind carbamazepine. More studies must be performed to further delineate this association.⁹
Product labeling for carbamazepine (Tegretol XR®) warns for the potential of developing a serious dermatological reaction from treatment with carbamazepine in HLA-B*1502 positive individuals.¹

- Carbamazepine should not be used in patients positive for HLA-B*1502 unless the benefits clearly outweigh the risks. Patients who test negative for the allele have a low risk of SJS/TEN, but should have routine monitoring for toxicity.¹
- Carbamazepine should be discontinued at the first sign of a rash, unless the rash is clearly not drug-related. If signs or symptoms suggest SJS/TEN, carbamazepine should not be resumed and alternative therapy should be considered.¹

Criteria
HLA-B*1502 variant testing is indicated in individuals with Asian ancestry prior to initiation of or during the first nine months of treatment with carbamazepine therapy.

References
HLA-B*5701 Genotyping for Abacavir Hypersensitivity

Procedures addressed

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<tr>
<td>HLA-B*5701 Genotyping</td>
<td>81381</td>
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</table>

What is HLA-B*5701

Definition

Abacavir is used in the treatment of patients with human immunodeficiency virus (HIV).

- The most important adverse effect limiting the use of abacavir is a hypersensitivity reaction (HSR) which occurs in approximately 5-8% of patients.¹
  - The abacavir HSR includes a combination of rash, fever, GI symptoms (such as nausea, vomiting, diarrhea, or abdominal cramping), constitutional symptoms (tachycardia, hypotension, myalgia, fatigue, pain, malaise, dizziness and headache) and respiratory symptoms.¹
  - Symptoms usually appear within the first six weeks of abacavir therapy, but can happen at any time.¹-³

- People with a positive HLA-B*5701 test are at risk for abacavir HSR. Not all HLA-B*5701 carriers will have immunologic-confirmed HSR.² In studies of people who have experienced an immunologically-confirmed HSR, about half (47.9%) test positive for the HLA-B*5701 allele.¹

- People with a negative HLA-B*5701 are at low risk for abacavir HSR. A negative HLA-B*5701 test result does not completely rule out the possibility of an HSR. Those who test negative should be monitored carefully for signs of toxicity, especially in the first six weeks of treatment.⁴

- Demographic risk factors for abacavir HSR show a higher risk in white and Hispanic populations (5-8%) compared to 2-3% in the black population.⁴,⁵ The frequency in Asian populations is very low.²
• Screening HIV-1 patients for HLA-B*5701 prior to starting abacavir can reduce the rate of clinically suspected HSR by approximately 60%.1

**Test information**

• HLA-B*5701 testing is performed on a blood or cheek swab sample. The test can be performed in different ways by different labs. Some labs will test for specific gene variants associated with the B*5701 haplotype, where other labs may sequence the DNA in the HLA-B region.

• In general, results can be interpreted as:

  o HLA-B*5701 positive – person is at high risk for developing abacavir HSR; abacavir-containing drugs should be avoided.

  o HLA-B*5701 negative – person is at lower risk for developing abacavir HSR; if abacavir treatment is used, this person should be monitored for toxicity.

**Guidelines and evidence**

• The Infectious Disease Society of America (2013)6 and the Department of Health and Human Services’ (DHHS) Panel on Antiretroviral Guidelines for Adults and Adolescents (2016)4 HIV guidelines recommend that:

  o HLA-B*5701 genotyping should be performed in all patients prior to initiating abacavir therapy.

  o HLA-B*5701 positive patients should not be prescribed abacavir; however, the guidelines state that if abacavir is used in HLA-B*5701 positive patients, careful monitoring for HSR is warranted.

  o A negative test result does not rule out the possibility of an HSR but makes the chance of HSR less likely.

  o Patients should be counseled about the potential for experiencing HSR before being treated with abacavir-containing drugs, regardless of HLA-B*5701 test results.

  o HLA-B*5701 positive status should be recorded as an abacavir allergy in the patient’s medical record.

• The DHHS’s Panel on Antiretroviral Therapy and Medical Management of HIV-Infected Children (2016) recommends against the use of abacavir in children who test positive for HLA-B*5701.7

• The Clinical Pharmacogenetics Implementation Consortium (2014) published an update to their Guidelines on HLA-B Genotype and Abacavir Dosing.8 A focused literature review found no new evidence to change their original (2012) recommendations, which include:

  o “HLA-B*5701 screening should be performed in all abacavir-naive individuals before initiation of abacavir-containing therapy.”
o “In abacavir-naive individuals who are HLA-B*5701-positive, abacavir is not recommended and should be considered only under exceptional circumstances when the potential benefit, based on resistance patterns and treatment history, outweighs the risk.”

o “There is some debate among clinicians regarding whether HLA-B*5701 testing is necessary in patients who had previously tolerated abacavir chronically, discontinued its use for reasons other than HSR, and are now planning to resume abacavir. The presence of HLA-B*5701 has a positive predictive value of ~50% for immunologically confirmed hypersensitivity, indicating that some HLA-B*5701-positive individuals can be, and have been, safely treated with abacavir. However, we were unable to find any data to show that HLA-B*5701-positive individuals with previous, safe exposure to abacavir had zero risk of HSR upon re-exposure.”

- Product labeling for abacavir-containing drugs recommends:
  - HLA-B*5701 testing prior to initiating treatment with abacavir and prior to reinitiating abacavir when HLA-B*5701 status is unknown even if the patient has previously tolerated treatment with abacavir.
  - For HLA-B*5701-positive patients, treatment with an abacavir-containing regimen is not recommended and should be considered only with close medical supervision and under exceptional circumstances when the potential benefit outweighs the risk.
  - Abacavir is contraindicated in patients with previous hypersensitivity to abacavir.
  - Discontinue abacavir at the first sign of a suspected hypersensitivity reaction.

- Careful monitoring for adverse effects is recommended during the first six weeks of abacavir therapy, when an HSR is most likely to happen. However, an HSR can occur at any time during treatment with abacavir.

Criteria
HLA-B*5701 testing is indicated in individuals with HIV-1 prior to the initiation of any abacavir-containing therapy.

References
3. Information for Healthcare Professionals: Abacavir (marketed as Ziagen) and Abacavir-Containing Medications. [Internet] Silver Spring (MD): US Food and Drug


Huntington Disease

Procedures addressed

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<thead>
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<tr>
<td>HTT Expansion Analysis</td>
<td>81401</td>
</tr>
</tbody>
</table>

What is Huntington disease

Definition

Huntington disease (HD) is an autosomal dominant neurodegenerative disorder causing progressive cognitive, motor, and psychiatric disturbances.\(^1\)

Prevalence

The prevalence of HD ranges from 3 to 7 per 100,000 people in populations of western European descent.\(^1\)

It is less frequent in individuals of Chinese, Japanese, Finnish or African descent. The prevalence of HD is believed to be highest in individuals living in the Lake Maracaibo region of Venezuela.\(^1\)

Cause

HD is caused by expansion of a CAG trinucleotide repeat mutation in the HTT gene. The number of CAG repeats is typically associated with the severity of disease.

<table>
<thead>
<tr>
<th>When a person has this number of CAG repeats ...</th>
<th>Then the person ...</th>
</tr>
</thead>
<tbody>
<tr>
<td>26 or fewer</td>
<td>is unaffected.</td>
</tr>
<tr>
<td>27 to 35</td>
<td>is in the intermediate range and is typically not affected with HD. However, any offspring are at risk for HD, because the repeat number can expand over generations.(^1)</td>
</tr>
<tr>
<td>36 to 39</td>
<td>is at risk for HD but may not develop symptoms.(^1)</td>
</tr>
<tr>
<td>40 or more</td>
<td>will develop HD symptoms.(^1)</td>
</tr>
</tbody>
</table>
Onset

The mean age of onset of symptoms is 35-44 years of age.\(^1\) Approximately 3-10% of individuals with HD have onset of symptoms before 21 years of age.\(^2\) This is known as juvenile HD. Juvenile HD most commonly results from paternally inherited HD mutations with larger CAG repeats.

Survival

Median survival time is 15-18 years after onset.\(^1\)

Inheritance

HD is an autosomal dominant condition. Because the CAG repeat number can expand over generations, children of individuals with HD are at increased risk. Typically, as the disease passes through generations

- severity of HD symptoms increases, and
- age of onset decreases.

This is seen more often when inherited through a male. This phenomenon is known as anticipation.\(^1\)

Treatment

There is no cure for HD. Some pharmacologic treatments may be effective in decreasing some of the associated symptoms, such as chorea, rigidity and psychiatric disturbances.\(^1\)

Test information

Introduction

Testing for Huntington disease includes analysis to determine the number of CAG repeats.

CAG repeat testing

Testing for Huntington disease is performed by determining the number of CAG repeats in the HTT gene.\(^1\) CAG repeat analysis has a mutation detection rate greater than 99%.\(^1\)

Diagnostic testing

Symptomatic HD testing is appropriate for individuals who have a known or suspected diagnosis of HD based on clinical symptoms.\(^3\)
Predictive testing

Predictive HD testing is appropriate for adults who have a known family history of HD, and wish to know their HD mutation status. Predictive testing should be performed in the context of thorough counseling (described below in Guidelines/Evidence). Predictive HD testing is generally not recommended for minors. Predictive testing for HD cannot accurately predict disease severity, type of symptoms, or rate of progression in asymptomatic individuals. However, an estimate of age of onset is possible based on the number of CAG repeats detected.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to Huntington disease testing.

United States Huntington's Disease Genetic Testing Group

The United States Huntington's Disease Genetic Testing Group (2003) has guidelines regarding genetic testing for Huntington disease.

Symptomatic testing

“Confirmatory testing by analysis of the HD gene may be offered at or after the time of the clinical diagnosis of HD. The presence of a CAG repeat expansion in a person with HD symptoms confirms the clinical impression and supports a diagnosis of HD.”

Predictive testing

Asymptomatic (predictive) testing is supported in the context of a predictive testing protocol that includes

- optional neurological exam
- psychological exam,
- social support
- pre- and post-test counseling regarding implications of positive and negative test results, and
- documented informed consent.

Predictive testing protocol support

The predictive testing protocol is also supported by guidelines from
• the International Huntington Association and the World Federation of Neurology Research Group on Huntington’s Chorea (1994)⁴
• the American College of Medical Genetics and Genomics⁵, and
• the National Society of Genetic Counselors.⁶

Criteria

Introduction

Requests for Huntington disease testing are reviewed using these criteria.

Criteria

• Clinical Consultation:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), and
  o Examination by a geneticist or physician familiar with genetic movement disorders, AND

• Previous Genetic Testing:
  o No previous genetic testing of HTT, AND

• Diagnostic Testing for Symptomatic Individuals:
  o For individuals 18 years of age or older:(at least 2 of the following)
    ▪ Progressive motor disability featuring involuntary movements (chorea) and gait disturbance, and/or
    ▪ Behavioral disturbances including:
      • Personality change
      • Depression
      • Cognitive decline, and/or
    ▪ Family history of Huntington disease
  o For individuals 17 years of age or younger: (at least 2 of the following)
    ▪ Progressive motor disability featuring involuntary movements (chorea) and gait disturbance, and/or
    ▪ Cognitive decline, and/or
    ▪ Stiffness or rigidity, and/or
    ▪ Epilepsy/myoclonus and tremor, and/or
Family history of Huntington disease, OR

Predictive Testing for Presymptomatic/Asymptomatic At-Risk Individuals:

- For individuals 18 years of age or older:
  - Known CAG trinucleotide repeat expansion in HTT in 1st, 2nd, or 3rd degree biologic relative, or
  - One or more 1st degree biologic relative(s) with clinical diagnosis of HD and mutation unknown/not yet tested

References

Introduction

These references are cited in this guideline.


Hypertrophic Cardiomyopathy Testing

Procedures addressed

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<tr>
<td>Hypertrophic Cardiomyopathy Genetic Testing Panel</td>
<td>S3865</td>
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<tr>
<td>Hypertrophic Cardiomyopathy Known Familial Mutation Analysis</td>
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<td>MYH7 Sequencing</td>
<td>81407</td>
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<tr>
<td>MYL2 Sequencing</td>
<td>81405</td>
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</tbody>
</table>
What is hypertrophic cardiomyopathy

Definition

Hypertrophic cardiomyopathy (HCM) is a genetic condition associated with unexplained thickening of the heart wall surrounding the left ventricle (called left ventricular hypertrophy or LVH).

- A clinical diagnosis is suggested by a nondilated left ventricle with a wall thickness of 13-15mm or more in adults, or ≥2 standard deviations in children. However, some individuals with HCM have smaller LV measurements and variable patterns of LVH may be observed.
- Other causes of LVH should be ruled out, including underlying cardiac disease (e.g., chronic hypertension, aortic stenosis), extreme physiologic hypertrophy (“athlete's heart”), and other multisystem disorders that may have LVH as a feature (e.g., Fabry disease, Friedreich's ataxia, Noonan syndrome, LEOPARD syndrome, Danon disease, Barth syndrome, Pompe syndrome).
- Signs and symptoms are variable ranging from a lifelong asymptomatic course to progressive heart failure and sudden cardiac death.
- HCM affects about 1 in 500 people, and is the most common cause of sudden cardiac death among young people under 35 - especially athletes.
• HCM is an autosomal dominant condition. First-degree relatives (parents/siblings/children) of someone with HCM have up to a 50% chance of also being affected. Longitudinal clinical screening is recommended for at-risk relatives.²,⁵,⁷

• HCM is caused by a mutation in one of at least 14 genes.² Genetic testing can be useful to confirm a diagnosis of inherited HCM in a person with unexplained LVH. A family history of LVH, heart failure, or sudden cardiac death supports the diagnosis of HCM but is not required to make a diagnosis. The severity and likelihood of cardiac death may be associated with the gene mutation that causes HCM.⁴

• Identifying a gene mutation does not significantly change management for someone diagnosed with HCM.⁶ However, once the disease-causing mutation is identified, at-risk relatives can have reliable genetic testing to define their risk and screening needs.⁷

### Test information

• HCM Sequencing Panels vary by laboratory but most laboratories test at least the eight genes most commonly linked to HCM. Mutations in the MYH7 and MYBC3 genes are most common.¹ About 35-60% of people clinically diagnosed with HCM will have a mutation in one of the genes commonly tested.¹,⁵

• Once a mutation is identified in a family member, the family mutation can be specifically identified with >99% accuracy in asymptomatic family members, or those with equivocal symptoms.²

### Guidelines and evidence

#### Diagnostic testing

• Evidence-based guidelines from the European Society of Cardiology published in 2014⁸ state:
  
  o “Genetic testing is recommended in patients fulfilling diagnostic criteria for HCM, when it enables cascade genetic screening of their relatives.” (Class 1, Level B)
  
  o “It is recommended that genetic testing be performed in certified diagnostic laboratories with expertise in the interpretation of cardiomyopathy-related mutations.” (Class 1, Level C)
  
  o “In the presence of symptoms and signs of disease suggestive of specific causes of HCM, genetic testing is recommended to confirm the diagnosis.” (Class 1, Level B)
  
  o “Genetic testing in patients with a borderline diagnosis of HCM should be performed only after detailed assessment by specialist teams.” (Class IIa, Level C)
• “Post-mortem genetic analysis of stored tissue or DNA should be considered in deceased patients with pathologically confirmed HCM, to enable cascade genetic screening of their relatives.” (Class IIa, Level C)

• The Cardiac Society of Australia and New Zealand (2013) made the following recommendation regarding the use of diagnostic testing for HCM:⁹
  o “Genetic testing may also help to discriminate between HCM and other causes of left ventricular hypertrophy, including hypertension and “athlete’s heart”.”

• A 2011 expert consensus statement from the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA)¹⁰ makes Class 1 recommendation that:
  o “Comprehensive or targeted (MYBPC3, MYH7, TNNI3, TNNT2, TPM1) HCM genetic testing is recommended for any patient in whom a cardiologist has established a clinical diagnosis of HCM based on examination of the patient’s clinical history, family history, and electrocardiographic/echocardiographic phenotype.”

• Evidence-based guidelines from the American College of Cardiology Foundation (ACCF) and the American Heart Association (AHA) published in 2011 state:
  o “Genetic testing for HCM and other genetic causes of unexplained cardiac hypertrophy is recommended in patients with an atypical clinical presentation of HCM or when another genetic condition is suspected to the cause.” (Class 1, Level of evidence B).⁵
  o “Genetic testing is reasonable in the index patient to facilitate the identification of first-degree family members at risk for developing HCM.” (Class IIa, Level of Evidence B).⁵

• Evidence-based practice guidelines for the genetic evaluation of cardiomyopathies, including HCM, from the Heart Failure Society of America (HFSA, 2009) state:
  o “Genetic testing should be considered for the one most clearly affected person in a family to facilitate family screening and management.” (evidence level A: The specific genetic test or clinical test has a high correlation with the cardiomyopathic disease of interest in reasonably large studies from multiple centers.)⁶

Predictive testing

• Evidence-based guidelines from the European Society of Cardiology published in 2014 state:⁸
  o “It is recommended that genetic testing be performed in certified diagnostic laboratories with expertise in the interpretation of cardiomyopathy-related mutations.” (Class 1, Level C)
o “Cascade genetic screening, after pre-test counseling, is recommended in first-degree adult relatives of patients with a definite disease-causing mutation.” (Class I, Level B)

o “Clinical evaluation, employing ECG and echocardiography and long-term follow-up, is recommended in first-degree relatives who have the same definite disease-causing mutation as the proband.” (Class 1, Level C)

o “First-degree relatives who do not have the same definite disease-causing mutation as the proband should be discharged from further follow-up but advised to seek re-assessment if they develop symptoms or when new clinically relevant data emerge in the family.” (Class IIa, Level B)

- The Cardiac Society of Australia and New Zealand (2013) made the following recommendation regarding the use of predictive testing for HCM:

  o “Identifying the disease-causing gene mutation can be very valuable for a family, as it can allow earlier management of at-risk members and avoid unnecessary screening of non-carriers.”

- A 2011 expert consensus statement from the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA) makes Class 1 recommendation that:

  o “Mutation-specific genetic testing is recommended for family members and appropriate relatives following the identification of the HCM-causative mutation in an index case.”

- Evidence-based guidelines from the American College of Cardiology Foundation (ACCF) and the American Heart Association (AHA) published in 2011 make the following Class I recommendations:

  o “Screening (clinical, with or without genetic testing) is recommended in first-degree relatives of patients with HCM.” (Level of Evidence: B)

  o “In individuals with pathogenic mutations who do not express the HCM phenotype, it is recommended to perform serial electrocardiogram (ECG), transthoracic echocardiogram (TTE), and clinical assessment at periodic intervals (12 to 18 months in children and adolescents and about every 5 years in adults), based on the patient’s age and change in clinical status.” (Level of Evidence: B)

### Criteria

#### Known Familial Mutation(s) for Hypertrophic Cardiomyopathy

- Genetic Counseling:
• Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous HCM-associated genetic testing inclusive of known family mutation, AND

• Diagnostic/Predisposition Testing for Presymptomatic/Asymptomatic Individuals:**
  o HCM known family mutation in 1st or 2nd degree biologic relative, OR

• Diagnostic Testing for Symptomatic Individuals:
  o HCM known family mutation in 1st or 2nd degree biologic relative
  o Echocardiogram demonstrating LVH without obvious cause (valvular disease, hypertension, infiltrative or neuromuscular disorder), and
  o Myocardial wall thickening of greater than or equal to 15mm (1.5cm), or
  o Presence of pathognomonic histopathologic features of HCM
    ▪ Myocyte disarray
    ▪ Hypertrophy
    ▪ Increased myocardial fibrosis, and
  o The results of the test will directly impact the diagnostic and treatment options that are recommended for the patient, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**NOTE:** Since symptoms may occur in childhood, testing of children who are at-risk for a pathogenic mutation may be appropriate, but requires genetic counseling and careful consideration of ethical issues related to genetic testing in minors.³

Hypertrophic Cardiomyopathy Genetic Testing Panel

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous genetic testing for HCM, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Echocardiogram demonstrating LVH without obvious cause (valvular disease, hypertension, infiltrative or neuromuscular disorder), and
  o Myocardial wall thickening of greater than or equal to 15mm (1.5cm), or
Presence of pathognomonic histopathologic features of HCM

- Myocyte disarray
- Hypertrophy
- Increased myocardial fibrosis, and

The results of the test will directly impact the diagnostic and treatment options that are recommended for the patient, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

Billing and reimbursement considerations

- When multiple CPT codes are billed for components of a panel and there is a more appropriate CPT code representing the panel, eviCore will redirect to the panel code(s).
  - If the panel includes required genes for CPT code 81439 (DSG2, MYBPC3, MYH7, PKP2, TTN), the panel will be approved for 81439, with the billed amount not to exceed the list price of the test, or
  - If the panel does not include all of the required genes for CPT code 81439, the panel will be approved for 81479 with the billed amount not to exceed the list price of the test.

- If the laboratory will not accept redirection to a panel code, the medical necessity of each billed component procedure will be assessed independently.
  - In general, only a limited number of panel components that are most likely to explain the member's presentation will be reimbursable. The remaining panel components will not be reimbursable.
  - When the test is billed with multiple stacked codes, only the following genes may be considered for reimbursement:
    - MYH7
    - MYBPC3
    - TNNT2
    - TNNI3
TPM1

References


10. Ackerman MJ, Priori SG, Willems S, et al. HRS/EHRA expert consensus statement on the state of genetic testing for the channelopathies and cardiomyopathies: this document was developed as a partnership between the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA). Europace. 2011;13(8):1077-1109. Available at: http://europace.oxfordjournals.org/content/europace/13/8/1077.full.pdf
Immunohistochemistry (IHC)

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Immunohistochemistry or immunocytochemistry, per specimen; each additional single antibody stain procedure (List separately in addition to code for primary procedure)</td>
<td>88341</td>
</tr>
<tr>
<td>Immunohistochemistry or immunocytochemistry, per specimen; initial single antibody stain procedure</td>
<td>88342</td>
</tr>
<tr>
<td>Immunohistochemistry or immunocytochemistry, per specimen; each multiplex antibody stain procedure</td>
<td>88344</td>
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</tbody>
</table>

What is immunohistochemistry

Definition

Immunohistochemistry (IHC) is a method used to determine the expression of biomarkers in tissue. Antibodies that detect specific antigens (proteins, biomarkers) are applied to tissue and attach to their target antigen. The antibodies are tagged with a visible label that allows the pattern or distribution of the antigen in the tissue to be directly visualized under a microscope.

Test information

Immunohistochemistry is used widely in pathology for diagnosis, sub-typing, and increasingly to identify therapeutic targets. It has also dramatically changed the approach to evaluating cancer of unknown primary. There are numerous other applications and some of the most common are outlined below:¹⁻³

- Diagnostic
  - Initial detection of malignancy, tumor classification (including cancers of unknown origin), and sub-typing
- Identification of infections (e.g., H. pylori induced gastritis or CMV infection)
- Neurodegenerative disorder classification

**Therapy Selection/Management**
- Identification of specific therapeutic targets (e.g., HER2 IHC to determine response to trastuzumab)
- Monitoring therapeutic response
- Further characterization of prognosis to gauge treatment aggressiveness

**Genetic Disorder Evaluation**
- Altered gene expression predictive of an underlying genetic disorder (e.g., loss of expression of the mismatch repair genes associated with Lynch syndrome)
- Skeletal muscle biopsy protein abnormalities that help establish a specific muscular dystrophy diagnosis

**Criteria**

This guideline addresses common issues in billing for immunohistochemistry-based tests using CPT codes 88341-88344. It is not intended to encompass immunohistochemistry-based tests billed using more specific CPT codes (e.g., 88360, 88361).

**Medical necessity requirements**

Given the numerous applications of IHC, it is not practical or desirable to create policy to address the specific use of stains in various settings. That said, it is possible to determine a maximum number of stains that should be reasonable for the vast majority of applications.

Several studies have included data on the average number of immunohistochemical (IHC) stains used per case in various settings. In a cost-effectiveness of IHC study, Raab (2000) modeled the analysis on a 5-antibody panel, which was the average number of antibodies ordered per case in that hospital system. No data was provided on the upper and lower limits of that range.¹

A study published by Shah, et al. (2012) looked at the use of IHC stains among different pathology practice settings, which included academic, private and commercial practices. The study concluded that regardless of where IHC was performed, the average number of stains ordered per case was similar among all groups although ranges varied considerably. Pathologists from private groups performed an average of 4 stains (range 1-13), whereas those in commercial laboratories used an average of 3 stains per case (range 1-7 for both). When broken down by organ system, the highest average was 6 stains per case for head and neck tissue.⁶
The National Comprehensive Cancer Network (NCCN) Guidelines for Treatment of Cancer by Site provide detailed guidelines on the use of individual IHC stains in the diagnosis and management of each cancer type addressed.\footnote{7}

National Comprehensive Cancer Network (NCCN, 2016) Guidelines on Occult Primary (Cancer of Unknown Primary [CUP]) state the following: “In patients with occult primary tumors, immunohistochemical studies are useful for the characterization of poorly differentiated or undifferentiated tumors and for cell-type determination and pathologic diagnosis. However, exhaustive IHC studies (in excess of 10-12 stains) have not been shown to increase the diagnostic accuracy in identifying the putative primary sites” NCCN recommends a tiered approach as follows: first tier determines tissue lineage, second tier can suggest putative primary sites, and an optional third tier to identify therapeutic targets in select patients.\footnote{8}

Based on these findings, most immunohistochemistry applications should rarely require more than 12 IHC stains per unique specimen. Therefore the following criteria will be applied when processing claims for IHC procedures:

- In addition to the first stain billed with one unit of 88342, reimbursement will routinely be limited to 11 units of CPT 88341 per specimen.
- Any claim for IHC, regardless of the number of units billed, may be subject to post-service medical necessity review if excess units are suggested based on the available clinical information.

**Billing and reimbursement considerations**

There are currently three codes for reporting qualitative IHC stains: 88341, 88342, and 88344. IHC stains are now reported per unique specimen instead of per block (paraffin-embedded tissue). Examples of unique specimens that may be evaluated on the same date of service are separate colon polyps or skin biopsies from different lesions.

- **Current codes:**
  - Codes 88342 and 88341 are reported for a single antibody stain procedure.
  - Code 88344 is used to report a multiplex staining procedure* (e.g., PIN-4, ADH-5, Uro-3 triple stain).
- **Retired codes:**
  - Code 88343 was deleted in 2015.
  - The HCPCS codes G0461 and G0462 are also no longer reportable.

*Multiplex staining refers to the use of two or more different antibodies mixed together (“cocktails”) that demonstrate different staining characteristics on a single slide. Multiplex does not refer to antibody cocktails such as Cytokeratin AE1/AE3 that do not show distinct color changes between antibodies. There are a limited, although expanding, number of multiplex stains with PIN4 being among the most frequently utilized (evaluation of prostate biopsies).\footnote{4}
Therefore, the following limitations will be applied when processing claims for IHC procedures:

- Qualitative immunohistochemistry procedure codes 88341-88344 should only be used when other, more specific, procedure codes are not available to describe the performed test, AND

- For single antibody stains:
  - One unit of 88342 should be used for the first single antibody applied to a unique specimen. Additional stains applied to that same specimen are billed with one unit of 88341 per stain to an allowable maximum (see Allowable Units policy below). 88341 should therefore not be billed without 88342 on the same date of service, and
  - It is generally unnecessary to test more than three separate specimens on the same date of service. Therefore, no more than three units of 88342 are routinely reimbursable on a single date of service, and
  - When more than one specimen is studied, the units of 88341 applicable to each unique specimen should be entered separately on the claim and each entry should have a corresponding unit of 88342 billed, OR

- For multiplex antibody stains:
  - One unit of 88344 may be used for a multiplex stain applied to a unique specimen. A multiplex stain is defined as a combination of antibodies that yield separately identifiable staining characteristics on a single slide, and
  - No more than one multiplex stain on one specimen should be necessary on a single date of service, AND

- Modifiers 26 and TC can be used to split codes 88341, 88342, and 88344 into their technical and professional components. When split, one unit of the technical component of a code and one unit of the professional component of a code will be viewed as the equivalent of one unit when calculating maximum allowable units for any code. Alternatively, the sum of units billed with the same modifier (e.g. TC) can fulfill the maximum allowable units regardless of whether the units for the other modifier (e.g., 26) are ever billed.

**Note** *Multiplex staining refers to the use of two or more different antibodies mixed together (“cocktails”) that demonstrate different staining characteristics on a single slide. Multiplex does not refer to antibody cocktails such as Cytokeratin AE1/AE3 that do not show distinct color changes between antibodies. There are a limited, although expanding, number of multiplex stains with PIN4 being among the most frequently utilized (evaluation of prostate biopsies).*
References


KRAS Testing for Anti-EGFR Response in Metastatic Colorectal Cancer

Procedures addressed

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<table>
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<tr>
<th>Procedures addressed by this guideline</th>
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<td>KRAS Exon 2 Targeted Mutation Analysis</td>
<td>81275</td>
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<tr>
<td>KRAS Targeted Mutation Analysis, Additional Variants</td>
<td>81276</td>
</tr>
<tr>
<td>KRAS Sequencing</td>
<td>81405</td>
</tr>
</tbody>
</table>

What is KRAS mutation analysis

Definition

KRAS mutation analysis on metastatic colorectal cancer (mCRC) tissue helps identify patients who are most likely to respond to EGFR-targeted therapy (Erbitux® and Vectibix®).1-4

- EGFR-targeted therapies usually bind EGFR, block its signaling to KRAS, and inhibit cellular proliferation, angiogenesis, and metastasis.3
- Approximately 40% of mCRC tumors have an activating KRAS mutation.3
- Anti-EGFR therapy is ineffective for treating mCRC tumors with an activating KRAS mutation because EGFR no longer controls KRAS activation.
- Thus, testing identifies the subset of patients who are resistant to anti-EGFR treatment, avoiding unnecessary drug toxicity and cost.3,5,6 In addition, some patients with KRAS mutant tumors were found to have an inferior outcome when treated with EGFR-targeted therapy.3,8

Test information

- **KRAS Targeted Mutation Analysis** identifies specific KRAS gene mutations — usually including at least the seven most common mutations in codons 12 and 13 that account for more than 95% of activating mutations.3,6 It requires very little tumor material for testing, and combines high sensitivity with efficiency. It is also relatively inexpensive and is designed to detect the most common mutations within the KRAS
gene. Because it does not evaluate the whole KRAS gene, it will miss the less common mutations. KRAS mutation analysis uses fresh, frozen, or paraffin-embedded tissue from either a primary tumor or metastasis.\textsuperscript{3,7} 

- **KRAS Gene Sequencing Analysis** identifies most clinically significant mutations in the KRAS gene, including both common and rare changes. It has the broadest coverage in KRAS testing, looking at most, if not all, coding areas within the gene. However, sequence analysis requires more and higher quality tumor material for testing than PCR. This typically translates into being less efficient and more expensive than targeted mutation analysis. Direct sequence analysis has lower analytical sensitivity than some targeted, PCR based assays. However, the clinical relevance of a small percentage of cells with mutant KRAS has not been established.

**Guidelines and evidence**

- Consensus from the National Comprehensive Cancer Network (NCCN, 2017) “strongly recommends KRAS/NRAS genotyping of tumor tissue (either primary tumor or metastasis) in all patients with metastatic colorectal cancer.” “Patients with known KRAS or NRAS mutations should not be treated with either cetuximab or panitumumab, either alone or in combination with other anticancer agents, because they have virtually no chance of benefit and the exposure to toxicity and expense cannot be justified.”\textsuperscript{2}

- Evidence based guidelines from the American Society of Clinical Oncology (ASCO, 2015) state: “All patients with mCRC who are candidates for anti-EGFR antibody therapy should have their tumor tested in a Clinical Laboratory Improvement Amendments–certified laboratory for mutations in both KRAS and NRAS exons 2 (codons 12 and 13), 3 (codons 59 and 61), and 4 (codons 117 and 146). The weight of current evidence indicates that anti-EGFR MoAb therapy should only be considered for treatment of patients whose tumor is determined to not have mutations detected after such extended RAS testing.”\textsuperscript{1}

- These guidelines do not recommend a specific test methodology.

**Criteria**

KRAS mutation testing is indicated in individuals with metastatic colorectal cancer prior to the initiation of treatment with cetuximab (Erbitux\textsuperscript{®}) or panitumumab (Vectibix\textsuperscript{®}) therapy.

**References**


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www.eviCore.com


## Procedures addressed

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<td>81490 - 81599; 0001M - 0009M</td>
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<tr>
<td>Proprietary Laboratory Analyses (PLA)</td>
<td>Molecular tests* within range (currently 0001U-0034U)</td>
</tr>
<tr>
<td>Molecular Infectious Testing</td>
<td>Molecular tests* within range 87149 – 87912 G0476</td>
</tr>
<tr>
<td>Molecular Cytopathology Procedures (Flow Cytometry, In Situ Hybridization)</td>
<td>88120 - 88121 88182 - 88199</td>
</tr>
<tr>
<td>Cytogenetics</td>
<td>88230 - 88299</td>
</tr>
<tr>
<td>Molecular Surgical Pathology Procedures (Immunohistochemistry, In Situ Hybridization)</td>
<td>88341 - 88344 88360 - 88361 88364 –88377 88380 - 88388</td>
</tr>
<tr>
<td>Other Molecular Codes</td>
<td>84999 86152 86153</td>
</tr>
<tr>
<td>Molecular HCPCS Codes</td>
<td>S3800 - S3870 G0452, G0464, G9143</td>
</tr>
</tbody>
</table>

**Note**  
*Generally defined as codes that include “DNA”, “RNA”, “nucleic acid”, “genotype”, “phenotype” or related language in the code description.*
Description

eviCore manages claims payment for our subscribing Health Plans. Procedure codes (CPT, HCPCS) are adjudicated against claims review and payment rules. Additional payment and coverage adjustments may be made in addition to those outlined in this policy.

The following claim reimbursement policies provide general guidance on what forms of review may be employed. They are intended to augment other clinical and administrative policies and do not represent all possible claims treatments.

Criteria: Claims Reimbursement

Introduction

All procedure codes included in the Molecular and Genomic Testing Laboratory Program (as outlined in the table at the top of this document) may be subject to claims review and payment policies. The following policies define many, but not all, of the most commonly applied claims edits.

Authorization Check

Required Authorization

Procedure codes that require medical necessity authorization are defined in published Plan procedure code management lists, which are generally available on the Health Plan’s website. All claims will be reviewed for the presence of any procedure code that requires authorization. When required, the following process is employed:

- The procedure code(s) requiring authorization will be checked against an authorization database.
- The procedure code will be released for further adjudication if an authorization is on file for all units of that procedure code and the code is billed with any stipulated additional information (e.g., modifier, unique test identifier).
- If any of the following are true, the Plan’s authorization requirements will be enforced. This may include denial for failure to comply with prior authorization requirement or post-service medical necessity review.
  - An authorization is not on file for all units of that CPT/HCPCS, or
  - The stipulated modifier is not appended to the code, or
• The authorization is not valid for the date of service.

Voluntary Prior Authorization

Providers may choose to seek prior authorization for one or more procedure codes that are in scope for the Molecular and Genomic Lab Management Program but do not routinely require pre-service review. This is termed voluntary prior authorization. A voluntary prior authorization will also be performed for all codes that do not require prior authorization when they are being billed as part of a test panel with any codes that do require authorization.

- If a voluntary authorization is on file for any procedure code, and the procedure code is billed with all stipulated information, the procedure will be released for further adjudication.
- If an authorization is not on file for all units of that procedure code, the stipulated information is not included on the claim, or the date of service is not valid, all or some of the units for that procedure may undergo any post-service medical necessity review processes in place for that procedure but will not be denied for failure to obtain authorization.

Substitutable Codes

Note that an authorization for a procedure code may be used to approve coverage for a DIFFERENT billed procedure code that is substantially similar in clinical intent to the authorized code (e.g., an authorization to perform CPT 81228 is substitutable if CPT 81229 is billed). Clinically reasonable substitution rules are automatically applied through the Claims Studio process. When substitution rules are invoked, the billed procedure code is the paid procedure code. See the companion Procedure Code Substitution table for additional details.

Supporting Documents

For more information, see the Supporting Documents section at the end of this guideline.

Post-Service Medical Necessity Determination

Many lab tests that are in scope for the Molecular and Genomic Lab Management Program are not managed through mandatory authorization requirements. Appropriate billing or medical necessity may be assessed upon claim submission (post-service) prior to payment as follows:

• All procedure codes managed under this program may be subject to post-service medical necessity review.
• Any and all available claims data (e.g., ICD code, age, gender, historical or co-existing procedures, etc.) may be used to determine medical necessity or identify cases requiring further review.
- Claims data may be sufficient to determine medical necessity without additional clinical information. When medical necessity is determined based on claims data alone, the claims information that will either support or refute medical necessity is defined in the clinical policy (e.g., submitted ICD codes do not support medical necessity for a procedure).

- When a case is identified for additional post-service medical necessity review, communication is sent to at least the rendering provider requesting additional information with the following possible outcomes:
  - If the required clinical information is provided and fulfills criteria, the procedure is approved and the claim is released for further adjudication.
  - If the required clinical information is provided and does not fulfill criteria, the procedure is denied for lack of medical necessity.
  - If the required clinical information is not provided within the specified timeframe, the procedure is denied for failure to comply with the post-service review process.

- The factors that may prompt post-service medical necessity review include, but are not limited to:
  - ICD codes that support clinical criteria are not reported on the claim.
  - A billed amount threshold is exceeded.
  - A particular procedure code is billed with other procedure codes (bundled testing whether defined by the laboratory as a panel or not).
  - The claim is submitted by a provider (participating and non-participating) selected for focused review.
  - Billing portrait demonstrates billing patterns selected for focused review.

- There are multiple sources of the rules established by this policy including CMS documents, published code definitions, specialty guidelines, peer reviewed literature, expert opinion, and claims experience with codes or providers.

**Gender Nondiscrimination**

Gender reported on a claim is one element used to determine medical necessity. In situations where the reported gender may not be consistent with the medical needs based on biological sex (e.g., transgender, transsexual, intersex individuals), the KX modifier should be appended to each billed procedure code that may have gender-related policy. The KX modifier will allow automated gender-specific edits to be bypassed.
**Lifetime Maximums**

In general, the same or similar tests performed on heritable DNA should not need to be performed more than once on the same person in that person's lifetime (e.g., gene sequencing or a similar mutation panel on a gene should not need to be repeated). Rarely, a procedure code may be billed twice for the same female member when subsequent instances represent testing on the female member’s fetus. It is the provider’s responsibility to determine if any contemplated genetic testing has already been performed for the member and to avoid unnecessary repeat testing.

Lifetime maximum rules will be applied for procedure codes that involve genetic testing of heritable DNA in the following manner:

- See the companion *Lifetime Maximums* table for a list of procedure codes subject to lifetime maximum policy.
  - Only a single date of service will be reimbursed for any procedure code with a lifetime maximum for a single individual.
  - While most procedure codes have a lifetime maximum of one unit, some have a limit of 2 (e.g. known familial mutations for recessive conditions).
  - Procedure codes representing tests that may reasonably be performed on a fetus through prenatal diagnosis are covered services more than once per lifetime. When applicable, claims should include the following ICD code to indicate prenatal diagnosis: O35.2X.

- All claims submitted for the defined procedure codes will be checked for previous payment in historical claims data.
- Testing more than once per lifetime is not medically necessary and such claims will be denied for reimbursement if:
  - The procedure code is known to have already been paid for that member, and
    - The member is a male, or
    - The member is a female, and
      - The code does not allow a prenatal diagnosis override, or
      - No ICD code suggesting prenatal diagnosis is submitted for a code that does allow a prenatal diagnosis override

**Supporting Documents**

For more information, see the Supporting Documents section at the end of this guideline.
Frequency Rules

Tests that do not involve unchanging, inherited DNA may be repeated for medically necessary reasons. Any limits to the frequency at which such tests should be repeated is defined in the applicable clinical policy. These frequency limits will be assessed at claim submission based on available historical claims data.

Maximum Units per Date of Service

Most procedure codes have a reasonable maximum number of expected units that should be billed on a single date of service. Maximum expected units are coded into claims systems to prevent billing, data entry, and payment errors.

The CMS National Correct Coding Initiative provides guidance on maximum units for many procedure codes through their Medically Unlikely Edits. When not provided by CMS, maximum units are established based on code definitions, specialty guidelines, peer reviewed literature, expert opinion and claims experience with those codes.

Maximum units per date of service rules are administered as follows:

- See the Maximum Units table for all procedure codes that have established maximum units.
- The allowable daily maximum units may only be addressed in this table and nowhere else in any other policy.
- Total billed units are calculated based on the combined number of times a procedure code is billed on a single date of service. This applies to codes billed with multiple units on a single claim line, units reported on separate claims lines on the same claim, or multiple units reported on separate claims for that date of service.
- When multiple units are billed, only the number of units up to the allowable daily maximum will be reimbursed.
- Some unusual circumstances justify exceeding the established maximum units per date of service.
  - When such exceptions are recognized in eviCore clinical policy, instructions for submitting claims with additional units are provided.
  - When exceptions are not specifically addressed in policy, reimbursement of additional units will be considered if supporting documentation is provided.

Supporting Documents

For more information, see the Supporting Documents section at the end of this guideline.

CCI Code Pair Edits

When two or more procedure codes are billed for the same member, on the same date of service, by the same provider, those codes must be compared to ensure the
procedures are distinct from each other and not mutually exclusive. The CMS National Correct Coding Initiative provides guidance through their Column One/Column Two code pair edits.

CCI code pair edits are administered as follows:

- Providers should bill only the most comprehensive procedure code(s) that represent the performed procedures with the fewest number of codes possible.
- See the **CCI Code Pair Edits** table for all code pairs considered to be components of one another or mutually exclusive. When two codes are billed together that appear in columns 1 and 2 of this table, the code in column 1 is paid, while the code in column 2 is denied as inclusive.
- Some unusual circumstances justify billing both codes in a pair for separate, reasonable indications. In such cases, an appropriate modifier may be used to override the CCI edit as outlined in the provided table. Additional supporting documentation to explain the necessity of both procedures may be required.

**Supporting Documents**

For more information, see the Supporting Documents section at the end of this guideline.

**Professional and Technical Component Modifiers**

Modifiers may be used to convey when only the professional or technical components of a test have been performed separately by the billing provider.

- Modifier 26 represents the professional component, such as the clinical interpretation of a test.
- Modifier TC represents the technical component, such as the equipment, supplies, and technical work.

The CMS Professional Component/Technical Component (PC/TC) indicators in the National Physician Fee Schedule Relative Value File are used to determine whether a procedure code is eligible for separate reimbursement for professional and technical components. The file is available at: [http://www.cms.gov/Medicare/Medicare-Fee-for-Service-Payment/PhysicianFeeSched/PFS-Relative-Value-Files.html](http://www.cms.gov/Medicare/Medicare-Fee-for-Service-Payment/PhysicianFeeSched/PFS-Relative-Value-Files.html). In some instances, professional services are billed separately from technical services even when the billing code does not have a modifier 26 or TC designation. This policy applies to all instances of services where there are separate professional and technical charges.

Professional and technical modifiers are administered as follows:

- See the **PC&TC Modifiers** table for all codes subject to the following modifier claims rules.
• If a procedure code is eligible for separate PC/TC reimbursement (PC/TC indicator 1), that code may be billed with no modifier to represent a global service (both PC and TC components) or the modifiers 26 or TC, but not both.

• Procedure codes that are designated as only global without being eligible for separate PC/TC component reimbursement should not be billed with modifier 26 or TC, and will be subject to denial if inappropriately billed.

• Physician specialty and CMS place of service codes are used to determine eligibility to bill professional or technical components.

Supporting Documents
For more information, see the Supporting Documents section at the end of this guideline.

Add-on Codes

Some procedure codes are defined as “add-on” codes that should always be supplemental to a separate primary code. The CMS National Correct Coding Initiative provides guidance through their Add-on Code Edits. Language in the code description or knowledge of testing practices may also be used.

Add-on code edits are administered as follows:
• See the Add-on Codes table for all codes subject to the add-on claims rules.
• Add-on codes will not be reimbursed when not billed with their appropriate primary code(s) on the same date of service by the same provider.

Supporting Documents
For more information, see the Supporting Documents section at the end of this guideline.

References


Leber Hereditary Optic Neuropathy (LHON) Genetic Testing

Procedures addressed

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<td>MT-ND4, MT-ND6 Targeted Mutation Analysis</td>
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<td>Whole Mitochondrial Genome</td>
<td>81460</td>
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What is Leber Hereditary Optic Neuropathy

Definition

Leber Hereditary Optic Neuropathy (LHON) is a mitochondrial disorder that mainly affects the eye. It is characterized by bilateral painless subacute vision loss that begins in the second and third decades of life. It usually has onset between 15-30 years of age, and leads to rapid, progressive blindness. Visual acuity usually deteriorates to 20/200 or worse.¹⁻³

- The primary cell type that is lost in LHON is the retinal ganglion cell, which is highly susceptible to disrupted ATP production and oxidative stress.⁴
- A diagnosis of LHON can be made clinically. "The pathologic hallmark of LHON is the selective degeneration of the retinal ganglion cell layer and optic nerve."¹
- LHON has three phases:¹
  - Presymptomatic/subacute phase: Mild abnormalities in the fundus may be present. Additionally, color vision, contrast, and electroretinogram may be mildly affected.
  - Acute phase: Onset features blurred or clouded central vision usually starting in one eye, followed by other eye within weeks to months. Onset involves both eyes simultaneously in about 25% of cases. The vision loss gets progressively worse with the blurred central field enlarging (called a scotoma). Evaluation of fundus in 80% of affected patients will show disk swelling, edema of the peripapillary nerve fiber layer, retinal telangiectasia, and increased vascular tortuosity without corresponding leakage on fluorescein angiography.
o Atrophic phase: Optic atrophy and worsening central scotoma will progress to severe impairment over the course of six weeks. Once the atrophic phase begins, visual acuity rarely recovers. Most individuals become legally blind.

- Within 1 year, 97% of those affected have involvement of the second eye, such that a patient presenting with a unilateral optic neuropathy for longer than 1 year is highly unlikely to suffer from LHON-related vision loss.

- Other neurologic features may include: tremor, peripheral neuropathy, myopathy, and/or movement disorders. Additionally, women may develop a multiple sclerosis-like progressive disease.

- Unaffected LHON point mutation carriers can display subclinical signs of disease on fundus examination, including peripapillary microangiopathy, zones of mild disc pseudoeoedema, and telangiectasia.

- Some clinicians treat children presymptomatically with antioxidants when their genetic status is known.

- People who have a pathogenic variant consistent with LHON should avoid alcohol and smoking.

- The prevalence of LHON in most populations is unknown. In Caucasian populations estimates range from 1 in 31,000 to 1 in 50,000. Men are about 4-5 times more likely to develop LHON than women.

- LHON is caused by point mutations in the mitochondrial genome which is separate from nuclear DNA.

- Several mtDNA mutations have been reported to cause LHON. However, 90% of affected individuals have one of three common mitochondrial mutations: G3460A (13%), G11778A (70%) and T14484C (14%).

- A 2016 expert-authored review stated the following regarding genotype-phenotype correlations:
  o The mtDNA mutation T14484C is associated with a partial recovery rate of 37%–58%, while the G11778A mutation has the lowest partial recovery rate of 4%. Patients with the G3460A mutation have an intermediate prognosis, with an approximate 20% partial recovery rate.
  o “m.3460G>A is associated with the worst impairment in visual function. m.11778G>A has an intermediate phenotype. Although published reports would appear to indicate otherwise, the m.3460G>A pathogenic variant is generally accepted among experts as having the worst visual recovery rate.”

- Earlier age of onset (younger than 20 years), a subacute time course of vision loss, and larger optic discs are all associated with a better visual prognosis.

- Mitochondrial DNA (mtDNA) is passed from the maternal gamete (oocyte) to the developing fetus, therefore mitochondrial diseases like LHON are inherited through the maternal lineage. Since sperm do not contribute mitochondria (and mtDNA) to a fetus, men cannot pass on any mitochondrial mutations they may carry.
About 60% of people with LHON have an identifiable maternal family history of disease. In the remaining 40%, the family history may be incomplete or the affected individual could have a new (de novo) mutation but this is rare.\(^1,\)\(^2\)

Not all people with an LHON disease-causing mtDNA mutation will develop symptoms. Only about 50% of males and 10% of females who have a known disease-causing LHON mutation will develop blindness.\(^2\) There must be other genetic and environmental factors that explain the variable appearance of symptoms and the gender differences.\(^1,\)\(^2\)

- Diseases like LHON that are attributed to mtDNA mutations have unique patterns of inheritance and penetrance governed by the principles of maternal inheritance, heteroplasmy, replicative segregation, and the critical threshold. Heteroplasmy and replicative segregation contribute to the heterogeneity of mitochondrial disease phenotypes, even among related individuals. Critical threshold is reached when the wild-type mtDNA cannot compensate for the mutant mtDNA in a cell or tissue. This accounts for targeted tissue involvement and age dependent onset. Even more variability is present because tissue-specific segregation of mutant mtDNA is stochastic during embryogenesis.\(^4\)

**Test information**

- An ophthalmological evaluation can confirm the diagnosis of LHON:\(^1,\)\(^2\)
  - Eye testing may include fundus exam, visual field testing, and imaging. Other testing, including angiography and electrophysiology, are sometimes warranted. This testing may reveal characteristic findings of LHON or rule out other causes of acute vision loss.
  - In cases where a diagnosis can't be confirmed by eye findings alone, molecular genetic testing may be useful.

- The LHON three mtDNA mutation panel involves targeted testing of three common mutations in mtDNA (G3460A, G11778A and T14484C).\(^1\)\(^3\) These three mutations account for over 90% of mtDNA mutations found in people with LHON.\(^1\)

- Full sequencing of the entire mitochondrial genome can be done to identify the remaining 10% of mtDNA mutation in individuals affected with LHON. Since the mitochondrial genome is highly polymorphic, this is not routinely offered unless clinical suspicion is very high and paternal transmission has been ruled out.\(^1\) If the status of heteroplasmy is of concern, next generation testing with high read depth may be preferable.\(^5\)

- The three LHON mutations are also included on a number of more general mitochondrial targeted mutation panels (in conjunction with genes for MELAS, MERFF and Leigh syndrome).

- A number of large panels sequence the mitochondrial genome in conjunction with nuclear-encoded mitochondrial genes for a broad approach to testing.
• DNA testing can be performed on a blood specimen. Muscle biopsy is generally not necessary, but some labs accept blood, saliva and muscle samples.\textsuperscript{6}

**Guidelines and evidence**

• No evidence-based U.S. testing guidelines were identified for LHON.

• Although not specific to genetic testing for LHON, the Mitochondrial Medicine Society (2015)\textsuperscript{7} developed consensus recommendations for the diagnosis and management of mitochondrial disease. Testing strategies, including strategies for genetic testing, were discussed.

  o Recommendations for DNA testing include the following:

    ▪ “Massively parallel sequencing/NGS of the mtDNA genome is the preferred methodology when testing mtDNA and should be performed in cases of suspected mitochondrial disease instead of testing for a limited number of pathogenic point mutations.”

    ▪ “Patients with a strong likelihood of mitochondrial disease because of a mtDNA mutation and negative testing in blood, should have mtDNA assessed in another tissue to avoid the possibility of missing tissue-specific mutations or low levels of heteroplasmy in blood; tissue-based testing also helps assess the risk of other organ involvement and heterogeneity in family members and to guide genetic counseling.”

    ▪ “Heteroplasmy analysis in urine can selectively be more informative and accurate than testing in blood alone, especially in cases of MELAS due to the common m.3243 A>G mutation.”

    ▪ “When considering nuclear gene testing in patients with likely primary mitochondrial disease, NGS methodologies providing complete coverage of known mitochondrial disease gene is preferred. Single-gene testing should usually be avoided because mutations in different genes can produce the same phenotype. If no mutation is identified via known NGS panels, then whole exome sequencing should be considered.”

• The European Federation of Neurological Sciences (2009)\textsuperscript{8} provide consensus-based guidelines for LHON genetic testing: “If the phenotype suggests syndromic mitochondrial disorder due to mtDNA point mutations (MELAS, MERRF, NARP, LHON), DNA-microarrays using allele-specific oligonucleotide hybridisation, real-time-PCR or single-gene sequencing are indicated.”

• The Clinical Molecular Genetics Society of the United Kingdom (2008)\textsuperscript{6} provided practice-based guidelines for the molecular diagnosis of mitochondrial disease: “Investigation for the G3460A, G11778A and T14484C mutations are indicated for all LHON referrals. Secondary mutations should not be investigated as their significance is unknown. Presymptomatic testing for LHON should be undertaken cautiously and homoplasy/heteroplasmy should be stated on the report.”

• A 2016 expert-authored review suggests the following testing strategy for those with a known or suspected diagnosis of LHON:
“Three common mtDNA pathogenic variants account for 90%-95% of LHON. Targeted analysis for one of these three variants should be performed first.”

“A multi-gene panel that includes the mitochondrial genes that encode subunits of NADH dehydrogenase, MT-ND1, MT-ND2, MT-ND4, MT-ND4L, MT-ND5, and MT-ND6, which are known to cause LHON and other genes of interest may also be considered.”

“Complete mtDNA sequencing may be considered if use of targeted testing and/or a multi-gene panel did not identify a pathogenic variant, clinical suspicion remains high, and there is no evidence of paternal transmission.”

For those seeking predictive testing (e.g. they are not currently affected), this review states:\textsuperscript{1}

“Testing of at-risk asymptomatic adults for LHON is possible ... Such testing is not useful in predicting age of onset, severity, or rate of progression of visual loss in asymptomatic individuals.”

“Testing of asymptomatic individuals younger than age 18 years who are at risk for adult-onset disorders for which no treatment exists is not considered appropriate.”

Criteria

**LHON known familial mutation testing**

- Genetic Counseling:
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous genetic testing for LHON, and
  - LHON causing mutation identified in 1st degree biological maternal relative, AND
- Predictive Testing for Asymptomatic Individual:
  - 18 years of age or older, or
  - Under the age of 18 years, and
    - Presymptomatic treatment with antioxidants is being considered, OR
- Diagnostic Testing for Symptomatic individuals:
  - Ophthalmology examination is suggestive, but not confirmatory, of a diagnosis of LHON, OR
- Prenatal Testing for At-Risk Pregnancies:
- LHON disease-causing mutation identified in a previous child or in the mother, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy

**LHON targeted mutation analysis (G3460A, G11778A and T14484C)**

- Genetic Counseling:
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  - No previous genetic testing for LHON, and
  - No known LHON mutation in the family, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Ophthalmology examination is suggestive, but not confirmatory, of a diagnosis of LHON, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

**Whole mtDNA sequencing**

- Genetic Counseling:
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Criteria for LHON targeted mutation analysis is met, AND

- No mutations identified in the targeted mutation analysis, AND

- Paternal transmission has been ruled out

**References**


Li-Fraumeni Syndrome

Introduction

Li-Fraumeni syndrome testing is addressed by this guideline.

Procedures addressed

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What is Li-Fraumeni syndrome

Definition

Li-Fraumeni syndrome (LFS) is a hereditary cancer-predisposition syndrome typically associated with soft tissue sarcoma, osteosarcoma, premenopausal breast cancer, brain tumor, and adrenocortical carcinomas. People with LFS also have an increased risk of a variety of other cancers.1-3

Cause

Historically, there are two forms of LFS: Classic LFS, and Li-Fraumeni-like syndrome (LFL).1 LFL shares some of the features for LFS, but has less strict clinical diagnostic criteria.1 LFS/LFL are caused by mutations in the TP53 gene.

Prevalence

Prevalence of inherited p53 mutations is estimated to be 1 in 20,000.1 The likelihood of detecting a TP53 mutation is about 70% in classic LFS cases and 40-50% in LFL cases.1
Inheritance

This condition is inherited in an autosomal dominant manner.¹ Children of an affected person have a 1 in 2 (50%) chance to be affected. Most TP53 mutations are inherited from an affected parent.¹ The frequency of de novo mutations is not well defined but may be as high as 20%.¹

Prognosis

About 50% of individuals with LFS/LFL will have cancer by 30 years of age, and 90% of individuals with LFS/LFL will have cancer by 60 years of age.¹

Test information

Introduction

Testing for Li-Fraumeni may include sequence analysis, deletion/duplication analysis, or known familial mutation analysis.

Sequence analysis

Complete p53 gene sequencing will detect approximately 95% of known mutations.¹ Limited seqencing of only certain regions of the p53 gene is also available. The detection rate of the limited sequencing tests varies between 70-90% depending on which portions of the gene are screened.¹

Deletion/duplication testing

Deletion/duplication testing may be considered as a reflex test if a mutation is not found by sequencing. This method will identify gene rearrangements in an additional 1% of cases.

Known familial mutation analysis

Once a mutation has been identified in the family, known familial mutation testing can be done for at-risk family members.¹²

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to Li-Fraumeni testing.
National Comprehensive Cancer Network

The National Comprehensive Cancer Network (2017) guidelines outline the following Li-Fraumeni syndrome testing criteria (quoted directly). These are considered a category 2A recommendation “lower level evidence with uniform NCCN consensus”.

- Individuals from a family with a known TP53 mutation, OR
- Classic Li-Fraumeni syndrome when ALL of the following are present:
  - Combination of an individual diagnosed less than age 45 years of age with a sarcoma; AND
  - First-degree relative diagnosed less than 45 years of age with cancer; AND
  - An additional first- or second-degree relative in the same lineage with cancer diagnosed less than 45 years of age, or a sarcoma at any age OR
- Chompret Criteria (2015 version), when ANY of the following are present:
  - Individual with a tumor from LFS tumor spectrum (for example, soft tissue sarcoma, osteosarcoma, CNS tumor, breast cancer, adrenocortical carcinoma), before 46 years of age, and at least one first- or second-degree relative with any of the aforementioned cancer (other than breast cancer if the proband has breast cancer) before the age of 56 years, or with multiple primaries at any age; OR
  - Individual with multiple tumors (except multiple breast tumors), two of which belong to LFS tumor spectrum with the initial cancer occurring before the age of 46 years; OR
  - Individual with adrenocortical carcinoma or choroid plexus carcinoma or rhabdomyosarcoma of embryonal anaplastic subtype, at any age of onset, regardless of the family history
- Early onset breast cancer
  - Individual with breast cancer diagnosed before 31 years. TP53 testing can be ordered alone, concurrently with BRCA1/2 testing and/or other gene testing or as a follow up test after negative BRCA1/2 testing.

Criteria

Introduction

Requests for Li-Fraumeni testing are reviewed using these criteria.

TP53 Known Familial Mutation Analysis

- Genetic Counseling:
- Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  - No previous genetic testing of TP53, AND

- Diagnostic and Predisposition Testing for Presymptomatic/Asymptomatic Individuals**:
  - Known family mutation in TP53

** Includes prenatal testing for at-risk pregnancies.

**TP53 Sequencing**

- Genetic Counseling:
  - Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy). AND

- Previous Testing:
  - No previous sequencing of TP53, and
  - No previous duplication/deletion analysis, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Classic Li-Fraumeni syndrome when **ALL** of the following are present:
    - Combination of an individual diagnosed less than age 45 years of age with a sarcoma; and
    - First-degree relative diagnosed less than 45 years of age with cancer; and
    - An additional first- or second-degree relative in the same lineage with cancer diagnosed less than 45 years of age, or a sarcoma at any age, OR
  
  - Chompret Criteria (2015) are met when **ANY** of the following are present:
    - Individual with a tumor from LFS tumor spectrum (eg, sarcoma, CNS tumor, breast cancer, osteosarcoma, adrenocortical carcinoma, leukemia, or lung bronchoalveolar cancer) before age 46 years, and at least one first- or second-degree relative with any of the aforementioned cancers (other than breast cancer if the proband has breast cancer) under the age of 56 years or with multiple primaries at any age; or
    - Individual with multiple tumors (except multiple breast tumors), two of which are LFS tumor spectrum (eg, sarcoma, CNS tumor, breast cancer, osteosarcoma, adrenocortical carcinoma, leukemia, or lung bronchoalveolar cancer) with the initial cancer occurring before the age of 46 years, regardless of the family history; or
• Individual with adrenocortical carcinoma or choroid plexus carcinoma or rhabdomyosarcoma of embryonal anaplastic subtype, at any age of onset, regardless of the family history, OR
  o Early onset breast cancer

• Individual with breast cancer diagnosed before 31 years of age

Predisposition Testing for Presymptomatic/Asymptomatic Individuals:

• One or more biologic relatives (1st, 2nd, or 3rd degree) with a clinical diagnosis of LFS/LFL (according to criteria above) and no known family mutation or no testing to date

TP53 Deletion/Duplication Analysis

• Genetic Counseling:
  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous deletion analyses of TP53, and
  o No mutation detected on full sequencing of TP53.

References

Introduction

These references are cited in this guideline.


**Procedures addressed**

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**What is liquid biopsy testing**

**Definition**

The use of cell-free circulating tumor DNA (ctDNA) to identify genetic mutations present in a tumor is also referred to as a liquid biopsy.

- The National Cancer Institute defines a liquid biopsy as “a test done on a sample of blood to look for cancer cells from a tumor that are circulating in the blood or for pieces of DNA from tumor cells that are in the blood. A liquid biopsy may be used to help find cancer at an early stage. It may also be used to help plan treatment or to find out how well treatment is working or if cancer has come back. Being able to take multiple samples of blood over time may also help doctors understand what kind of molecular changes are taking place in a tumor.”

- Cell-free circulating tumor DNA (ctDNA) is released into circulation by tumors. It can be found in various substances, including blood, urine, saliva, etc.
Analysis of ctDNA is performed to help identify indicators of disease recurrence or disease progression. It can also help to determine if a specific treatment is indicated.\(^1\)

Liquid biopsies can be used to more easily obtain serial sampling of a tumor. This is particularly useful since genetic mutations that are used in treatment decisions can change as the tumor progresses.\(^1\) ctDNA is also thought to be a more representative sample of the entire tumor genome as well as any metastases that may be present.\(^1\)

Traditional methods of performing biopsies on tumor tissue pose the following problems:\(^1,3\)
- Biopsies are invasive, involve risks, are typically costly, and are typically difficult to obtain.
- Treatment decisions often rely on one single biopsy, while tumors are usually heterogeneous in nature, tumor characteristics can evolve, and information regarding metastases may not be not known.\(^1\)

The use of liquid biopsies can help overcome some of the above problems with traditional biopsies since they can be completed non-invasively.

This policy will only address the use of ctDNA as a liquid biopsy in solid tumors. This policy does not address the use of ctDNA as a liquid biopsy in hematological malignancies.

**Test information**

- Testing methodology relies on the presence of ctDNA in circulation.
- Various laboratories have recently developed liquid biopsy tests (e.g. Guardant Health, Trovagene, Biocept, Transgenomic).
- Different methodologies are used to detect ctDNA in blood. These include spectrophotometric methods, fluorescent dyes, or quantitative PCR-based methods.\(^1\)
- Once ctDNA is detected, it is typically analyzed by one of the following methods:
  - Standard testing methodologies, such as PCR or sequencing, are used to identify targeted mutations. In order to detect targeted mutations in ctDNA, the genetic changes present in the tumor need to be known.\(^1\)
Methodologies, such as NGS-based approaches or array-CGH, are used to identify untargeted mutations. These include whole genome sequencing or whole exome sequencing. Use of these approaches allows testing with no knowledge of genetic mutations that are present in the tumor.1

**Guidelines and evidence**

- The National Comprehensive Cancer Network (NCCN, 2017) states the following in regards to liquid biopsies for EGFR T790M testing in patients with non-small cell lung cancer:4
  - In patients with a sensitizing EGFR mutation at progression, tissue biopsy testing for EGFR T790M testing should be performed.
  - “Recent data suggest that plasma genotyping (also known as liquid biopsy or plasma biopsy) may be considered instead of tissue biopsy to detect whether patients have T790M; however, if the plasma biopsy is negative, then tissue biopsy is recommended if feasible.”

- Many laboratories are developing liquid biopsies assays. For many of these assays, analytical validity studies have been performed; however, data regarding the clinical validity and clinical utility of these tests is still emerging.3,5-9

- The TRACERx study (Tracking Non-small cell lung cancer evolution through therapy (Rx)) is a large, prospective clinical trial being conducted to evaluate “the relationship between intra-tumor heterogeneity and clinical outcome following surgery and adjuvant therapy.”10 Researchers plan to analyze patient’s tumors before surgery and multiple times after surgery during their treatment regimen. Tumor tissue and ctDNA in patient’s blood will be examined in approximately 840 patients with NSCLC. This trial is expected to continue until 2023.10

**Criteria**

- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References
## Long QT Syndrome Testing

### Procedures addressed

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What is Long QT syndrome
Definition

LQTS is caused by mutations in a number of genes, most of which are related to the functioning of sodium or potassium ion channels in the heart.\(^1\) Testing may offer prognostic information in some cases, as specific genes and even specific mutations within those genes may have some correlation to risk for sudden death, effectiveness of beta-blocker therapy, and preventive strategies.\(^1,3,4\)

- Signs and symptoms of long QT syndrome (LQTS) are variable, but may include a prolonged QT interval on an electrocardiogram, torsades de pointes, syncope, seizures, cardiac arrest, and sudden cardiac death.\(^1,2\)
- Symptoms typically occur in young individuals who are otherwise healthy.\(^1\) Certain events — such as exercise, emotional stress, a startle, or sleep — can trigger arrhythmia in individuals with LQTS.\(^1\) Patients are recommended to avoid these activities when possible.\(^1\)
- Screening for LQTS is by electrocardiography (ECG or EKG), and sometimes includes an ambulatory ECG (Holter monitor), and/or an exercise- or medication-induced stress test.\(^1,3\) In many cases, the diagnosis of LQTS can be made based on personal and family history and clinical findings.\(^1\) However, approximately 10-40% of LQTS patients will not have diagnostic ECG changes.\(^4\)
- Several forms of LQTS exist. The autosomal dominant Romano-Ward syndrome is the most common form, with a prevalence of 1 in 3000 to 1 in 5000.\(^1,2\) It affects all ethnic groups.\(^1\) All forms of LQTS are estimated to affect at least 1 in 2500 people.\(^4\)
- Genetic LQTS must be differentiated from acquired LQTS, which can be caused by exposure to certain medications, certain heart conditions, bradycardia, electrolyte imbalances, dietary deficiencies, or intracranial disease.\(^1\)

Test information

- Genetic testing for LQTS is typically performed with a sequencing panel. Commercially available genetic testing exists for the AKAP9, ANK2, CACNA1C, CAV3, CALM1, CALM2, CALM3, KCNE1, KCNE2, KCNH2, KCNJ2, KCNJ5, KCNQ1, SCN4B, SCN5A, SNTA1, and TRDN genes associated with LQTS.\(^1\) Mutations in three genes (KCNQ1, KCNH2, and SCN5A) account for the majority of cases.\(^1,2\) Testing will find a mutation in approximately 75% of patients with a clinical diagnosis of LQTS.\(^4\) Composition of test panels varies by laboratory.
- Deletion/duplication testing for the AKAP9, ANK2, CACNA1C, CAV3, KCNE1, KCNE2, KCNH2, KCNJ2, KCNQ1, SCN4B, SCN5A, SNTA1 genes is also available.
• Once the causative mutation has been identified in a family member, other at-risk relatives only need to be tested for that mutation — not a panel of genes. Testing by known familial mutation analysis is greater than 99% accurate.\(^1\)

Guidelines and evidence
A 2013 expert consensus statement from the Heart Rhythm Society (HRS), the European Heart Rhythm Association (EHRA), and the Asia Pacific Heart Rhythm Society incorporates genetic test results into the recommended diagnostic criteria:\(^5\)

• LQTS is diagnosed:
  o In the presence of an LQTS risk score ≥3.5 in the absence of a secondary cause for QT prolongation and/or
  o In the presence of an unequivocally pathogenic mutation in one of the LQTS genes or
  o In the presence of a corrected QT interval for heart rate using Bazett's formula (QTc) ≥500 ms in repeated 12-lead electrocardiogram (ECG) and in the absence of a secondary cause for QT prolongation.

• LQTS can be diagnosed in the presence of a QTc between 480 and 499 ms in repeated 12-lead ECGs in a patient with unexplained syncope in the absence of a secondary cause for QT prolongation and in the absence of a pathogenic mutation.

A 2011 expert consensus statement from the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA) makes the following recommendations regarding genetic testing:\(^4\)

• “Comprehensive or LQT1-3 (KCNQ1, KCNH2, and SCN5A) targeted LQTS genetic testing is recommended for any patient in whom a cardiologist has established a strong clinical index of suspicion for LQTS based on examination of the patient’s clinical history, family history, and expressed electrocardiographic (resting 12-lead ECGs and/or provocative stress testing with exercise or catecholamine infusion) phenotype.” [Class I, “is recommended”]\(^4\)

• “Comprehensive or LQT1-3 (KCNQ1, KCNH2, and SCN5A) targeted LQTS genetic testing is recommended for any asymptomatic patient with QT prolongation in the absence of other clinical conditions that might prolong the QT interval (such as electrolyte abnormalities, hypertrophy, bundle branch block, etc., i.e., otherwise idiopathic) on serial 12-lead ECGs defined as QTc>480ms (prepuberty) or >500ms (adults).” [Class I, “is recommended”]\(^4\)

• “Comprehensive or LQT1-3 (KCNQ1, KCNH2, and SCN5A) targeted LQTS genetic testing may be considered for any asymptomatic patient with otherwise idiopathic QTc values>460ms (prepuberty) or >480ms (adults) on serial 12-lead ECGs.” [Class IIB “may be considered”]\(^4\)

• “Mutation specific genetic testing is recommended for family members and other appropriate relatives subsequently following the identification of the LQTS-causative mutation in an index case.” [Class I, “is recommended”]\(^4\)
• Older American College of Cardiology/American Heart Association/European Society of Cardiology (2006) guidelines on the management of ventricular arrhythmias made no specific evidence-based recommendations about genetic testing for LQTS, but do state:
  o “[Genetic testing is] useful for risk stratification and for making therapeutic decisions,” and they highlight the benefit for identifying family members for counseling and preventative management. They conclude: “Although genetic analysis is not yet widely available, it is advisable to try to make it accessible to LQTS patients.”

Criteria

Long QT Syndrome Known Familial Mutation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous genetic testing for Long QT Syndrome inclusive of known family mutation, AND
• Diagnostic and Predisposition Testing:
  o Long QT Syndrome family mutation identified in 1st degree relative(s). (Note: 2nd or 3rd degree relatives may be considered when 1st degree relatives are unavailable or unwilling to be tested), AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy

Long QT Syndrome Sequencing or Multigene Panel

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous genetic testing for Long QT Syndrome, AND
• Diagnostic Testing for Symptomatic Individuals:
  o Clinical signs indicating moderate to high pre-test probability of Long QT syndrome, but diagnosis cannot be made with certainty by other methods (i.e. Schwartz criteria of 2-3), or
• Confirmation of prolonged QTc or T-wave abnormalities [>460ms (prepuberty) or >480ms (adults)] on serial 12-lead ECGs on exercise or ambulatory ECG, or during pharmacologic provocation testing and acquired cause has been ruled out, or
  • A prolonged or borderline prolonged QT interval on ECG or Holter monitor and acquired cause has been ruled out, or
  • Profound congenital bilateral sensorineural hearing loss and prolonged QTc, OR

• Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  • Biologic relative(s) (1st degree) diagnosed with LQTS clinically whose genetic diagnosis is unknown, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Long QT Syndrome Deletion/Duplication Analysis

• Genetic Counseling:
  • Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  • No mutation identified with long QT full gene sequence analysis, or
  • Neither or only one mutation in KCNQ1 or KCNE1 identified in an individual with profound congenital bilateral sensorineural hearing loss and prolonged QTc, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

References


Lynch Syndrome Genetic Testing

Introduction

Lynch syndrome genetic testing is addressed by this guideline.

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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<tr>
<td>MSH6 Known Familial Mutation Analysis</td>
<td>81299</td>
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<td>PMS2 Known Familial Mutation Analysis</td>
<td>81318</td>
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<tr>
<td>EPCAM Deletion/Duplication Analysis</td>
<td>81403</td>
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</tbody>
</table>

What is Lynch syndrome

Definition

Lynch syndrome, also called hereditary, non-polyposis colorectal cancer (HNPCC), is a hereditary cancer syndrome that is the most common cause of colon and endometrial cancer.
Prevalence

Lynch syndrome affects approximately 1 in 35 colorectal and endometrial cancer patients and around 1 in 370 individuals in the general population. Lynch syndrome accounts for 2-4% of all colorectal cancer cases.\(^1-3\)

Cancer risks

Lynch syndrome is associated with an 82% lifetime risk for colorectal cancer and a 15-60% risk of endometrial cancer.\(^4,5\) The risk also increases for development of the following cancer types:

- small bowel
- stomach
- ovarian
- pancreatic
- ureteral and renal pelvis
- biliary tract
- brain
- sebaceous adenoma, and
- keratoacanthoma tumors.\(^1,5\)

Onset

The average ages of diagnosis for colorectal, endometrial, and gastric cancers are 44-61, 48-62, and 56 years, respectively.\(^4\) Ovarian cancer diagnoses are typically earlier, with an average age of diagnosis of 42.5 years, roughly one-third of cases being diagnosed before the age of 40.\(^4\)

Diagnosis

Lynch syndrome should be suspected when the personal and family cancer history meets the Revised Bethesda Guidelines or the Amsterdam II Criteria (see below).\(^6,7\)

Cause

Lynch syndrome is caused by mutations in any one of at least the following five genes.\(^4,8\)

MLH1

MLH1 accounts for 32%-50% of Lynch syndrome-causing mutations. Sequencing identifies most mutations. An additional 5% can only be identified by a deletion and duplication analysis.
MSH2

MSH2 accounts for 38-40% of Lynch syndrome-causing mutations. Most are found by sequencing, but 20% are detectable only by deletion/duplication analysis.

MSH6

MSH6 accounts for 7%-10% of Lynch syndrome-causing mutations. Most will be found by sequencing, but an estimated 7% are detectable only by deletion and duplication analysis.

PMS2

PMS2 accounts for <5% of Lynch syndrome-causing mutations. Most are found by sequencing, but 20% are only detectable by deletion and duplication analysis.

EPCAM

EPCAM accounts for about 1%-3% of Lynch syndrome-causing mutations. To date, all mutations have been deletions detectable by deletion/duplication analysis, not sequencing.

Inheritance

Lynch syndrome is an autosomal dominant syndrome that is associated with a germline mutation in one of at least five genes: MLH1, MSH2, MSH6, PMS2, and EPCAM. Children of an affected individual have a 50% risk to inherit a mutation. Lynch syndrome mutations inherited in an autosomal recessive manner cause constitutional MMR deficiency syndrome (CMMR-D). Testing for CMMR-D is not addressed in this summary.

Associated syndromes

Lynch syndrome includes the variants Muir-Torre syndrome (one or more Lynch syndrome-associated cancers and sebaceous neoplasms of the skin) and Turcot syndrome (Lynch syndrome with glioblastoma).

Test information

Introduction

Testing for Lynch syndrome may include tumor testing, gene sequencing, deletion/duplication analysis, known familial mutation testing, or multigene panel testing.
Testing approaches

Testing those with a suspected Lynch syndrome-related cancer should begin with microsatellite instability or immunohistochemistry testing on tumor tissue. The following table lists and describes the various testing scenarios.

<table>
<thead>
<tr>
<th>When ...</th>
<th>Then ...</th>
</tr>
</thead>
<tbody>
<tr>
<td>tumor tests suggest Lynch syndrome</td>
<td>that individual should be offered genetic testing to look for a mutation that causes Lynch syndrome.¹,⁸⁻¹⁰</td>
</tr>
<tr>
<td>immunohistochemistry studies are abnormal</td>
<td>those results may suggest which of four possible mismatch repair genes is likely to harbor a mutation.</td>
</tr>
<tr>
<td>tumor tests are normal, and a strong family history of Lynch syndrome-associated cancers is present</td>
<td>genetic testing may still be warranted, or tumor testing in another family member with the most suspicious cancer history may be considered.⁸</td>
</tr>
<tr>
<td>tumor screening is not possible, and the individual meets the guideline criteria</td>
<td>direct genetic testing may be reasonable.</td>
</tr>
</tbody>
</table>

Genetic testing

Genetic testing usually starts with sequencing and deletion and duplication analysis of the MLH1 and MSH2 genes¹ because they account for most Lynch syndrome cases.⁴ The first person tested should be the relative most likely to have Lynch syndrome in the family.

When the family Lynch syndrome mutation is known, at-risk relatives should be tested for that specific mutation only. This is often called single site mutation analysis. Detection rates approach 100%.

Guidelines and evidence

Introduction

This section includes relevant guidelines and evidence pertaining to Lynch syndrome genetic testing.

Multiple society recommendations

The US Multi-Society Task Force (2014), the National Society of Genetic Counselors and the Collaborative Group of the Americas on Inherited Colorectal Cancer (NSGC/CGA-ICC, jointly published, 2012), the National Comprehensive Cancer Network (NCCN, 2016), and the American College of Gastroenterology (ACG); (2015)
have practice guidelines that address Lynch syndrome genetic testing. Generally, these recommendations agree:\textsuperscript{1,8,9,11}

- Test colorectal or endometrial tumors by microsatellite instability and/or immunohistochemistry first when tissue is available.

- Individuals with abnormal microsatellite instability and/or immunohistochemistry results (and no demonstrated BRAF mutation or hypermethylation of MLH1) should be offered genetic testing to identify a Lynch syndrome disease-causing mutation. Results from tumor testing should guide the genetic testing cascade. When tumor testing is not possible or results are inconclusive, genetic testing for an inherited mutation is indicated if a patient with a suspected Lynch syndrome-related cancer meets one of the first three Bethesda Guidelines or the family meets the Amsterdam Criteria (see tables below). If no affected family member is available for testing, at-risk relatives can consider genetic testing if the family meets the Amsterdam Criteria. However, only a mutation positive result can be clearly interpreted. Mutation negative results must be interpreted with caution; the chance of inconclusive results is high because the family mutation may not be detectable. Once a Lynch syndrome disease-causing mutation has been identified, at-risk relatives should be offered genetic testing for that specific mutation.

“The Multi-Society Task Force is composed of gastroenterology specialists with a special interest in CRC, representing the following major gastroenterology professional organizations: American College of Gastroenterology, American Gastroenterological Association Institute, and the American Society for Gastrointestinal Endoscopy. Also, experts on LS from academia and private practice were invited authors of this guideline. Representatives of the Collaborative Group of the Americas on Inherited Colorectal Cancer and the American Society of Colon and Rectal Surgeons also reviewed this manuscript. In addition to the Task Force and invited experts, the practice committees and Governing Boards of the American Gastroenterological Association Institute, American College of Gastroenterology, American Society for Gastrointestinal Endoscopy reviewed and approved this document.”

**Society of Gynecologic Oncology**

The Society of Gynecologic Oncology recommends “all women who are diagnosed with endometrial cancer should undergo systematic clinical screening for Lynch syndrome (review of personal and family history) and/or molecular screening. Molecular screening of endometrial cancer for Lynch syndrome is the preferred strategy when resources are available.” Universal molecular tumor testing for either all endometrial cancer or cancers diagnosed at age less than 60, regardless of personal or family cancer history, is a sensitive strategy for identifying women with Lynch syndrome.\textsuperscript{12}

**Revised Bethesda Guidelines**

According to the *Revised Bethesda Guidelines*, consider Lynch syndrome tumor screening when any one of the following criteria are met:
• colorectal cancer is diagnosed before the age of 50
• presence of synchronous or metachronous colorectal cancer, or other Lynch syndrome-associated tumor***, regardless of age
• microsatellite unstable (MSI-H) tumor pathology before the age of 60, examples include
  o tumor-infiltrating lymphocytes
  o Crohn's-like lymphocytic reaction
  o mucinous or signet-ring differentiation
  o medullary growth pattern, or
  o other reported features
• at least one first-degree relative, including parent, sibling, or child with a Lynch syndrome-related tumor***, one of whom was diagnosed before the age of 50, or
• at least two first- or second-degree relatives with Lynch syndrome-related tumors*** at any age.

Amsterdam II Criteria

According to Amsterdam II Criteria, Lynch syndrome is likely when all of the following criteria are met:
• there are at least three relatives with Lynch syndrome associated tumors***
• one affected relative is a first-degree relative (parent, sibling, child) of the other two
• affected relatives are in two or more successive generations
• at least one Lynch syndrome-related tumor was diagnosed before age 50, and
• FAP has been excluded on the basis of no polyposis.

Tumors must be verified by pathology.

***Lynch syndrome-associated tumors include
• colorectal
• endometrial
• small bowel
• stomach
• ovarian
• pancreatic
• ureteral and renal pelvis
• biliary tract
• brain tumors, usually glioblastomas associated with Turcot syndrome variant
• sebaceous adenomas, and
• keratoacanthomas, associated with a Muir-Torre syndrome variant.

Criteria

Known Familial Mutation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Testing:
  o No previous testing for inherited Lynch syndrome mutations, AND
• Family History:
  o Known MLH1, MSH2, MSH6, PMS2, or EPCAM mutation in a close blood relative (1st, 2nd, or 3rd degree), AND
• Age- 18 years and older, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Gene Sequencing and/or Deletion/Duplication Analysis of MLH1, MSH2, MSH6, PMS2, or EPCAM

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Testing:
  o Gene requested has not been tested previously by the same methodology (i.e., sequencing or deletion/duplication analysis), AND
• Age- 18 years or older, AND
• Diagnostic Testing for Symptomatic Individuals\textsuperscript{1,13}
  o Personal history of colorectal cancer (or other Lynch syndrome-related tumor\textsuperscript{10}), and
  o If colorectal cancer (see figure A):
    ▪ MSI testing of tumor tissue shows MSI-high, or
    ▪ IHC testing of tumor tissue detects absence of MLH1, MSH2, MSH6, and/or PMS2 encoded protein products, and
BRAF mutation analysis and/or MLH1 hypermethylation analysis performed if indicated (according to figure A) and not consistent with sporadic CRC (sporadic CRC is likely when the tumor has MLH1 promoter hypermethylation and/or the BRAF V600E mutation.), or

- If other LS-associated tumor:
  - Endometrial cancer diagnosed before age 50, or
  - Presence of synchronous or metachronous Lynch syndrome-associated tumors, regardless of age, or
  - Amsterdam II criteria are met:
    - ≥ 3 close blood relatives (1st, 2nd, or 3rd degree) with Lynch syndrome-associated tumor (symptomatic member can be one of the three), and
    - One should be a first-degree relative of the other two, and
    - ≥ 2 successive generations affected, and
    - ≥ 1 diagnosed before age 50, or
  - 5% or greater risk of Lynch syndrome based on one of the following mutations prediction models (MMRPro or MMRpredict)\(^1,10,11\), or
  - 2.5% or greater risk of Lynch syndrome based on PREMM[5]\(^\_14\), OR

- Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  - ≥ 3 close blood relatives (1st, 2nd, or 3rd degree) with Lynch syndrome-associated tumor, where Amsterdam II criteria are met:
    - One should be a first degree relative of the other two, and
    - ≥ 2 successive generations affected, and
    - ≥ 1 diagnosed before age 50, and
    - Familial adenomatous polyposis (FAP) ruled out, and
  - IHC and/or LS genetic testing results from affected family member are unavailable, OR
  - 5% or greater risk of Lynch syndrome based on one of the following mutations prediction models (MMRPro or MMRpredict)\(^1,10,11\), OR
  - 2.5% or greater risk of Lynch syndrome based on PREMM[5]\(^\_14\), AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

***Lynch syndrome-associated tumors include colorectal, endometrial, small bowel, stomach, ovarian, pancreatic, ureteral and renal pelvis, biliary tract, brain/CNS tumors (usually glioblastomas associated with Turcot syndrome variant), sebaceous adenomas, and keratoacanthomas (associated with Muir-Torre syndrome variant).
Billing and reimbursement considerations

- For individuals that have had previous tumor testing (MSI and/or IHC), the testing algorithm as outlined in Figure A must be followed for payment of claim.
• Lynch syndrome genetic testing for those with colorectal cancer is generally not indicated in the absence of abnormal MSI and/or IHC results on the colorectal tumor. MSI and/or IHC became part of the standard NCCN recommended evaluation for all people with colorectal cancer under the age of 70 (at a minimum) in May 2013. As a result, most people affected with colorectal cancer who are appropriate candidates for Lynch syndrome testing should have access to MSI and/or IHC. Lynch syndrome genetic testing without MSI and/or IHC results will only be considered necessary in extenuating circumstances and will require medical necessity review.
+ “Individuals with abnormal MSI and/or IHC tumor results and no germline mutation detected in the corresponding gene(s) may still have undetected Lynch syndrome. At this time, no consensus has been reached as to whether these patients should be managed as Lynch syndrome or managed based on personal/family history. Growing evidence suggests that the majority of these individuals with abnormal tumor results and no germline mutation found have double somatic mutations/changes in the MMR
genes. Although the efficacy has not yet been proven, genetic testing of the corresponding gene(s) could be performed on tumor DNA to assess for somatic mutations. Individuals found to have double somatic mutations/changes in the MMR genes likely do not have Lynch syndrome and management should be based on personal/family history.”

++“If strong family history (i.e. Amsterdam criteria) or additional features of hereditary cancer syndromes (multiple colon polyps) are present, additional testing may be warranted in the proband, or consider tumor testing in another affected family member due to the possibility of a phenocopy.”

References

Introduction

The following references are cited throughout the Lynch Syndrome documentation.


Lynch Syndrome Tumor Screening - First-Tier

Procedures addressed

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What is Lynch syndrome tumor screening

Definition

Lynch syndrome, also called hereditary non-polyposis colorectal cancer (HNPCC), is the most common known hereditary cause of colon and endometrial cancer. It affects approximately 1 in 35 colorectal and endometrial cancer patients and around 1 in 370 individuals in the general population. Lynch syndrome accounts for 2-4% of all colorectal cancer cases.\(^1\)\(^3\)

- Lynch syndrome is associated with a high lifetime risk for colorectal cancer (up to 82%) and endometrial cancer (15-60%), diagnosed at an earlier than usual age. The risk is also increased for small bowel, stomach, ovarian, pancreatic, ureteral and renal pelvis, biliary tract, brain, sebaceous adenoma, and keratoacanthoma tumors.\(^1\)\(^4\)\(^5\)
- Lynch syndrome is caused by mutations in the following mismatch repair genes: MLH1 and MSH2 (together account for 90% of HNPCC mutations), MSH6 (up to 10%), and PMS2 (<5%).\(^4\) An additional gene called EPCAM (or TACSTD1), was found to account for about 1% of Lynch syndrome cases.\(^4\)
- Lynch syndrome gene mutations are inherited in an autosomal dominant manner (children of an affected individual have a 50% risk to inherit a mutation), but family history alone is unreliable for identifying Lynch syndrome cases.\(^1\)\(^4\) Lynch syndrome mutations inherited in an autosomal recessive manner cause Constitutional MMR-Deficiency syndrome (CMMR-D).\(^4\)\(^5\)
• Individuals with colorectal or endometrial cancer due to Lynch syndrome often have abnormal immunohistochemistry (IHC) and/or microsatellite instability (MSI) results on their tumors. These tests have good sensitivity and can identify individuals at sufficient risk for Lynch syndrome to warrant follow-up genetic testing.¹

• Most often, tumor screening is offered to those with cancer and a family history that suggests Lynch syndrome (see guidelines below).¹,⁶,⁷

• Identifying at-risk individuals is necessary for appropriate surveillance and risk reduction.¹

Test information

• Both immunohistochemistry and microsatellite instability evaluate formalin-fixed, paraffin-embedded tumor tissue for evidence of mismatch repair defects. Lynch syndrome is caused by mutations in mismatch repair genes.

  o **Immunohistochemistry (IHC)** detects the presence or absence of MLH1, MSH2, MSH6, ± PMS2 mismatch repair proteins.¹,⁵ Most Lynch syndrome-causing mutations result in protein truncation or absent protein expression⁷, which leads to abnormal IHC staining. As a result, IHC will detect an estimated 83%-94% of underlying Lynch syndrome mutations in colorectal tumors.²,⁸ IHC has the distinct benefit of identifying the gene most likely to have a mutation.⁴,⁸ DNA testing can then be targeted to that specific gene.

  o **Microsatellite Instability (MSI)** compares normal and tumor tissue to detect microsatellite (stretches of repetitive DNA) size changes. Lynch syndrome mutations often cause the size of microsatellites to be unstable.³ When tumor tissue shows high microsatellite instability (MSI-H), it is indirect evidence of an underlying Lynch syndrome gene mutation. Depending on the panel of MSI markers, 80-91% of MLH1 and MSH2 mutations and 55-77% of MSH6 and PMS2 mutations will be detected by MSI testing.²

• No specific tumor screening strategy has been recommended, but studies suggest that both MSI and IHC are cost-effective.¹,²

• MSI and IHC together have better sensitivity for Lynch syndrome than either test alone⁴, and may be used simultaneously or sequentially.

Guidelines and evidence

• The National Comprehensive Cancer Network (NCCN, 2016) has published practice guidelines that address MSI and IHC tumor screening for Lynch syndrome:¹

  o Routine tumor testing for Lynch syndrome is supported either for all CRC patients or CRC patients diagnosed at < 70 years and also those ≥70 years who meet the Bethesda guidelines.
o “IHC and/or MSI screening of all colorectal and endometrial cancers (usually from surgical resection but may be performed on biopsies) regardless of age at diagnosis or family history, has been implemented at some centers to identify individuals at risk for Lynch syndrome. This approach was recently endorsed for colorectal cancer by the Evaluation of Genomic Applications in Practice and Prevention Working Group from the CDC and shown to be cost effective.”

o “An alternative approach is to test all patients with CRC diagnosed prior to age 70 years plus patients diagnosed at older ages who meet the Bethesda guidelines.”

“This approach gave a sensitivity of 95.1% (95%CI, 89.8-99.0%) and a specificity of 95.5% (95%CI, 94.7-96.1%). This level of sensitivity was better than that of both the revised Bethesda and Jerusalem (testing all patients diagnosed with CRC at age <70) recommendations. While this new selective strategy failed to identify 4.9% of Lynch syndrome cases, it resulted in approximately 35% fewer tumors undergoing MMR testing.”

o “Endometrial cancer <50 y is not included in the revised Bethesda guidelines; however, recent evidence suggests that these individuals should be evaluated for Lynch syndrome.”

• Consider Lynch syndrome tumor screening if any one of the following are met:9

  o Colorectal cancer diagnosed before age 50
  o Presence of synchronous or metachronous colorectal cancer, or colorectal cancer with other Lynch syndrome-associated tumors,** regardless of age
  o Microsatellite unstable (MSI-H) tumor pathology before age 60 (e.g., tumor-infiltrating lymphocytes, Crohn’s-like lymphocytic reaction, mucinous/signet-ring differentiation, medullary growth pattern, or other reported features)
  o Colorectal cancer diagnosed in a patient with at least one first-degree relative (parent, sibling, child) with a Lynch syndrome-related tumor*, one of whom was diagnosed before age 50
  o Colorectal cancer diagnosed in a patient with at least two first- or second-degree relatives with Lynch syndrome-related tumors * at any age

**Lynch syndrome-associated tumors include colorectal, endometrial, small bowel, stomach, ovarian, pancreatic, ureteral and renal pelvis, biliary tract, brain tumors (usually glioblastomas associated with Turcot syndrome variant), sebaceous adenomas, and keratoacanthomas (associated with Muir-Torre syndrome variant).

• An evidence-based recommendation from the Centers for Disease Control and Prevention sponsored Evaluation of Genomic Applications in Practice and Prevention Working Group (EGAPP, 2009) found sufficient evidence to recommend Lynch syndrome tumor screening to all individuals with newly diagnosed colorectal cancer since morbidity and mortality can be significantly improved for the patient and at-risk relatives through management changes once Lynch syndrome is
diagnosed.² Although not yet standard of care, some centers have instituted screening for all newly diagnosed colorectal and endometrial cancer.¹

- A National Society of Genetic Counselors and the Collaborative Group of the Americas on Inherited Colorectal Cancer (2012) Joint Practice Guideline makes the following recommendations:¹⁰
  - “Microsatellite instability (MSI) and immunohistochemistry (IHC) tumor analyses should be performed on CRC or endometrial cancers as the first-line testing strategy for any patient being evaluated for Lynch syndrome (this includes individuals with CRC or endometrial cancer who meet Amsterdam I or II criteria or Bethesda guidelines).”
  - “MSI testing should include, at a minimum, the five markers included in the NCI panel.”
  - “MSI and IHC should be performed on pretreated specimens.”
  - “MSI and IHC can be technically challenging assays and should be performed in laboratories that have experience with these tests to minimize the possibility of false positive or false negative results.”
  - “MSI and IHC should be performed, when possible, on an affected relative’s tumor when an unaffected patient is being evaluated for Lynch syndrome.”
  - “Direct germline genetic testing (refers to both DNA sequencing and a technology that detects large rearrangements, insertions, deletions and duplications) may be considered on an affected or unaffected patient being evaluated for Lynch syndrome when MSI and IHC testing are not feasible.”
  - This guideline also notes that “Approximately 25% of individuals with Lynch syndrome are not going to meet Amsterdam or Bethesda criteria so limiting MSI and IHC to individuals who meet these criteria only is inadequate and will miss a large number of individuals with Lynch syndrome.”

- The Multi-Society Task Force (2014)⁷ recently published a consensus statement on genetic evaluation for Lynch syndrome and recommended that “Testing for MMR deficiency of newly diagnosed CRC should be performed. This can be done for all CRCs, or CRC diagnosed at age 70 years or younger, and in individuals older than 70 years who have a family history concerning for LS. Analysis can be done by IHC testing for the MLH1 / MSH2 / MSH6 / PMS2 proteins and / or testing for MSI. Tumors that demonstrate loss of MLH1 should undergo BRAF testing or analysis of MLH1 promoter hypermethylation.” The Multi-Society Task Force on Colorectal Cancer additional endorsed utilizing The Colorectal Cancer Risk Assessment Tool to aid in identifying individuals with possible Lynch syndrome.⁷,¹¹
  - The Multi-Society Task Force is composed of gastroenterology specialists with a special interest in CRC, representing the following major gastroenterology professional organizations: American College of Gastroenterology, American Gastroenterological Association Institute, and the American Society for Gastrointestinal Endoscopy. Also, experts on LS from academia and private
practice were invited authors of this guideline. Representatives of the Collaborative Group of the Americas on Inherited Colorectal Cancer and the American Society of Colon and Rectal Surgeons also reviewed this manuscript. In addition to the Task Force and invited experts, the practice committees and Governing Boards of the American Gastroenterological Association Institute, American College of Gastroenterology, American Society for Gastrointestinal Endoscopy reviewed and approved this document.

- The American Gastroenterology Association (AGA; 2015) recommends “testing the tumors of all patients with colorectal cancer with either immunohistochemistry (IHC) or for microsatellite instability (MSI) to identify potential cases of Lynch syndrome versus doing no testing for Lynch syndrome.”

- The American College of Gastroenterology (ACG; 2015) states that “All newly diagnosed colorectal cancers (CRCs) should be evaluated for mismatch repair deficiency. Analysis may be done by immunohistochemical testing for the MLH1/MSH2/MSH6/PMS2 proteins and/or testing for microsatellite instability (MSI). Tumors that demonstrate loss of MLH1 should undergo BRAF testing or analysis for MLH1 promoter hypermethylation.”

- The Society of Gynecologic Oncology recommends “all women who are diagnosed with endometrial cancer should undergo systematic clinical screening for Lynch syndrome (review of personal and family history) and/or molecular screening. Molecular screening of endometrial cancer for Lynch syndrome is the preferred strategy when resources are available.” Universal molecular tumor testing for either all endometrial cancer or cancers diagnosed at age less than 60, regardless of personal or family cancer history, is a sensitive strategy for identifying women with Lynch syndrome.

- The US Food and Drug Administration (FDA) has approved “Keytruda for the treatment of adult and pediatric patients with unresectable or metastatic solid tumors that have high microsatellite instability (MSI-H) or mismatch repair deficiency (dMMR). This indication covers patients with solid tumors that have progressed following prior treatment and who have no satisfactory alternative treatment options and patients with colorectal cancer that has progressed following treatment with certain chemotherapy drugs.”

Criteria

Lynch Syndrome tumor screening may be considered for individuals with Lynch syndrome-related cancer according to the revised Bethesda criteria and guidelines from the National Comprehensive Care Network (NCCN).

- Testing may be considered for individuals who meet ANY of the following criteria:
  - All colorectal cancers regardless of age, OR
  - All endometrial cancers regardless of age, OR
- Treatment with Keytruda is being considered, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

“Lynch syndrome-related cancers include colorectal, endometrial, gastric, ovarian, pancreas, ureter and renal pelvis, brain (usually glioblastoma as seen in Turcot syndrome), and small intestinal cancers, as well as sebaceous gland adenomas and keratoacanthomas in Muir-Torre syndrome. However, “tumor testing strategies apply to colorectal and endometrial cancers. Limited data exist regarding the efficacy of tumor testing in other LS tumors.”\(^1\)

References


14. US Food and Drug Administration. FDA approves first cancer treatment for any solid tumor with a specific genetic feature. Available at: https://www.fda.gov/newsevents/newsroom/pressannouncements/ucm560167.htm
Lynch Syndrome Tumor Screening - Second-Tier

MOL.TS.199.A
v2.0.2018

Procedures addressed

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<td>81210</td>
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<tr>
<td>MLH1 Promoter Methylation Analysis</td>
<td>81288</td>
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What are BRAF mutation and MLH1 promoter methylation testing for Lynch Syndrome

Introduction

Lynch syndrome, also called hereditary non-polyposis colorectal cancer (HNPCC), is the most common known hereditary cause of colon and endometrial cancer. It affects approximately 1 in 35 colorectal and endometrial cancer patients and around 1 in 370 individuals in the general population. Lynch syndrome accounts for 2-4% of all colorectal cancer cases.¹-⁴

- Lynch Syndrome is associated with a high lifetime risk for colorectal cancer (up to 82%) and endometrial cancer (15-60%), diagnosed at an earlier than usual age. The risk is also increased for small bowel, stomach, ovarian, pancreatic, ureteral and renal pelvis, biliary tract, brain, sebaceous adenoma, and keratoacanthoma tumors.¹,⁵⁻⁶ Lynch syndrome is an autosomal dominant syndrome that is associated with a germline mutation in one of at least five genes: MLH1, MSH2, MSH6, PMS2, and EPCAM. Children of an affected individual have a 50% risk to inherit a mutation.⁵
- People suspected to have colorectal or endometrial cancer caused by Lynch syndrome generally have tumor screening studies first.¹,⁷⁻⁸ Tumors caused by Lynch syndrome often show microsatellite instability (MSI) and absent protein from one or more mismatch repair genes (MLH1, MSH2, MSH6, +/- PMS2) by immunohistochemistry (IHC).¹,⁵
- If MSI or IHC shows signs of Lynch syndrome, the next step is usually Lynch syndrome genetic testing.¹,²,⁵
- However, another step may be useful before genetic testing when IHC indicates absent MLH1 protein. Absent MLH1 may be caused by Lynch syndrome, but is also
frequently a sporadic finding in colorectal and endometrial cancers. Additional testing can help determine whether MLH1-negative colorectal and endometrial tumors (not other Lynch syndrome-associated tumors) are sporadic or are associated with Lynch syndrome.¹²⁵

- The most common cause of absent MLH1 protein is sporadic methylation of the MLH1 gene, which causes the gene to make no protein.³
- This MLH1 methylation is often associated with a sporadic mutation in the BRAF gene (in colorectal tumors only; not endometrial).
- BRAF is part of a cell signaling pathway that helps control cell growth. About 6-8% of colorectal cancer tumors have a BRAF mutation.⁹ A single mutation, called V600E (previously called V599E), accounts for about 90% of these BRAF mutations.³
- When MLH1 protein is absent and a BRAF mutation is present, the colorectal cancer is rarely caused by Lynch syndrome (i.e., the cancer is usually sporadic).³
- When MLH1 protein is absent, the tumor is negative for a BRAF V600E mutation, and MLH1 promoter methylation is present, the cancer is still generally sporadic. However, other types of mutations (e.g., MLH1 epimutations that cause widespread hypermethylation or MLH1 promoter variants) may cause this result.¹²
- BRAF gene mutations that are inherited or occur in tumors are relevant to several other diagnoses, including:
  - Colorectal Cancer Anti-EGFR Therapy Response
  - Thyroid Cancer Prognosis
  - Noonan Syndrome

Test information

- For Lynch syndrome-related testing, BRAF mutation analysis +/- MLH1 promoter methylation studies are done on colorectal tumor tissue. MLH1 promoter methylation studies (not BRAF) are done on endometrial tumor tissue. Sporadic BRAF mutations do not appear to be responsible for MLH1 methylation in endometrial tumors.²
- When BRAF is being tested because MLH1 protein was absent on colorectal tumor IHC, most laboratories test only for the BRAF V600E mutation. However, some laboratories sequence all or part of the BRAF gene (sometimes for reasons other than Lynch syndrome screening). Targeted mutation analysis is generally less expensive than gene sequencing. Because the V600E accounts for most BRAF colorectal cancer mutations, targeted mutation analysis for this one mutation is sufficient. Results of testing for this single mutation are expected to be reliable.³
• BRAF mutation analysis and MLH1 promoter methylation studies may be offered as panels or in reflex options. For instance, BRAF mutation analysis may be a reflex test when MLH1 IHC results are abnormal. MLH1 promoter methylation studies may be done as reflex test if BRAF mutation analysis is negative.

Guidelines and evidence
The following organizations address when BRAF and/or MLH1 promoter methylation studies should be employed in evaluating the likelihood a tumor is caused by Lynch syndrome. This section does not address who should have MSI and/or IHC tumor screening for Lynch syndrome at the time of cancer diagnosis.

• The National Comprehensive Cancer Network (NCCN, 2016) includes BRAF V600E mutation and MLH1 promoter methylation status in their table that outlines “tumor testing results and additional testing strategies.”

• For colorectal tumors that show no MLH1 protein by IHC (+/- PMS2 negative), they state “consider BRAF/methylation studies.”

• They recommend the following based on the BRAF results:

<table>
<thead>
<tr>
<th>BRAF V600E Mutation</th>
<th>MLH1 Promoter Methylation</th>
<th>Lynch Syndrome Genetic Testing?</th>
</tr>
</thead>
<tbody>
<tr>
<td>Positive</td>
<td>Not necessary</td>
<td>No</td>
</tr>
<tr>
<td>Negative</td>
<td>Positive</td>
<td>Most likely a sporadic cancer; genetic testing only if the family history is compelling.</td>
</tr>
<tr>
<td>Negative</td>
<td>Negative</td>
<td>Pursue MLH1 and/or PMS2 genetic testing**.</td>
</tr>
</tbody>
</table>

Note  ** If genetic testing is negative, consider somatic MMR genetic testing.¹

• “Growing evidence suggests that the majority of these individuals with abnormal tumor results and no germline mutation found have double somatic mutations/changes in the mismatch repair (MMR) genes. Although the efficacy has not yet been proven, genetic testing of the corresponding gene(s) could be performed on tumor DNA to assess for somatic mutations. Individuals found to have double somatic mutations/changes in the mismatch repair (MMR) genes likely do not have LS and management should be based on personal/family history.”¹

• The National Society of Genetic Counselors and the Collaborative Group of the Americas on Inherited Colorectal Cancer (NSGC/CGA-ICC, jointly published, 2012) guidelines state:²
  o “Both somatic hypermethylation of the MLH1 gene (an epigenetic change) and somatic mutations of the BRAF gene have been described in sporadic CRCs exhibiting MSI and/or loss of expression of MLH1. These somatic events are rarely seen in LS CRCs and therefore may be useful in determining whether a MSI-high CRC is more likely to be sporadic.”
“MLH1 promoter methylation and BRAF V600E mutation testing may help to reduce the number of germline genetic tests needed when IHC reveals absence of MLH1 and PMS2. However, NSGC and the CGAICC did not find enough data to recommend one test over the other or both concomitantly.”

The likelihood of identifying a germline MLH1 with both DNA sequencing and MLPA analysis is approximately 33% when MLH1 +/- PMS2 are absent on IHC and MLH1 promoter hypermethylation is not present.

- The Centers for Disease Control and Prevention sponsored Evaluation of Genomic Applications in Practice and Prevention Working Group (EGAPP, 2009) published evidence-based recommendations focused on Lynch syndrome tumor screening by MSI and IHC. They include some information about BRAF mutation analysis and MLH1 promoter methylation, but do not make formal recommendations regarding these two tests. However, the CDC website provides additional information about these guidelines. For BRAF V600E mutation analysis, they find adequate evidence of clinical validity and utility with an overall recommendation of “Sufficient evidence to recommend use for the benefit of relatives.”

- The CDC website does not address MLH1 promoter methylation, but an EGAPP supplemental evidence review (that accompanied the recommendations) states: “This supplemental evidence review did not involve a formal search or statistical summary concerning the literature on methylation testing. The literature suggests, however, that BRAF V600E mutation testing and methylation testing of the MLH1 promoter region among CRC cases with absent MLH1 protein might avoid similar numbers of sequencing tests with little loss in Lynch syndrome detection.”

- The Multi-Society Task Force (2014) recently published a consensus statement on genetic evaluation for Lynch syndrome and recommended that “Testing for MMR deficiency of newly diagnosed CRC should be performed. This can be done for all CRCs, or CRC diagnosed at age 70 years or younger, and in individuals older than 70 years who have a family history concerning for LS. Analysis can be done by IHC testing for the MLH1 / MSH2 / MSH6 / PMS2 proteins and/or testing for MSI. Tumors that demonstrate loss of MLH1 should undergo BRAF testing or analysis of MLH1 promoter hypermethylation.”

- The American Gastroenterology Association (AGA; 2015) suggests “that in patients with colorectal cancer with IHC absent for MLH1, second-stage tumor testing for a BRAF mutation or for hypermethylation of the MLH1 promoter should be performed rather than proceeding directly to germline genetic testing.”

- The American College of Gastroenterology (ACG; 2015) states that “All newly diagnosed colorectal cancers (CRCs) should be evaluated for mismatch repair deficiency. Analysis may be done by immunohistochemical testing for the MLH1/MSH2/MSH6/PMS2 proteins and/or testing for microsatellite instability (MSI). Tumors that demonstrate loss of MLH1 should undergo BRAF testing or analysis for MLH1 promoter hypermethylation.”
The Society of Gynecologic Oncology recommends "all women who are diagnosed with endometrial cancer should undergo systematic clinical screening for Lynch syndrome (review of personal and family history) and/or molecular screening. Molecular screening of endometrial cancer for Lynch syndrome is the preferred strategy when resources are available." Universal molecular tumor testing for either all endometrial cancer or cancers diagnosed at age less than 60, regardless of personal or family cancer history, is a sensitive strategy for identifying women with Lynch syndrome.13

***The Multi-Society Task Force is composed of gastroenterology specialists with a special interest in CRC, representing the following major gastroenterology professional organizations: American College of Gastroenterology, American Gastroenterological Association Institute, and the American Society for Gastrointestinal Endoscopy. Also, experts on LS from academia and private practice were invited authors of this guideline. Representatives of the Collaborative Group of the Americas on Inherited Colorectal Cancer and the American Society of Colon and Rectal Surgeons also reviewed this manuscript. In addition to the Task Force and invited experts, the practice committees and Governing Boards of the American Gastroenterological Association Institute, American College of Gastroenterology, American Society for Gastrointestinal Endoscopy reviewed and approved this document.

Criteria

BRAF V600E Mutation Analysis or MLH1 Promoter Methylation Status

- Previous Testing:
  - IHC testing has been performed and indicates a loss of MLH1 protein, AND
- Diagnostic Testing for Symptomatic Individuals:
  - Personal history of colorectal or endometrial**** cancer, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy

**** MLH1 methylation only

References


Mammaprint 70-Gene Breast Cancer Recurrence Assay

Procedure addressed

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<tbody>
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<td>Mammaprint 70 Gene Signature</td>
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What is Mammaprint

Definition

Mammaprint® is a 70-gene expression test designed to predict the chance of later-in-life recurrence of breast cancer in women with newly diagnosed, early stage breast cancer.1 It is FDA cleared for use along with other standard prognostic methods, such as disease staging, grading and other tumor marker analyses.2

- Mammaprint is intended to assist patients and providers considering treatment with adjuvant chemotherapy. Patients assigned a "low risk" may choose hormone therapy (tamoxifen) alone and forego chemotherapy. Patients assigned a "high risk" may benefit from more aggressive treatment and choose to do chemotherapy.1
- Mammaprint is designed for women with breast cancer who have:1,2
  - Stage I or II invasive carcinoma
  - Tumor size <5.0 cm
  - Node-negative (no metastasis to lymph nodes)
  - Estrogen receptor-positive (ER+) or -negative (ER-) disease

Test information

- Mammaprint uses a microarray platform to analyze the expression level of 70 genes in the tumor. These 70 genes are thought to be critical in the cellular pathways to cancer metastasis.1
• Based on the test results, patients are assigned either a low risk or a high risk for a distant recurrence. Low risk corresponds to a 10% risk of recurrence by 10 years without any additional adjuvant treatment. In contrast, those in the high risk group have a 29% risk of recurrence by 10 years without any additional adjuvant treatment.¹

Guidelines and evidence
• The National Comprehensive Cancer Network (NCCN) 2016 Clinical Practice Guidelines for Breast Cancer state that:³
  o “The NCCN Panel members acknowledge that many assays, including PAM50 and MammaPrint, have been clinically validated for prediction of prognosis. However, based on the currently available data, the panel believes that the 21-gene assay has been best-validated for its use as a prognostic test as well as in predicting who is most likely to respond to systemic chemotherapy.”

• Evidence-based clinical guidelines from the American Society of Clinical Oncology (ASCO, 2016) state the following:⁴
  o “If a patient has ER/PgR-positive, HER2-negative (node-positive or node-negative) breast cancer, the clinician should not use the 70-gene assay (MammaPrint; Agendia, Irvine, CA) to guide decisions on adjuvant systemic chemotherapy. Type: evidence based. Evidence quality: intermediate. Strength of recommendation: moderate.”
  o “If a patient has HER2-positive breast cancer, the clinician should not use the 70-gene assay (MammaPrint) to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: low. Strength of recommendation: moderate.”
  o “If a patient has TN breast cancer, the clinician should not use the 70-gene assay (MammaPrint) to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: insufficient. Strength of recommendation: strong.”

• European Society of Medical Oncology (ESMO) 2015:⁵
  o “Gene expression profiles, such as MammaPrint (Agendia, Amsterdam, the Netherlands), Oncotype DX Recurrence Score (Genomic Health, Redwood City, CA), Prosigna (Nanostring Technologies, Seattle, WA) and EndoPredict (Myriad Genetics), may be used to gain additional prognostic and/or predictive information to complement pathology assessment and to predict the benefit of adjuvant chemotherapy. The three latter tests are designed for patients with ER-positive early breast cancer only.”
  o “In cases of uncertainty regarding indications for adjuvant chemotherapy (after consideration of other tests), gene expression assays, such as MammaPrint, Oncotype DX, Prosigna and Endopredict, may be used, were available.”
o “In cases when decisions might be challenging, such as luminal B HER2-negative and node-negative breast cancer, commercially available molecular signatures for ER-positive breast cancer, such Oncotype DX, EndoPredict, Prosigna, and for all types of breast cancer (pN0–1), such as MammaPrint and Genomic Grade Index, may be used in conjunction with all clinicopathological factors, to help in treatment decision making.”

  o “The Panel considered the role of multi-parameter molecular marker assays for prognosis separately in years 1-5 and beyond 5 years, and their value in selecting patients who require chemotherapy. Oncotype DX®, MammaPrint®, PAM-50 ROR® score, EndoPredict® and the Breast Cancer Index® were all considered usefully prognostic for years 1-5. Beyond 5 years, the Panel was divided almost equally on the prognostic value of Oncotype DX (despite the available data from NSABP Trial B-14 [32]); EndoPredict® (despite the report of Dubsky et al. [36]); and Breast Cancer Index (despite the report of Zhang et al. [37]). (All these reports show the respective tests to be prognostic beyond 5 years.) PAM50 ROR® score was agreed to be clearly prognostic beyond 5 years, and a clear majority rejected the prognostic value of MammaPrint® in this time period.”

• The Evaluation of Genomic Applications in Practice and Prevention (EGAPP, 2009) Working Group reviewed the evidence for MammaPrint and concludes:⁷
  o “It is unclear what population of patients would derive benefit from use of the test, and what the magnitude of that benefit would be. Prospective data from trials like MINDACT will be extremely valuable.”
  o “Overall, published evidence supports MammaPrint as a better predictor of the risk of distant recurrence than traditionally used tumor characteristics or algorithms, but its performance in therapeutically homogeneous populations is not yet known with precision, and it is unclear for how many women the lowest predicted risks are low enough to forgo chemotherapy.”
  o “No evidence is available to permit conclusions regarding the clinical utility of MammaPrint to select women who will benefit from chemotherapy.”
  o “To conclude, the literature on the 70-gene signature includes numerous studies that focused more on its biological underpinning and less on the clinical implications of this gene expression profile, although it has now received FDA approval for clinical use.”

• The US Food and Drug Administration (FDA) cleared Mammaprint for clinical use on fresh tissue samples in 2007.² The FDA cleared Mammaprint for clinical use on FFPE samples in 2015.¹
• While the clinical validity of the test has been established, data regarding the clinical utility of MammaPrint is still emerging.8-16
  
  o Cardoso et al. (2016)17 conducted the MINDACT trial (randomized, phase III trial), evaluating a total of 1550 early-stage breast cancer patients, who were considered at high clinical risk as assessed by a version of the Adjuvant! Online algorithm, and low genomic risk as assessed by the MammaPrint. Chemotherapy was administered in those patients at low clinical and genomic risk, and avoided in those patients at high clinical and genomic risk. (Patients with low clinical risk were not evaluated in the study). The primary goal was to assess whether a 92% survival rate at 5 years of follow up (lower boundary of the 95% CI) was achievable among patients with high-risk clinical features and a low-risk gene-expression profile who did not receive chemotherapy. Results showed that the rate of survival at 5 years of follow-up was nearly 95% (95% CI, 9.25 to 96.2) among those patients who did not receive chemotherapy. Study authors reported similar rates of survival without distant metastasis in a subgroup of patients who had ER+, HER2-, and either node-negative or node-positive disease. It should be noted that the study was designed to test those patients with high clinical risk with MammaPrint. Patients with low clinical risk were not tested with MammaPrint; however, a subgroup analysis of this population showed that the 5-year recurrence risk may be too high to forego chemotherapy (low clinical risk: hazard ratio [HR], 1.17 vs high clinical risk: HR, 0.78).
  
  o Additional well-designed studies are needed to examine the optimal patient population who could achieve the greatest clinical benefit with MammaPrint testing.

Criteria

• Previous Testing:
  
  o No repeat MammaPrint testing on the same sample when a result was successfully obtained, and
  
  o No previous gene expression assay (e.g. Prosigna, Oncotype DX) performed on the same sample when a result was successfully obtained, AND

• Testing Multiple Samples:
  
  o When more than one breast cancer primary is diagnosed:
    
    ▪ There should be reasonable evidence that the tumors are distinct (e.g., bilateral, different quadrants, different histopathologic features, etc.), and
    
    ▪ There should be no evidence from either tumor that chemotherapy is indicated with or without knowledge of the MammaPrint test result (e.g., histopathologic features or previous MammaPrint result of one tumor suggest chemotherapy is indicated), and
If both tumors are to be tested, both tumors must independently meet the required clinical characteristics outlined below and the MammaPrint score from one tumor must be determined before testing of another tumor.

- Required Clinical Characteristics:
  - Invasive breast cancer meeting all of the following criteria:
    - Nonmetastatic, and
    - Tumor size ≤5cm in greatest dimension, and
  - Patient has no regional lymph node metastasis (pN0) or only micrometastases (pN1mi, malignant cells in regional lymph node(s) not greater than 2.0mm), and
  - Patient has not received Tamoxifen therapy, and
  - Chemotherapy is a treatment option for the patient but has not been initiated, and
  - Results from this MammaPrint test will be used in making chemotherapy treatment decisions, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


Mammostrat Breast Cancer Recurrence Assay

Procedures addressed

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What is the Mammostrat Breast Cancer Recurrence Assay

Definition

The Mammostrat® Breast Cancer Recurrence Assay is an immunohistochemical (IHC) assay that measures levels of five proteins in tumor tissue associated with risk of breast cancer recurrence.¹

- It is used in people with newly diagnosed, early stage breast cancer.
- The assay looks at five proteins and determines their expression levels in the tumor. The expression levels of these five markers are thought to influence whether the tumor will metastasize, increasing the patient’s chance of recurrence. These levels are then translated into a risk index, given as a percent chance of recurrence over 10 years.
- Physicians and patients may use the risk index as one factor in determining the course of treatment. Patients in the high risk category may benefit more from aggressive treatment, whereas patients in the low risk category may elect to forgo the aggressive chemotherapy.²

Test information

- The Mammostrat assay measures the expression level of five proteins by immunohistochemistry. These markers are believed to be associated with breast cancer recurrence.³
  - p53 plays a role in cell cycle regulation. Mutations in the p53 gene are associated with tumor growth.
  - HTF9C is implicated in DNA replication and cell cycle control.
• **CEACAM5** is normally expressed in embryonic tissue, but is also found in some tumors.
• **NDRG1** may have a role in helping tumors survive aggressive treatment.
• **SLC7A5** can, when overexpressed, help sustain the high growth rate of cancer.

- These levels are then translated into a quantitative “risk index” via a proprietary algorithm, which divides patients into groups with low, moderate, or high risk of recurrence:

<table>
<thead>
<tr>
<th>Risk index</th>
<th>Risk of breast cancer recurrence over 10 years</th>
</tr>
</thead>
<tbody>
<tr>
<td>Low</td>
<td>7.6%</td>
</tr>
<tr>
<td>Moderate</td>
<td>16.3%</td>
</tr>
<tr>
<td>High</td>
<td>20.9%</td>
</tr>
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</table>

**Guidelines and evidence**

• The NCCN does not specifically mention the use of Mammostrat in its most recent guidelines.
• The American Society of Clinical Oncology (ASCO, 2016) published a clinical practice guideline on the use of biomarkers to guide decision-making in women with early-stage invasive breast cancer. They recommend:
  - “If a patient has ER/PgR-positive, HER2-negative (node-positive or node-negative) breast cancer, the clinician should not use the five-protein assay (Mammostrat; Clarient, a GE Healthcare company, Aliso Viejo, CA) to guide decisions on adjuvant systemic therapy. Type: evidence based. Evidence quality: intermediate. Strength of recommendation: moderate.”
  - “If a patient has HER2-positive breast cancer or TN breast cancer, the clinician should not use the five-protein assay (Mammostrat) to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: insufficient. Strength of recommendation: strong.”

• A 2010 clinical study tested the assay’s ability to accurately predict risk of breast cancer recurrence in a cohort of 1,812 women with early stage breast cancer:
  - “The Mammostrat markers are biologically independent of one another and measure aspects of physiology distinct from proliferation, HER2 status, and hormone receptor status already assessed by IHC assays that are standard of care. Collectively these data add support to a potential role for Mammostrat in management of early-stage breast cancer.”
Criteria
• This test is considered investigational and/or experimental.
  o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  o In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

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<tr>
<td>TGFBR2 Sequencing</td>
<td>81405</td>
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</tbody>
</table>

What is Marfan syndrome

Definition

Marfan syndrome is an autosomal dominant disorder that affects connective tissue in many parts of the body. It affects about 1 in 5000 to 1 in 10000 individuals.¹

- Symptoms can present in males or females at any age and typically worsen over time. Infants who present with symptoms typically have the most severe disease course.¹
- Signs and symptoms of Marfan syndrome usually include (some combination of the following):¹
  - Cardiovascular system — dilatation of the aorta, predisposition for aortic tear or rupture, mitral valve prolapse (with or without congestive heart failure), tricuspid valve prolapse, and enlargement of the proximal pulmonary artery.¹
  - Skeletal system — long bone overgrowth and joint laxity, long arms and legs, scoliosis, sternum deformity (pectus excavatum or carinatum), pes planus, long thin fingers and toes, micrognathia, retrognathia, high-arched palate, deep set eyes, malar hypoplasia, down- slanting palpebral fissures, and long thin face.¹
• Ocular system — severe myopia, dislocated lens of eye (ectopia lentis), detached retina, glaucoma, early cataracts.¹
• Other symptoms – dural ectasia (stretching of the dural sac), hernias, stretch marks on the skin, and lung bullae.¹

- Clinical diagnosis-Ghent Criteria¹⁻³
  - With no known family history, a Marfan syndrome diagnosis is confirmed if any ONE of the following is met:¹⁻³
    - Significant aortic dilation (Z-score ≥2)/dissection + ectopia lentis**
    - Significant aortic dilation (Z-score ≥2)/dissection + FBN1 mutation
    - Aortic dilation/dissection + sufficient points from other system findings**
    - Ectopia lentis + FBN1 mutation known to be associated with aortic disease
  - With a known family history, the presence of any ONE of the following is diagnostic:¹⁻³
    - Ectopia lentis
    - Significant aortic root enlargement (Z-score ≥2 in those >20 years of age or ≥3 in those <20 years of age)**
    - Sufficient points (>7) from other system findings**

** Marfan syndrome can be clinically diagnosed in these cases, provided there are not other findings that more strongly suggest Sphrintzen-Goldberg syndrome, Loeys-Dietz syndrome, or vascular Ehlers-Danlos syndrome, which have clinical overlap. Or, these conditions are unlikely based on genetic or collagen testing.

- Systemic scoring system¹⁻³
  - Wrist and Thumb Sign - 3 points
  - Wrist or Thumb Sign - 1 point
  - Pectus Carinatum deformity - 2 points
  - Pectus Excavatum or chest asymmetry -1 point
  - Hindfoot deformity - 2 points
  - Plan pes planus -1 point
  - Pneumothorax - 2 points
  - Dural Ectasia - 2 points
  - Protrusio Acetabulae - 2 points
  - Reduced upper seg/lower seg and inc. arm span and height ratio - 1 point
  - Scoliosis or thoracolumbar kyphosis - 1 point
- Reduced elbow extension - 1 point
- 3 of 5 facial features: Dolichocephaly, enophthalmos, downslanting palpebral fissures, malar hypoplasia, retrognathia - 1 point
- Skin striae - 1 point
- Myopia - 1 point
- Mitral Valve Prolapse - 1 point

  o According to the Ghent criteria, many of the manifestations of Marfan syndrome can emerge with age. Therefore, it is not advisable to establish definitive alternative diagnosis in individuals younger than age 20 years who have some physical manifestations of Marfan syndrome but not enough for a clinical diagnosis. In this circumstance, the following is suggested:\textsuperscript{1,2}

  “If the systemic score is <7 and/or borderline aortic root measurements (Z-score <3) are present (without an FBN1 pathogenic variant), use of the term “nonspecific connective tissue disorder” is suggested until follow-up echocardiographic evaluation shows aortic root dilation (Z-score ≥3).”\textsuperscript{1,2}

  “If an FBN1 pathogenic variant is identified in simplex or familial cases but aortic root Z-score is below 3.0, the term “potential Marfan syndrome” should be used until the aorta reaches this threshold.”\textsuperscript{1,2}

• Diagnostic evaluations recommended:
  o Ophthalmologist evaluation with someone familiar with Marfan\textsuperscript{1}
  o Evaluation for skeletal manifestations by an orthopedist\textsuperscript{1}
  o Cardiovascular evaluations\textsuperscript{1}
  o Medical genetics evaluation\textsuperscript{1}

Genetics

• Marfan syndrome is caused by mutations in the FBN1 gene, located on chromosome 15.\textsuperscript{1,4}

• Marfan syndrome is inherited in an autosomal dominant fashion. Everyone has 2 copies of the FBN1 gene. If one of these genes has a mutation, it is enough to cause Marfan syndrome. It affects males and females equally.\textsuperscript{1}

• A person who is found to have a FBN1 mutation has a 50% chance to pass the mutation to his/her children. Prenatal testing is available when the FBN1 mutation in the family is known.

• Genetic testing for Marfan syndrome typically starts with sequencing of the FBN1 gene. If negative, deletion/duplication of FBN1 should be considered.\textsuperscript{1}

• Mutations in the TGFBR1 or TGFBR2 gene have been found in some individuals with a clinical suspicion of MFS and no identifiable FBN1 mutation.\textsuperscript{1,3}
TGFBR1/2 are associated with Loeys-Dietz syndrome (LDS). Some features of MFS and LDS overlap. However, people with LDS typically have a greater risk of frequent aortic dissection and rupture at smaller dimensions and in early childhood.¹

- The presence of a mutation in the FBN1 gene alone does not diagnose Marfan syndrome. FBN1 mutations may cause conditions other than Marfan syndrome. Conversely, some people who meet the clinical diagnostic criteria for Marfan syndrome do not have an identifiable FBN1 mutation.¹

- Approximately 25% of cases of Marfan syndrome are the result of a new genetic change (de novo mutation) in the affected person and are not inherited from a carrier parent.¹

**Test information**

- More than 1300 mutations have been found in the FBN1 gene that cause Marfan syndrome.⁴
- **FBN1 Sequencing** identifies an FBN1 gene mutation in approximately 70-93% of people with a clinical diagnosis of Marfan syndrome.¹
- **FBN1 Deletion/Duplication Analysis** can be performed to look for other types of gene mutations when sequencing is negative. The percentage of people with a clinical diagnosis of Marfan syndrome and a deletion/duplication mutation is unknown.¹
- **FBN1 Known Familial Mutation.** If a FBN1 mutation is found in an affected person, other family members may be offered testing.¹,⁵
- **Additional Testing Information**
  - **TGFBR1/2 Testing.** If a mutation is not found in FBN1 and there is a strong clinical suspicion of Marfan syndrome, TGFBR1/2 genetic testing may be indicated. Given the increased risk of aortic dissection and rupture at smaller dimensions and in early childhood in LDS,¹ it is important to confirm whether there is a mutation in one of these two genes.
Panel Testing. There are other conditions which can cause familial aortic aneurysm and dissections and/or have overlapping features with Marfan syndrome. Many laboratories offer panel testing for FBN1 as well as other genes that cause these conditions. Detection rates of expanded panels vary by laboratory and depend on the genes included and the methods used for testing. A thorough clinical evaluation along with appropriate imaging studies will point to a specific diagnosis in many cases. Testing for conditions that are clinically indicated is most appropriate. Testing multiple genes, without supporting clinical features, has the potential to yield results that are difficult to interpret. The chance that a variant of uncertain significance will be found increases as more genes are tested. According to the American College of Medical Genetics and Genomics, “There is no case of classic, bona fide MFS due to mutations in a gene other than FBN1.” Therefore, when there is a strong clinical suspicion for Marfan syndrome, genetic testing for genes other than FBN1 is typically not needed, with the exception of TGFBR1/2 testing.

Guidelines and evidence

- The European Society of Cardiology (ESC, 2014) stated the following:7
  - “Once a familial form of TAAD is highly suspected, it is recommended to refer the patient to a geneticist for family investigation and molecular testing.” (Class I, Level C)

- The Canadian Cardiovascular Society (2014) stated the following:8
  - “We recommend clinical and genetic screening for suspected Marfan syndrome to clarify the nature of the disease and provide a basis for individual counseling" (Strong recommendation, High quality evidence)
  - “We recommend that genetic counseling and testing be offered to first degree relatives of patients in whom the causal mutation of a TAD-associated gene is identified. We recommend that aortic imaging be offered only to mutation carriers." (Strong recommendation, low quality evidence)

- Joint evidence-based guidelines from ACCF/AHA/AATS/ACR/ASA/SCA/SCAI/SIR/STS/SVM (2010) for the diagnosis and management of thoracic aortic disease include Marfan syndrome. Genetic testing for Marfan syndrome is addressed in the following guidelines statements:
  - “If the mutant gene (FBN1, TGFBR1, TGFBR2, COL3A1, ACTA2, MYH11) associated with aortic aneurysm and/or dissection is identified in a patient, first-degree relatives should undergo counseling and testing. Then, only the relatives with the genetic mutation should undergo aortic imaging.”5 [Class 1, Level of Evidence C. Recommendation that procedure or treatment is useful/effective. It is based on very limited populations evaluated and only expert opinion, case studies or standard of care.]5
  - “The criteria for Marfan syndrome is based primarily on clinical findings in the various organ systems affected in the Marfan syndrome, along with family history and FBN1 mutations status.”5
o Recommend echo at baseline, repeat at 6 months to look for progression then yearly if stable (Class 1).

o Determining genetic etiology guides prophylactic aortic surgery.

- An international group of Marfan syndrome experts initially proposed clinical diagnostic criteria for Marfan syndrome in 1996, called the Ghent nosology that gained wide acceptance.\(^9\)

- The Ghent criteria were updated in 2010 and now address the role of FBN1 genetic testing in the diagnosis of Marfan syndrome.\(^2\) They do not include guidelines about when to test for a familial mutation, but do indicate that finding a familial mutation is not sufficient evidence alone to make a definitive diagnosis, stating: “If an FBN1 mutation is identified in sporadic or familial cases but aortic root measurements are still below Z=3, we propose to use the term ‘potential MFS’ [Marfan syndrome] until the aorta reaches threshold.”

- According to the American College of Medical Genetics and Genomics, “There is no case of classic, bona fide MFS due to mutations in a gene other than FBN1. However, current clinical molecular testing of FBN1 successfully detects mutations in such unequivocal patients in only about 90-95% of cases. For all of these reasons, searching for mutations in FBN1 continues to have a circumscribed role in the diagnosis of equivocal cases. Said differently, MFS remains, by and large, a clinical diagnosis.”\(^6\)

Criteria

**FBN1 Known Familial Mutation Analysis**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous genetic testing of FBN1, and
  - FBN1 mutation identified in 1\textsuperscript{st} degree biological relative, OR

- Prenatal Testing for At-Risk Pregnancies:
  - FBN1 mutation identified in a previous child or either parent

**FBN1 Sequencing**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Testing:
  o No previous FBN1 sequencing, and
  o No known FBN1 mutation in the family, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Genetic testing is necessary because there is uncertainty in the clinical diagnosis, and
    ▪ Aortic root enlargement (Z-score >2.0) and a systemic score <7, without ectopia lentis, or
    ▪ Ectopia lentis, or
  o An individual has a clinical diagnosis of Marfan syndrome based on the revised Ghent Criteria, and
    ▪ Genetic testing is needed in order to offer testing to family members, or
    ▪ Genetic testing is needed for prenatal diagnosis purposes.

FBN1 Deletion/Duplication Analysis

• Criteria for FBN1 Sequencing are met, AND
• No previous deletion/duplication analysis of FBN1, AND
• No mutations detected in full sequencing of FBN1

TGFBR1/2 Known Familial Mutation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous genetic testing of TGFBR1/2, and
  o TGFBR1/2 mutation identified in 1st degree biological relative

TGFBR2 Sequencing

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
• Diagnosed with Marfan syndrome based on the Ghent criteria (Member met testing guidelines for FBN1 sequencing)
  
• Diagnostic Testing for Symptomatic Individuals:
  
  o There is a strong clinical suspicion of MFS based on the Ghent criteria (Member met testing guidelines for FBN1 sequencing)

TGFBR1 Sequencing

• Genetic Counseling:
  
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  
  o No previous TGFBR1 testing performed, and
  
  o No mutations detected in full sequencing or deletion/duplication analysis of FBN1, and
  
  o No mutations detected in full sequencing of TGFBR2, AND

• Diagnostic Testing for Symptomatic Individuals:
  
  o There is a strong clinical suspicion of MFS based on the Ghent criteria (Member met testing guidelines for FBN1 sequencing)

References


# Maturity-Onset Diabetes of the Young (MODY) Testing

## Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan’s procedure code list for management requirements.

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What is MODY

Definition

Maturity-onset diabetes of the young (MODY) is a type of monogenic diabetes characterized by non-insulin-dependent diabetes and early onset (usually before age 25).\textsuperscript{2-4}

- Diabetes is a disorder that results in elevated blood glucose. Over time, the disorder can cause various health problems, including diseases of the heart, kidneys, eyes, and nervous system. The condition affects 29.1 million people in the United States, or 9.3% of the population.\textsuperscript{1}
- The most common types of diabetes are type 1 and type 2. The genetic basis of these type of diabetes is largely unknown. The disease are thought to be the result of a combination of multiple genetic and environmental risk factors.\textsuperscript{1}
  - Risk factors for type 2 diabetes include being 45 or older, having a family history of diabetes, being overweight, and having certain health problems such as hypertension.
- Monogenic forms of diabetes are caused by a mutation in a single gene. They are rare, accounting for approximately 2% of all diabetes cases.\textsuperscript{2-4}
- Diabetes evaluations may include assessment of pancreatic autoantibodies, plasma glucose levels, hemoglobin A1C assessment (HbA1C), and oral glucose tolerance testing (OGTT). For young individuals in whom a diagnosis of type 1 or type 2 diabetes is considered unlikely, genetic testing for monogenic diabetes may be considered, especially in the presence of a strong family history.\textsuperscript{1}
- MODY is inherited in an autosomal dominant manner. When a parent has a MODY mutation, each of her/his offspring have a 50% risk of inheriting the mutation.\textsuperscript{2-4}
- There are 13 known MODY genes, and three account for the majority of cases:\textsuperscript{2-4}
  - MODY3: Mutations in the hepatocyte nuclear factor-1 alpha (HNF1A) gene are the most common cause of MODY, accounting for about half of cases. This type is characterized by a progressive insulin secretory defect due to beta-cell failure. Laboratory evaluations are negative for pancreatic islet cell antibodies (ruling out type 1) and glycosuria is detectable even at low blood glucose levels (<10 mmol/l). Treatment of choice for people with this type of MODY is sulfonylureas, and a majority of patients can be transferred from insulin to oral agents.
  - MODY2: Mutations in the glucokinase gene (GCK) are the next most common cause of MODY, accounting for about 20-25% of cases. GCK encodes the glucokinase enzyme, which acts as the pancreatic glucose sensor. Mutations result in lifelong, stable, mild fasting hyperglycemia. HbA1C values are usually just above the high normal range. People with GCK mutations rarely require treatment. This type of MODY may be detected during pregnancy, when glucose tolerance testing is routinely performed.
- MODY1: Mutations in the hepatocyte nuclear factor-4 alpha (HNF4A) gene cause a clinical presentation similar to HNF1A. However, mutations in this gene are much less common (<10% of MODY). Age of onset may be later, and there is not a low renal threshold. HNF4A mutations can also cause high birth weight in newborns and transient neonatal hypoglycemia. These patients are also more sensitive to sulfonylurea treatment.

- The remaining ten genes are rare causes of MODY, each accounting for <1%:2-4
  - MODY5: Caused by heterozygous mutations in HNF1B. The vast majority of HNF1B mutations cause Renal Cysts and Diabetes Syndrome, which is associated with diabetes, renal cysts, genitourinary malformations, pancreatic atrophy, hyperuricemia, and abnormal liver function tests.
  - MODY8: Caused by heterozygous mutations in CEL. Affected individuals also have pancreatic exocrine dysfunction (diabetes-pancreatic-exocrine dysfunction syndrome).
  - Others include: MODY4 (PDX1/IPF-1), MODY6 (NEUROD1), MODY7 (KLF11), MODY9 (PAX4), MODY10 (INS), MODY11 (BLK), MODY12 (ABCC8) and MODY13 (KCNJ11).

- Other monogenic causes of pediatric diabetes include the following (not meant to be an all-inclusive list):3,5-6
  - Permanent neonatal diabetes mellitus (PNDM), defined as persistent hyperglycemia in the first 6 months of life. It is most commonly caused by mutations in the ABCC8, KCNJ11, and INS genes. Biallelic mutations in GCK and PDX1 are less common causes.
  - Transient neonatal diabetes mellitus (TNDM), which accounts for ~50% of all neonatal diabetes. Affected individuals are at risk for recurrence later in life. 70% of TNDM cases are due to 6q24 methylation defects, while ABCC8 and KCNJ11 combined account for an additional 26% of cases.
  - Cystic fibrosis, caused by biallelic CFTR mutations (for more information, see test-specific guideline, Cystic Fibrosis Testing).
  - Immune dysregulation, polyendocrinopathy, and enteropathy, X-linked (IPEX syndrome), due to mutations in FOXP3.
  - Maternally inherited diabetes and deafness (MIDD), caused by mutations in mitochondrial genes: MT-TL1, MT-TK, or MT-TE.
  - Wolcott-Rallison syndrome, due to mutations in EIF2AK3.
  - Wolfram syndrome, caused by mutations in WFS1 and less often CISD2.
o Other genes associated with PNDM and extra-pancreatic features, including GATA6, GLIS3, IER3IP1, NEUROG3, PTF1A, and RFX6.

**Test information**

- There are various methods used to test for mutations in genes which can cause MODY.
  - Single gene analysis
  - Panel testing using next generation sequencing

- Until recently, most sequencing tests used the Sanger sequencing methodology that was originally developed in the 1970s. Sanger sequencing is labor intensive and did not lend itself to high-throughput applications.

- Next generation sequencing (NGS), which is also sometimes called massively parallel sequencing, has been developing since about 2005 to allow larger scale and more efficient gene sequencing. NGS relies on sequencing many copies of small pieces of DNA simultaneously and using bioinformatics to assemble the sequence.

- The efficiency of NGS has led to an increasing number of large, multi-gene testing panels. NGS panels that test several genes at once are particularly well-suited to conditions caused by more than one gene or where there is considerable clinical overlap between conditions, making it difficult to reliably narrow down likely causes. As a result, several laboratories have begun to combine genes involved in certain conditions, which often have both of those characteristics. However, NGS may not perform as well as Sanger sequencing in some applications. Results may also be obtained that cannot be adequately interpreted based on the current knowledgebase. When a sequence variation is identified that has not been previously characterized or shown to cause the disorder in question, it is called a variant of uncertain significance (VUS). VUSs are relatively common findings when sequencing large amounts of DNA with NGS.

- MODY multi-gene panels include a wide variety of genes associated with MODY and monogenic diabetes in general. The following are example panels (not intended to be a complete list):
  - Ambry Genetics: MODY Panel Next Generation Sequencing and Deletion/Duplication Analysis of HNF1A, HNF1B, HNF4A, GCK, PDX1
  - Athena: Monogenic Diabetes (MODY) Evaluation (2-Gene, 3-Gene, 4-Gene, and 5-Gene available)
  - GeneDx: MODY Sequencing Panel
  - Prevention Genetics: Maturity Onset Diabetes of the Young (MODY) Sequencing and Deletion/Duplication Panels
Multi-gene panels may also include genes associated with other types of monogenic diabetes and glycemic disorders, such as neonatal diabetes, syndromic diabetes, and familial hyperinsulinism.

Under certain circumstances, technologies used in multi-gene testing may fail to identify mutations that might be identifiable through single-gene testing. If high clinical suspicion remains for a particular syndrome after negative multi-gene test results, consultation with the testing lab and/or additional targeted genetic testing may be warranted.

Multi-gene tests vary in technical specifications (e.g., depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis).

Since genes can be easily added or removed from multi-gene tests over time by a given lab, medical records must document which genes were included in the specific multi-gene test used from each patient, and in which labs they were performed.

Additionally, tests should be chosen that maximize the likelihood of identifying mutations in the genes of interest and that will alter patient management.

Guidelines and evidence

The American Diabetes Association (2017) states: “Children and adults, diagnosed in early adulthood, who have diabetes not characteristic of type 1 or type 2 diabetes that occurs in successive generations (suggestive of an autosomal dominant pattern of inheritance) should have genetic testing for maturity-onset diabetes of the young” (Grade A recommendation).7

The National Academy of Clinical Biochemistry (2011) states (direct quotes):8

- Routine measurement of genetic markers is not of value at this time for the diagnosis or management of patients with type 1 diabetes. For selected diabetic syndromes, including neonatal diabetes, valuable information can be obtained with definition of diabetes-associated mutations. A (moderate)
- There is no role for routine genetic testing in patients with type 2 diabetes. These studies should be confined to the research setting and evaluation of specific syndromes. A (moderate)

The International Society for Pediatric and Adolescent Diabetes (2014) makes the following recommendations:3

- “The diagnosis of maturity-onset diabetes of the young (MODY) should be suspected in cases with”
  - “A family history of diabetes in one parent and first degree relatives of that affected parent in patients who lack the characteristics of type 1 diabetes [no
islet autoantibodies, low or no insulin requirements 5 yr after diagnosis (stimulated C-peptide >200 pmol/L)] and lack the characteristics type 2 diabetes (marked obesity, acanthosis nigricans).”

- “Mild stable fasting hyperglycemia which does not progress. Such cases should be tested for glucokinase (GCK) gene mutations, which is the commonest cause of persistent, incidental hyperglycemia in the pediatric population (B).”

- “Specific features can suggest subtypes of MODY, such as renal developmental disease or renal cysts (HNF1B-MODY) and macrosomia and/or neonatal hypoglycemia (HNF4A-MODY) (C).”

- “In familial autosomal dominant symptomatic diabetes, mutations in the hepatocyte nuclear factor 1α (HNF1A) gene (HNF1A-MODY) should be considered as the first diagnostic possibility, while mutations in the GCK gene are the most common cause in the absence of symptoms or marked hyperglycemia (B).”

- “Three genes are responsible for the majority of MODY cases (GCK, HNF1A, and HNF4A) … However, up to 13 different genes have been reported to cause autosomal dominant non-insulin dependent diabetes but these are so unusual they do not need to be tested for in children with diabetes except in a research setting or when there are additional phenotypes such as pancreatic exocrine dysfunction.”

• The European Molecular Genetics Quality Network (2008) makes the following recommendations for testing (paraphrased due to their length):^4

- Testing for GCK mutations (presentation outside of pregnancy):
  - Persistent, stable elevation of fasting blood glucose (5.5-8 mmol/l)
  - HbA1c just above the upper limit of normal (rarely exceeds 7.5%)  
  - Oral glucose tolerance testing demonstrates a small increment (4.6 mmol/l is often used to prioritize testing)
  - May have a family history consistent with autosomal dominant inheritance

- Testing for GCK mutations (for evaluation of gestational diabetes):
  - Persistent elevation of fasting blood glucose (5.5-8 mmol/l) before, during and after pregnancy
  - At least one oral glucose tolerance test with an increment of <4.6 mmol/l (either during or after pregnancy)

- Testing for HNF1A mutations:
  - Young-onset diabetes (<25 years old)
  - Non-insulin-dependent diabetes
- Family history of diabetes (at least two generations)
- Absence of pancreatic islet autoantibodies
- Glycosuria at blood glucose levels <10 mmol/l
- Marked sensitivity to sulfonylureas
- Features suggestive of monogenic diabetes (lack of obesity or evidence of insulin resistance, absence of acanthosis nigricans, etc)

  - Testing for HNF4A mutations:
    - Should be considered when HNF1A analysis is normal but the clinical features are strongly suggestive of HNF1A
    - “When diabetic family members have marked macrosomia (>4.4 kg at term) or if diazoxide-responsive neonatal hyperinsulinism has been diagnosed in the context of familial diabetes.”
    - “Macrosomic babies with diazoxide-responsive hyperinsulinism and a strong family history of diabetes should be considered for HNF4A mutation screening.”

  - Syndromic forms of diabetes, including HNF1B and CEL mutations, “are not included in these guidelines since testing is guided by the non-endocrine pancreatic or extra-pancreatic clinical features.”

Criteria

This guideline applies to all MODY testing, including single genes as well as multi-gene panels, which are defined as assays that simultaneously test for more than one MODY gene. Medical necessity determination generally relies on criteria established for testing individual genes.

Medical necessity criteria differ based on the type of testing being performed (i.e., individual MODY genes separately chosen versus pre-defined panels of MODY genes) and how that testing will be billed (one or more individual MODY gene procedure codes, specific panel procedure codes, or unlisted procedure codes).

These guidelines are for gene testing in the context of MODY evaluation only. For gene testing in non-MODY contexts (e.g., neonatal diabetes, familial hyperinsulinism, etc.), refer to the general policies, Genetic Testing to Diagnose Non-Cancer Conditions and Genetic Testing by Multigene Panels, as appropriate.

HNF1A Sequencing and Deletion/Duplication Analysis

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous HNF1A gene sequencing or deletion/duplication analysis, and
  o No known mutation in biologic relative, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Member has a diagnosis of diabetes prior to 25 years of age, and
  o Member has a biological parent with diabetes, and
  o Member does NOT have symptoms consistent with a specific condition or specific gene mutation, and
  o Member does NOT have any of the following features:
    ▪ Extra-pancreatic manifestations (e.g., congenital malformations and other signs of syndromic diabetes), or
    ▪ Pancreatic autoantibodies suggestive of type 1 diabetes, or
    ▪ Body mass index (BMI) ≥ 35 kg/m², or
    ▪ Acanthosis nigricans, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**HNF4A Sequencing and Deletion/Duplication Analysis**

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous HNF4A gene sequencing or deletion/duplication analysis, and
  o No known mutation in biologic relative, and
  o Member has previous HNF1A testing with no deleterious mutation found, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**GCK Sequencing and Deletion/Duplication Analysis**

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous GCK gene sequencing or deletion/duplication analysis, and
• No known mutation in biologic relative, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Member has previous HNF1A testing with no deleterious mutation found, or
  o Member has a personal history of the following features presenting outside of pregnancy:
    ▪ Persistent, stable elevation of fasting blood glucose (5.5-8 mmol/L), and
    ▪ HbA1C that is no more than mildly elevated (≤7.5%), and
    ▪ At least one oral glucose tolerance test demonstrates a small increment (<4.6 mmol/L), or
  o Member has a personal history of the following features in the context of gestational diabetes:
    ▪ Persistent elevation of fasting blood glucose (5.5-8 mmol/L) before, during, and after pregnancy, and
    ▪ At least one oral glucose tolerance test demonstrates a small increment (<4.6 mmol/L) either during or after pregnancy, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Sequencing and Deletion/Duplication Analysis of ABCC8, BLK, CEL, HNF1B, INS, KCNJ11, KLF11, NEUROD1, PAX4, and PDX1

Sequencing and deletion/duplication analysis of these genes in the context of MODY testing is not a covered benefit.

• The clinical utility of these tests for the evaluation of MODY has not been well established. Mutations in HNF1A, GCK, and HNF4A are responsible for the majority of cases of MODY, making them the most common known genetic causes of the disorder. There are other genes associated with MODY, but mutations in each gene account for <1% of cases of MODY, therefore incremental mutation yield of individual gene testing is expected to be very low. In addition, medical management guidelines have not been established for most of these forms of MODY.
• Gene testing is not covered strictly for the indication of MODY testing. Testing in other contexts may meet medical necessity criteria (e.g., HNF1B testing for individuals with symptoms of Renal Cysts and Diabetes Syndrome, CEL testing for individuals with diabetes and pancreatic exocrine dysfunction, or certain gene tests for individuals with neonatal diabetes or familial hyperinsulinism). For gene testing in non-MODY contexts, refer to Genetic Testing for Non-Cancer Conditions.

MODY Multi-Gene Panels

When separate procedure codes will be billed for individual MODY genes (e.g., Tier 2 MoPath codes 81400-81408), each individually billed test will be evaluated separately. The below criteria for single gene testing will be applied.

If the member meets the following criteria, the entire panel will be approved. However, the laboratory will be redirected to use a panel CPT code for billing purposes (e.g. 81479):

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous MODY genetic testing, and
  o No known mutation in biologic relative, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Member has a diagnosis of diabetes prior to 25 years of age, and
  o Member has a family history of diabetes consistent with autosomal dominant inheritance, and
  o Member does NOT have symptoms consistent with a specific condition or specific gene mutation, and
  o Member does NOT have any of the following features:
    ▪ Extra-pancreatic manifestations (e.g., congenital malformations and other signs of syndromic diabetes), or
    ▪ Pancreatic autoantibodies suggestive of type 1 diabetes, or
    ▪ Body mass index (BMI) $\geq 35$ kg/m$^2$, or
    ▪ Acanthosis nigricans, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

When a multi-gene panel is being requested and will be billed with a single panel CPT code (e.g. 81479), the panel will be considered medically necessary when the following criteria are met:
• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous MODY genetic testing, and
  o No known mutation in biologic relative, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Member has a diagnosis of diabetes prior to 25 years of age, and
  o Member has a family history of diabetes consistent with autosomal dominant inheritance, and
  o Member does NOT have symptoms consistent with a specific condition or specific gene mutation, and
  o Member does NOT have of the following features:
    ▪ Extra-pancreatic manifestations (e.g., congenital malformations and other signs of syndromic diabetes), or
    ▪ Pancreatic autoantibodies suggestive of type 1 diabetes, or
    ▪ Body mass index (BMI) ≥ 35 kg/m^2, or
    ▪ Acanthosis nigricans, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Billing and reimbursement considerations

• When multiple CPT codes are billed for components of a panel and there is a more appropriate CPT code representing the panel, eviCore will redirect to the panel code(s).

• If the laboratory will not accept redirection to a panel code, the medical necessity of each billed component procedure will be assessed independently.
  o In general, only a limited number of panel components that are most likely to explain the member’s presentation will be reimbursable. The remaining panel components will not be reimbursable.
  o When a MODY multi-gene panel is billed with multiple stacked codes, only the following genes may be considered for reimbursement:
    ▪ HNF1A
    ▪ GCK
References


2. Genetic and Rare Diseases Information Center. Maturity-onset diabetes of the young. Available at: https://rarediseases.info.nih.gov/diseases/3697/maturity-onset-diabetes-of-the-young.


MGMT Testing for Malignant Glioma Alkylation Agent Response

Procedures addressed

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<table>
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<tr>
<td>MGMT Promoter Methylation Analysis</td>
<td>81287</td>
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</table>

What is MGMT

Definition

MGMT is the O6-methylguanine- DNA methyltransferase gene, which encodes an essential DNA repair enzyme. MGMT expression in tumors causes resistance to DNA- alkylation drugs. MGMT repairs the damage produced by these DNA cross linking agents.1

- Gene methylation is a control mechanism that regulates gene expression. If the MGMT gene is hypermethylated, its expression is absent (“turned off”) or reduced (“turned down”). With less MGMT DNA repair protein present, the tumor is typically more responsive to alkylation drugs.2
- Glioblastoma is a common and aggressive brain tumor that is often treated with alkylation drugs.2 Temozolomide is a standard systemic chemotherapy shown to be effective for malignant gliomas.2
- About 40-50% of glioblastoma tumors exhibit MGMT hypermethylation, leading to increased chemosensitivity.3,4
- Treatment of gliomas often includes resection, radiation, and chemotherapy. For frail or elderly patients, combined treatment may not be tolerated; therefore, treatment with a single agent (radiation therapy or chemotherapy) or chemotherapy with deferred radiation therapy may be considered.1

Test information

- MGMT promoter methylation testing is performed on paraffin embedded tumor tissue. Quantitative methylation-sensitive PCR or pyrosequencing is used to determine MGMT gene promoter methylation levels.
Guidelines and evidence

The National Comprehensive Cancer Network (NCCN, 2016) stated that:

- “MGMT promoter methylation is particularly useful in treatment decisions for elderly patients with high grade gliomas (grades III-IV).”
- “Patients with glioblastoma that are not MGMT promoter methylated derive less benefit from treatment with temozolomide compared to those whose tumors are methylated.”

In September 2012, Alberta Health Services published a Clinical Practice Guideline on Glioblastoma. It concluded:

- “Determination of MGMT promoter methylation status may assist in determination of prognosis.”
- “…whenever possible, determination of MGMT promoter methylation status should be conducted, as it may assist in determination of prognosis.”

An analysis of epigenetic promoter methylation of the MGMT gene in 206 patients with glioblastoma demonstrated:

- Significantly improved median survival for those with a methylated MGMT promoter—21.7 months for those treated with temozolomide compared to 15.3 months for those treated with radiotherapy alone (p=0.007).
- Marginally improved median survival for those without a methylated MGMT promoter—12.7 months for those treated with temozolomide versus 11.8 months for those treated with radiotherapy alone (p=0.06).
- MGMT promoter methylation was an independent prognostic factor for favorable response to any glioblastoma treatment (HR=0.45, 95% CI 0.32 – 0.61; p<0.001).

Criteria

Testing criteria:

- Diagnosis of glioblastoma (or gliosarcoma)¹, and
- Adjuvant temozolomide chemotherapy is being considered¹, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


Mitochondrial DNA Deletion Syndromes

Procedures addressed

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<tr>
<td>mtDNA Deletion Analysis</td>
<td>81465</td>
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</table>

What are mtDNA deletion syndromes

Definition

Mitochondrial DNA deletion syndromes include three overlapping phenotypes: Kearns-Sayre syndrome (KSS), Pearson syndrome, and progressive external ophthalmoplegia (PEO).¹ ²

- The three phenotypes may be observed in different members of the same family or may evolve in a given individual over time.¹
  - KSS is a multisystem disorder defined by three key signs and symptoms: onset before age 20 years (typically in childhood), pigmentary retinopathy, and PEO. Affected individuals must also have at least one of the following: cardiac conduction block, cerebrospinal fluid protein concentration >100 mg/dL, or cerebellar ataxia. Other findings may include short stature, hearing loss, dementia, limb weakness, diabetes mellitus, hypoparathyroidism, and growth hormone deficiency.¹ ²
  - Pearson syndrome includes the findings of sideroblastic anemia and exocrine pancreas dysfunction. It is usually fatal in infancy. Those surviving into childhood develop features of KSS.¹ ³
  - Symptoms may first occur between the first and fifth decade of life and may not appear in any particular order.¹
  - PEO is a mitochondrial myopathy characterized by findings including drooping of the eyelids (ptosis), paralysis of the extraocular muscles (ophthalmoplegia), and variably severe proximal limb weakness.¹
  - Rarely Leigh syndrome can manifest due to a mtDNA deletion which is characterized by basal ganglia and brain stem lesions.¹

- These conditions are caused by pathogenic variants in mitochondrial DNA (mtDNA). Pathogenic variants can be sporadic (not inherited) or maternally inherited. A female who carries the mtDNA mutation at high mutation load will
typically pass it on to all of her children. A male who carries the mtDNA mutation cannot pass it on to his children.\textsuperscript{1-3}

- The wide variability in clinical presentation depends on how much mutant mtDNA is present in a tissue (heteroplasmy), which organs and tissues have mutant mtDNA, and how vulnerable those tissues are to impaired mitochondrial function (threshold effect).\textsuperscript{1}
- Management is usually symptomatic and supportive.\textsuperscript{1}
- An epidemiologic study of an adult population in the North East of England estimated the prevalence of large-scale mtDNA deletions at 1.2:100,000.\textsuperscript{4}

**Test information**

- Diagnosis of mtDNA deletion syndromes is based on a combination of clinical findings and genetic testing.\textsuperscript{1,2}
- Findings in KSS and PEO may include elevated lactate and pyruvate levels in blood and cerebrospinal fluid while at rest, with excessive increases in blood after moderate activity. MRI can demonstrate leukencephalopathy, often associated with cerebral or cerebellar atrophy or basal ganglia lesions.\textsuperscript{1} Biochemical studies may also be performed, though: "It is important to note that biochemical abnormalities may not be present during periods when the mitochondrial disease is quiescent/dormant."\textsuperscript{5}
- Detection rate for cases of KSS and PEO by deletion/duplication analysis is 90% and 50% respectively.\textsuperscript{1}
  - In cases of KSS and PEO, the disease-causing rearrangements can be detected on a muscle specimen but typically are undetectable in blood, therefore mutational analysis is best obtained through muscle biopsy by NGS.\textsuperscript{1} The same would apply to the rare cases of Leigh syndrome.\textsuperscript{1}
  - For Pearson syndrome, the rearrangements can be detected in blood by whole mitochondrial genome amplification followed by massively parallel sequencing detecting about 90% of those affected.\textsuperscript{1,2}
- Any molecular genetic test for a mtDNA mutation should ideally be directed by the clinical phenotype and results of other clinical, laboratory, and radiological investigations.\textsuperscript{2}
- Genetic test results alone cannot predict the exact course or phenotype of the disease. Therefore, testing is not appropriate for asymptomatic at-risk individuals.\textsuperscript{1,2}

**Guidelines and evidence**

- No specific evidence-based U.S. testing guidelines were identified.
• Case reports and a limited number of case series are the primary evidence base available for the diagnosis of mitochondrial disease. There are few prospective studies. The Mitochondrial Medicine Society developed consensus recommendations using the Delphi method and published them in 2015.6

  o Recommendations for DNA testing
    ▪ “Massively parallel sequencing/NGS of the mtDNA genome is the preferred methodology when testing mtDNA and should be performed in cases of suspected mitochondrial disease instead of testing for a limited number of pathogenic point mutations.”
    ▪ “Patients with a strong likelihood of mitochondrial disease because of a mtDNA mutation and negative testing in blood, should have mtDNA assessed in another tissue to avoid the possibility of missing tissue-specific mutations or low levels of heteroplasmy in blood; tissue-based testing also helps assess the risk of other organ involvement and heterogeneity in family members and to guide genetic counseling.”
    ▪ “Heteroplasmy analysis in urine can selectively be more informative and accurate than testing in blood alone, especially in cases of MELAS due to the common m.3243 A>G mutation.”
    ▪ “When considering nuclear gene testing in patients with likely primary mitochondrial disease, NGS methodologies providing complete coverage of known mitochondrial disease gene is preferred. Single-gene testing should usually be avoided because mutations in different genes can produce the same phenotype. If no mutation is identified via known NGS panels, then whole exome sequencing should be considered.”

• A workshop of the National Institute of Neurological Disorders and Stroke (2008) summarizes:5

  o “The diagnosis of mitochondrial diseases is complicated by their heterogeneous presentations and by the lack of screening procedures or diagnostic biomarkers that are both sensitive and specific. The workshop panelists explained that diagnosis is often a lengthy process beginning with a general clinical evaluation followed by metabolic screening and imaging and finally by genetic tests and more invasive biochemical and histological analyses. The identification of known mitochondrial mutations in tissue has greatly aided diagnosis. However, even when clinical features and family history strongly suggest mitochondrial disease, the underlying genetic mutation can elude detection, and there is no current screening procedure that would be practical for all cases of suspected mitochondrial disease.”

• The European Federation of Neurological Sciences (2009) 7 provided molecular diagnostic evidence-based guidelines for these conditions:

  o “If the phenotype suggests syndromic MID [mitochondrial disorders] due to mtDNA deletion (mtPEO, KSS, Pearson's syndrome), mtDNA analysis starts
with RFLP or Southern-blot from appropriate tissues. mtDNA deletions with low heteroplasmy rate may be detected only by long-range PCR. If neither a single deletion nor multiple deletions are found, mtDNA sequencing is recommended.”

Criteria

Known Familial Mutation Testing

- Genetic Counseling
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing
  - No previous genetic testing in the individual for mtDNA deletion syndromes, and
  - A maternal deletion identified in the mother, AND
- Diagnostic Testing for Symptomatic Individual:
  - Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of a mtDNA deletion syndrome, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy

mtDNA Deletion Testing

- Genetic Counseling
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Testing:
  - No previous genetic testing for mtDNA deletions,** and
  - No known mitochondrial pathogenic variants or deletions in the family, AND
- Diagnostic Testing for Symptomatic Individuals:
  - Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of a mtDNA deletion syndrome, and
  - Genetic testing is needed to confirm the diagnosis, AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy

** Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.
References


Mitochondrial Encephalomyopathy, Lactic Acidosis, and Stroke-like Episodes (MELAS) Genetic Testing

Procedures addressed

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<td>MT-TL1 Targeted Mutation Analysis</td>
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<tr>
<td>MT-ND5 Targeted Mutation Analysis</td>
<td>81401</td>
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<tr>
<td>Whole Mitochondrial Genome</td>
<td>81460</td>
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</table>

What is MELAS

Definition

Mitochondrial Encephalomyopathy, Lactic Acidosis, and Stroke-like Episodes (MELAS) is a progressive, multisystem genetic disease.¹

- The estimated prevalence of MELAS is about 16-18/100,000 individuals.²,³
- MELAS symptoms can present at any age. Most cases present between 2-10 years of age, but some present between 10-40 years of age.¹
- Individuals with MELAS typically experience disease progression that results in death. Median survival time from point of diagnosis is about 16.9 years, with a subgroup of 20.8% who are more severely affected and die within 7.3 years of diagnosis.¹ Overall, children and young adults diagnosed with MELAS who have classical symptoms have a shorter lifespan than older adults with milder symptoms.
- Typical initial clinical presentation includes stroke-like episodes or cortical blindness often occurring with generalized tonic-clonic seizures, and these episodes may be recurrent and associated with altered consciousness. Almost all individuals with MELAS (94%) have lactic acidemia. Individuals may also have recurrent headaches, anorexia, recurrent vomiting, possibly exercise intolerance or proximal limb weakness, Wolff-Parkinson-White, and diabetes mellitus. Short stature in children and sensorineural hearing loss in both children and adults is also common.¹
• The natural history of MELAS involves gradual impairment of motor abilities, vision, and cognitive ability by adolescence or young adulthood due to recurring stroke-like episodes.¹

• There is no cure for MELAS. Several types of treatment, however, have demonstrated benefit in affected individuals. The use of oral and intravenous (IV) L-arginine and citrulline has shown reduction of frequency and/or severity of stroke-like episodes.⁴⁻⁹ Both endurance and resistance exercise have been studied and shown to increase mitochondrial metabolism.⁶ Vitamin and cofactor supplementation including CoQ10, alpha lipoic acid, and riboflavin should be offered, and addition of folinic acid and L-carnitine should be considered, especially if there is documented deficiency.⁴

• At-risk individuals may benefit from assessment to initiate baseline evaluations (neurology, cardiology, ophthalmology, and audiology) and potential intervention prior to exhibiting clinical manifestations.⁵ Screening for diabetes mellitus by fasting serum glucose concentration and glucose tolerance test is recommended.¹

• Diagnosis of MELAS is based on a combination of clinical findings and genetic testing.¹,¹¹

• MELAS is caused by mutations in the mitochondrial DNA (mtDNA) that are always maternally inherited. This means that a female who carries the mtDNA mutation will pass it on to all of her children. A male who carries the mtDNA mutation will not pass it on to his children.¹,¹¹

• Mutations in the mtDNA gene, MT-TL1, cause MELAS. A majority of affected individuals with classic symptoms, about 80%, have a specific mutation, A3243G.¹,¹⁰,¹¹ Other rare mtDNA mutations in the MT-TL1 gene, T3271C and A3252G, and in 9 other mtDNA genes are also associated with MELAS.¹,¹¹

• Genetic test results alone cannot predict the exact course or phenotype of the disease.¹,¹¹ For all mtDNA mutations, clinical expressivity depends on the three following factors:¹
  o The relative abundance of mutant mtDNA, mutational load (heteroplasmy)
  o The organs and tissues in which the mutant mtDNA is found (tissue distribution), and
The vulnerability of each tissue to impaired oxidative metabolism (threshold effect).

- There is suggested clinical utility with the use of genetic testing for MELAS at the present time. Each patient and family is unique therefore it necessary to consider the case to determine the clinical utility in regards to impactful management. This may include changes to stroke treatment, treatment during illness, the use of anesthesia, the use of exercise as treatment, and the use of vitamin and xenobiotics.

### Test information

- The investigation and diagnosis of patients with mitochondrial respiratory chain disease often necessitates a combination of techniques including muscle histocytochemistry, biochemical assessment and molecular genetic studies along with clinical assessment. Any molecular genetic test for a mtDNA mutation should ideally be directed by the clinical phenotype and results of these other investigations.

- Targeted mutation testing for MELAS is available at many laboratories. The specific mutations included in these targeted tests can vary by laboratory; however, they typically include the most common pathogenic variant found in MELAS, m.3243 A>G.

- The common MELAS mutations are also included on a number of more general mitochondrial targeted mutation panels (in conjunction with genes for LHON, MERRF and Leigh syndrome).

- Full sequencing of the entire mitochondrial genome can be done to identify the remaining rare mtDNA mutation in individuals affected with MELAS. Since the mitochondrial genome is highly polymorphic, this is not routinely offered unless clinical suspicion is very high and paternal transmission has been ruled out. If the status of heteroplasmy is of concern, next generation testing with high read depth may be preferable.

- A number of large panels sequence the mitochondrial genome in conjunction with nuclear-encoded mitochondrial genes for a broad approach to testing.

- DNA testing can be performed on a blood specimen. Muscle biopsy is generally not necessary, but some labs accept blood, saliva and muscle samples.
• A muscle biopsy or heteroplasmy analysis in urine may be recommended for testing of A3243G variant in cases with a clinical presentation of classic MELAS and where the variant is not detected on blood or urine specimens. If the status of heteroplasmy is of concern, next generation testing with high read depth may be preferable, however certain targeted mutation analysis can detect low level heteroplasmy.

Guidelines and evidence
• No specific evidence-based U.S. testing guidelines for MELAS were identified.
• The Mitochondrial Medicine Society (2015) developed consensus recommendations for the diagnosis and management of mitochondrial disease. Testing strategies, including strategies for genetic testing, were discussed.
  o Recommendations for DNA testing
    ▪ “Massively parallel sequencing/NGS of the mtDNA genome is the preferred methodology when testing mtDNA and should be performed in cases of suspected mitochondrial disease instead of testing for a limited number of pathogenic point mutations.”
    ▪ “Patients with a strong likelihood of mitochondrial disease because of a mtDNA mutation and negative testing in blood, should have mtDNA assessed in another tissue to avoid the possibility of missing tissue-specific mutations or low levels of heteroplasmy in blood; tissue-based testing also helps assess the risk of other organ involvement and heterogeneity in family members and to guide genetic counseling. Heteroplasmy analysis in urine can selectively be more informative and accurate than testing in blood alone, especially in cases of MELAS due to the common m.3243A>G mutation.”
  o Recommendations for pathology testing
    ▪ “Muscle (and/or liver) biopsies should be performed in the routine analysis for mitochondrial disease when the diagnosis cannot be confirmed with DNA testing.”
• The European Federation of Neurological Sciences (EFNS, 2009) provided molecular diagnostic consensus-based guidelines based on literature reviews:
  o “If the phenotype suggests syndromic mitochondrial disease due to mtDNA point mutations (MELAS, MERRF, NARP, LHON) DNA-microarrays using allele-specific oligonucleotide hybridization, real-time-PCR or single-gene sequencing are indicated.”
• The clinical utility of genetic testing for MELAS was described by a workshop of the National Institute of Neurological Disorders and Stroke (2008):
  o “The diagnosis of mitochondrial diseases is complicated by their heterogeneous presentations and by the lack of screening procedures or diagnostic biomarkers that are both sensitive and specific. The workshop panelists explained that diagnosis is often a lengthy process beginning with a general clinical evaluation followed by metabolic screening and imaging and finally by genetic tests and
more invasive biochemical and histological analyses. The identification of known mitochondrial mutations in tissue has greatly aided diagnosis. However, even when clinical features and family history strongly suggest mitochondrial disease, the underlying genetic mutation can elude detection, and there is no current screening procedure that would be practical for all cases of suspected mitochondrial disease."

  o Initial screening includes testing for blood lactate, urine amino acids, acylcarnitine profile, and MRI. “It is important to note that biochemical abnormalities may not be present during periods when the mitochondrial disease is quiescent/dormant.”

• The Clinical Molecular Genetics Society (CMGS) of UK (2008) provided practice-based guidelines for the molecular diagnosis of mitochondrial disease:11

  o In cases with strong clinical evidence, testing should begin with checking for the common A3243G mutation. Testing for the rare mutations including T3271C and A3252G is not routinely indicated unless there is strong clinical diagnosis of MELAS testing.

Criteria

MELAS Known Familial Mutation Testing

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing
  o No previous genetic testing in the individual for MELAS, and
  o MELAS pathogenic variant identified in 1st degree biological maternal relative, AND

• Predictive Testing for Asymptomatic Individual:
  o 18 years of age or older, or
  o Under the age of 18 years, and
    ▪ Presymptomatic screening for diabetes mellitus is being considered, OR

• Diagnostic Testing for Symptomatic Individual:
  o Clinical exam and biochemical testing suggestive, but not confirmatory, of a diagnosis of MELAS, OR

• Prenatal Testing for At-Risk Pregnancies:
• MELAS causing pathogenic variant in a previous child or in the mother, AND
  • Rendering laboratory is a qualified provider of service per the Health Plan policy

MELAS Targeted Mutation Analysis (A3243G)

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous genetic testing for MELAS, and
  o No known MELAS pathogenic variants in the family, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Clinical exam and biochemical testing suggestive, but not confirmatory, of a diagnosis of MELAS by one or more of the following:
    ▪ Lactic acidosis both in blood and in the CSF, and/or
    ▪ Muscle biopsy showing ragged red fibers, and/or
    ▪ Respiratory chain enzyme studies that are consistent with a diagnosis of MELAS, and/or
    ▪ Stroke-like episodes before the age of 40 years, and/or
    ▪ Encephalopathy with seizures and/or dementia, and
  o Genetic testing is needed to confirm the diagnosis, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

MELAS Targeted Mutation Analysis (G13513A, T3271C, and A3252G)

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Criteria for MELAS targeted mutation analysis (A3243G) is met, AND
• No pathogenic variants identified in the targeted mutation analysis (A3243G)

Whole mtDNA Sequencing

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Criteria for MELAS targeted mutation analysis is met, AND
• No pathogenic variants identified in the targeted mutation analysis (A3243G, G13513A, T3271C, and A3252G), AND
• Paternal transmission has been ruled out

References


Mitochondrial Genetic Testing

Procedures addressed

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<td>NuclearEncoded Mitochondrial Gene Sequencing Panel</td>
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What are mitochondrial disorders

Definition

Mitochondrial disorders arise from mutations in both nuclear and mitochondrial (mtDNA) components of the respiratory chain. They comprise a clinically diverse group of diseases that may present at any age and affect a single organ or present as a multi-system condition in which neurologic and myopathic features predominate. Extensive clinical variability and phenotypic overlap exists among the many discrete mitochondrial disorders.

- Mitochondrial disease is suspected in patients with a combination of clinical features in:
  - Muscle: proximal myopathy or cardiomyopathy
  - Nervous system: encephalopathy, seizures, dementia, stroke-like episodes, ataxia and spasticity and migraine
  - Eye: ptosis, ophthalmoparesis, ophthalmoplegia, optic atrophy, pigmentary retinopathy
  - Sensorineural hearing loss
  - Diabetes mellitus
  - Mid or late pregnancy loss

- Mitochondrial disease is not curable. However, in some cases, specific treatment recommendations can be made based on a person’s definitive diagnosis. Individuals at-risk for mitochondrial conditions may also benefit from clinical
assessment to initiate baseline evaluations (neurology, cardiology, ophthalmology, and audiology) and potential intervention prior to exhibiting clinical manifestations.\textsuperscript{1,2}

- Mitochondrial conditions caused by nuclear DNA variants can be maternally or paternally inherited and may follow autosomal dominant, autosomal recessive, and X-linked inheritance.

- Mitochondrial conditions caused by mtDNA are always maternally inherited. Pathogenic variants in the mtDNA may be de novo or maternally inherited. This means that a female who carries a mtDNA mutation at high mutation load will typically pass it on to all of her children. However, due to the meiotic bottleneck, the heteroplasmy level may vary significantly between generations. A male who carries the mtDNA mutation will not pass it on to his children.\textsuperscript{1,3} mtDNA deletions are rarely transmitted (less than 1\% empiric risk).\textsuperscript{4} If the mother is symptomatic, then the recurrence risk if approximately 4\%.\textsuperscript{5}

- For all mtDNA mutations, clinical expressivity depends on the three following factors:\textsuperscript{1}
  - The ratio of mutant mtDNA, mutational load (heteroplasmy)
  - The organs and tissues in which the mutant mtDNA is found (tissue distribution), and
  - The vulnerability of each tissue to impaired oxidative metabolism (threshold effect).

- Analysis of an individual’s family history may provide information regarding most likely inheritance patterns for a suspected mitochondrial condition. This may guide decisions to perform mtDNA sequencing, mtDNA deletion/duplication testing, nuclear encoded DNA sequencing, and/or nuclear encoded DNA deletion/duplication testing.

- While genetic test results alone cannot predict the exact course or phenotype of the disease, severity does correlate with mutation load for mitochondrial DNA mutations.\textsuperscript{3,6}

- Identification of a pathogenic variant in a proband can allow for informative testing of relatives at risk for diabetes, seizures, hearing loss, optic atrophy, and other findings.

### Test information

- The investigation and diagnosis of patients with mitochondrial disease often necessitates a combination of techniques including muscle histocytochemistry, biochemical assessment and molecular genetic studies along with clinical assessment.\textsuperscript{3} Any molecular genetic test for a mtDNA mutation should ideally be directed by the clinical phenotype and results of these other investigations.\textsuperscript{3}

- While biochemical analyses of an affected tissue may be informative, they are not sensitive or specific enough to definitively diagnose most mitochondrial conditions.
• Due to overlap of clinical findings of mitochondrial conditions and non-mitochondrial conditions, affected individuals are more likely to have multiple tests performed before a molecular genetic cause is identified. If an individual’s clinical findings clearly correlate with a specific mitochondrial condition, then testing can be focused on the most appropriate approach for that condition. However, if the clinical picture strongly suggests a mitochondrial condition but there is uncertainty about which subset of conditions, then larger mtDNA or nuclear DNA testing panels may be appropriate.

• “Approaches to molecular genetic testing of a proband to consider are serial testing of single genes, multi-gene panel testing (simultaneous testing of multiple genes), and/or genomic testing (e.g., sequencing of the entire mitochondrial genome, genome sequencing, or exome sequencing to identify a pathogenic variant in a nuclear gene). In many individuals in whom molecular genetic testing does not yield or confirm a diagnosis, further investigation of suspected mitochondrial disease can involve a range of different clinical tests, including muscle biopsy for respiratory chain function.”

• The efficiency of next generation sequencing (NGS) has led to an increasing number of large, multi-gene testing panels. NGS panels that test several genes at once are particularly well-suited to conditions caused by more than one gene or where there is considerable clinical overlap between conditions making it difficult to reliably narrow down likely causes. As a result, several laboratories have begun to combine genes involved in certain conditions which often have both of those characteristics.

• Mitochondrial Genome Sequencing Panels and Mitochondrial Genome Deletion/Duplication Panels:
  o Whole Mitochondrial Genome Sequencing: NGS testing is capable of simultaneously detecting point mutations, deletions, and point mutation heteroplasmies. Typically, Sanger sequence analysis will miss heteroplasmy below 20%. With suitable depth of coverage, NGS can detect heteroplasmy down to ~1%. 7-8
  o Mitochondrial Genome Deletion/Duplication Panels: Testing for deletions and duplications of the entire mitochondrial genome.

• For some, but not all, mtDNA conditions, such as MERFF, if mtDNA genetic testing is negative in a blood sample in a person with symptoms of the mtDNA condition, testing can be done on other specimens. Typically this is done when the phenotype is highly suggestive of presence of a mutation associated with a specific gene or set of genes, or when there is a need to assess reproductive risk.
  o The potential for informativeness versus the invasiveness and procedural costs are factors to consider. For instance, muscle biopsy also allows enzymatic analysis of the electron transport chain, light and ultrastructural microscopy, and mtDNA copy number analysis, which may provide highly useful information for some conditions, such as MERFF.
Genetic testing can also be done on skin fibroblasts, urinary sediment, or buccal mucosa. If cultured fibroblasts are used, measures such as limited passaging and uridine supplementation should be taken to reduce selection against mutant genotypes.

- Nuclear Encoded Mitochondrial Gene Sequencing Panel: A number of large panels are available that sequence numerous nuclear-encoded mitochondrial genes for a broad approach to testing. Multi-gene panel tests, even for similar clinical scenarios, vary considerably laboratory by laboratory in the genes that are included and in technical specifications (e.g. depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis).

Guidelines and evidence
- No specific evidence-based U.S. testing guidelines were identified.
- The Mitochondrial Medicine Society developed consensus recommendations using the Delphi method and published them in 2015.

Recommendations for DNA testing
- Massively parallel sequencing/NGS of the mtDNA genome is the preferred methodology when testing mtDNA and should be performed in cases of suspected mitochondrial disease instead of testing for a limited number of pathogenic point mutations.
- Patients with a strong likelihood of mitochondrial disease because of a mtDNA mutation and negative testing in blood, should have mtDNA assessed in another tissue to avoid the possibility of missing tissue-specific mutations or low levels of heteroplasmy in blood; tissue-based testing also helps assess the risk of other organ involvement and heterogeneity in family members and guides genetic counseling.
- When considering nuclear gene testing in patients with likely primary mitochondrial disease, NGS methodologies providing complete coverage of known mitochondrial disease genes is preferred. Single-gene testing should usually be avoided because mutations in different genes can produce the same phenotype. If no mutation is identified via known NGS panels, then whole exome sequencing should be considered.

Recommendations for pathology testing
- Biopsy should only be considered when the diagnosis cannot be confirmed with DNA testing of other more accessible tissues. Muscle (and/or liver) biopsies are often not necessary and should be avoided when possible due to their invasive nature, unless other types of analyses such as pathology, enzymology, or mtDNA copy number analyses are required for diagnosis.

- The American College of Medical Genetics and Genomics (ACMG, 2013) states the following regarding testing individuals with isolated autism for mitochondrial disorders:
“As with metabolic disorders, testing for mitochondrial disorders in persons with ASDs is recommended only if supporting symptoms or laboratory abnormalities are present.”

- The European Federation of Neurological Sciences (2009) provided molecular diagnostic consensus-based guidelines based on literature reviews: “If the phenotype suggests syndromic mitochondrial disease due to mtDNA point mutations (MELAS, MERRF, NARP, LHON) DNA-microarrays using allele-specific oligonucleotide hybridisation, real-time-PCR or single-gene sequencing are indicated.”

- The European Federation of Neurological Sciences (2009) provided molecular diagnostic consensus-based guidelines based on literature reviews: “If the phenotype suggests syndromic mitochondrial disease due to mtDNA point mutations (MELAS, MERRF, NARP, LHON) DNA-microarrays using allele-specific oligonucleotide hybridisation, real-time-PCR or single-gene sequencing are indicated.”

- The Clinical Molecular Genetics Society (CMGS) of the United Kingdom (2008) practice-based guidelines for the molecular diagnosis of mitochondrial disease state that: “In cases with strong clinical evidence, testing should begin with checking for the common mutation, m.8344A>G. Subsequent testing for other mutations, such as m.8356T>C, may be indicated in cases with a strong clinical indication of MERRF”. “For routine referrals for NARP, presence of T8993G and T8993C mutations should be investigated.”

- A workshop of the National Institute of Neurological Disorders and Stroke (2008) summarizes:

  “The diagnosis of mitochondrial diseases is complicated by their heterogeneous presentations and by the lack of screening procedures or diagnostic biomarkers that are both sensitive and specific. The workshop panelists explained that diagnosis is often a lengthy process beginning with a general clinical evaluation followed by metabolic screening and imaging and finally by genetic tests and more invasive biochemical and histological analyses. The identification of known mitochondrial mutations in tissue has greatly aided diagnosis. However, even when clinical features and family history strongly suggest mitochondrial disease, the underlying genetic mutation can elude detection, and there is no current screening procedure that would be practical for all cases of suspected mitochondrial disease.”

Criteria

Whole mtDNA Sequencing

- Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Member has not had previous whole mtDNA sequencing performed, AND
Biochemical testing appropriate for the suspected disorder has been performed and is not confirmatory of a diagnosis of a specific mitochondrial condition, AND

Member has multiple organ system involvement defined as altered function in two or more organ systems, AND

Member has one or more of the following clinical features: proximal myopathy, cardiomyopathy, encephalopathy, seizures, dementia, stroke-like episodes, ataxia, spasticity, ptosis, ophthalmoparesis, ophthalmoplegia, optic atrophy, pigmentary retinopathy, sensorineural hearing loss, diabetes mellitus, mid- or late pregnancy loss, MRI and/or MRS imaging results consistent with a mitochondrial process, and/or pathology results consistent with a mitochondrial process, AND

Member’s clinical presentation does not fit a well-described syndrome for which single-gene or targeted panel testing is available (e.g. LHON), AND

Alternate etiologies have been considered and ruled out when possible (e.g., environmental exposure, injury, infection), AND

Family history strongly suggests mitochondrial inheritance (e.g. paternal transmission has been ruled out)

Whole mtDNA Deletion/Duplication Analysis

Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

Member has not had previous whole mtDNA deletion/duplication analysis performed, AND

Biochemical testing appropriate for the suspected disorder has been performed and is not confirmatory of a diagnosis of a specific mitochondrial condition, AND

Member has multiple organ system involvement defined as altered function in two or more organ systems, AND

Member has one or more of the following clinical features: proximal myopathy, cardiomyopathy, encephalopathy, seizures, dementia, stroke-like episodes, ataxia, spasticity, ptosis, ophthalmoparesis, ophthalmoplegia, optic atrophy, pigmentary retinopathy, sensorineural hearing loss, diabetes mellitus, mid- or late pregnancy loss, MRI and/or MRS imaging results consistent with a mitochondrial process, and/or pathology results consistent with a mitochondrial process, AND

Member’s clinical presentation does not fit a well-described syndrome for which single-gene or targeted panel testing is available (e.g. LHON), AND

Alternate etiologies have been considered and ruled out when possible (e.g., environmental exposure, injury, infection), AND
• Family history strongly suggests mitochondrial inheritance (e.g. paternal transmission has been ruled out)

Nuclear Encoded Mitochondrial Gene Sequencing Panel

• Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Member has not had a previous nuclear encoded mitochondrial gene sequencing panel testing performed, AND
• Biochemical testing appropriate for the suspected disorder has been performed and is not confirmatory of a diagnosis of a specific mitochondrial condition, AND
• Member has multiple organ system involvement defined as altered function in two or more organ systems, AND
• Member has one or more of the following clinical features: proximal myopathy, cardiomyopathy, encephalopathy, seizures, dementia, stroke-like episodes, ataxia, spasticity, ptosis, ophthalmoparesis, ophthalmoplegia, optic atrophy, pigmentary retinopathy, sensorineural hearing loss, diabetes mellitus, mid- or late pregnancy loss, MRI and/or MRS imaging results consistent with a mitochondrial process, and/or pathology results consistent with a mitochondrial process, AND
• Member’s clinical presentation does not fit a well-described syndrome for which single-gene or targeted panel testing is available (e.g. LHON), AND
• Alternate etiologies have been considered and ruled out when possible (e.g., environmental exposure, injury, infection), AND
• Family history DOES NOT strongly suggest mitochondrial inheritance (e.g. paternal transmission is observed, autosomal inheritance is likely)

Exclusions

• Testing addressed in this guideline applies to patients in whom a mitochondrial disorder is suspected based on a constellation of findings commonly seen in these conditions, while not fitting clearly into one of the discrete mitochondrial syndromes. This guideline is not applicable in the following cases:
  o The patient’s findings fit into a discrete mitochondrial syndrome for which more specific testing is appropriate. Please see one of the following guidelines for information on specific mitochondrial conditions (MELAS, LHON, MNGIE, MERRF, NARP, etc); or
  o The patient’s findings could be explained nonspecifically by a mitochondrial disorder or other neurological or myopathic condition not related to mitochondrion for which a different genetic test may be considered; or
Individuals who have no increased risk above the general population risk to have inherited a mitochondrial disease and have just one of the following findings in isolation: fatigue; muscle weakness; developmental delay; autism; migraines; abnormal biochemical test results (e.g., elevated lactate); psychiatric symptoms.

Billing and reimbursement considerations

- Whole mtDNA Sequencing will only be considered for coverage when billed under the appropriate panel CPT code: 81460
- Whole mtDNA Deletion/Duplication will only be considered for coverage when billed under the appropriate panel CPT code: 81465
- Nuclear Encoded Mitochondrial Gene Sequencing Panels will only be considered for coverage when billed under the appropriate panel CPT code: 81440
- If the panel will be billed with separate procedure codes for each gene analyzed and the member meets criteria for Whole mtDNA Sequencing, Whole mtDNA Deletion/Duplication, or Nuclear Encoded Mitochondrial Gene Sequencing Panel, the testing will be approved but the laboratory will be redirected to the appropriate CPT code for billing purposes.
- If the panel cannot be redirected to 81460, 81465, or 81440 for any reason, the medical necessity of each billed procedure will be assessed independently.
- If more than one test or procedure code is requested at one time, the member meets criteria for all tests requested, and each test is equally likely based on personal history, clinical findings, and family history, the testing will be tiered in the following order: 81460, 81465, 81440.

References


Mitochondrial Neurogastrointestinal Encephalopathy (MNGIE)

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>TYMP Known Familial Mutation</td>
<td>81403</td>
</tr>
<tr>
<td>TYMP Sequencing</td>
<td>81405</td>
</tr>
<tr>
<td>TYMP Deletion/Duplication</td>
<td>81479</td>
</tr>
</tbody>
</table>

What is MNGIE

Definition

Mitochondrial Neurogastrointestinal Encephalopathy (MNGIE) is a multisystem mitochondrial disease. MNGIE is typically characterized by progressive gastrointestinal dysmotility, which may present with nausea, dysphagia, reflux, early satiety, vomiting after a meal, episodic abdominal pain, bloating, and/or diarrhea. Additionally individuals may present with cachexia (a wasting syndrome), ptosis/ophthalmoplegia (drooping/weakness of the eyelid), leukoencephalopathy on brain MRI, or peripheral neuropathy (tingling, numbness, and/or pain in the extremities). Symptoms may first occur between the first and fifth decade of life and may not appear in any particular order.

- MNGIE is caused by biallelic mutations in the TYMP gene and is inherited in an autosomal recessive pattern, meaning parents of an affected individual must be obligate carriers. The chance of having another child with MNGIE to the same parents is 25%.
- Prevalence of MNGIE is largely unknown but the condition appears to be rare. Approximately 120 cases have been reported.
- Management can be supportive, and may include assistance with swallowing difficulties, medication for nausea and vomiting, gastrostomy and parental nutrition for nutritional support, pain medications for neuropathy, and physical therapy and occupational therapy.
  - In individuals with advanced illness, liver transplant or allogenic hematopoietic stem cell transplant, have been suggested as possible curative treatment.
options, although risks and benefits of these procedures must be properly weighed.\textsuperscript{2,3}

- Peritoneal dialysis has also been suggested as a method of reduction of the thymidine concentration and should be considered as an additional or alternative form of treatment.\textsuperscript{4}

Test information

- “The TYMP gene encodes thymidine phosphorylase, a cytosolic enzyme that catalyzes the phosphorylation of thymidine or deoxyuridine to thymine or uracil, and is thus essential for the nucleotide salvage pathway.”\textsuperscript{5}
- Mutations that disrupt the function of TYMP will therefore disrupt the enzyme activity causing it to decrease and levels of thymidine or deoxyuridine to increase.
- Reduced thymidine phosphorylase enzyme activity or elevated thymidine and deoxyuridine levels are consistent with a diagnosis of MNGIE.\textsuperscript{1}
- Genetic testing of the TYMP gene can help to diagnosis a person with MNGIE.
  - The majority of TYMP mutations are detected by gene sequencing. TYMP deletions and duplications are less common.
    - Complete sequencing of TYMP for pathogenic mutations is necessary to diagnosis MNGIE.
    - If only one TYMP mutation identified or variant of uncertain significance results are returned, pursue gene TYMP deletion/duplication analysis.\textsuperscript{1}

Guidelines and evidence

- No specific evidence-based U.S. testing guidelines were identified.
- Although not specific to genetic testing for MNGIE, the Mitochondrial Medicine Society (2015)\textsuperscript{6} developed consensus recommendations for the diagnosis and management of mitochondrial disease. Testing strategies, including strategies for genetic testing, were discussed.
  - Recommendations for DNA testing include the following:
    - “Massively parallel sequencing/NGS of the mtDNA genome is the preferred methodology when testing mtDNA and should be performed in cases of suspected mitochondrial disease instead of testing for a limited number of pathogenic point mutations.”
    - “Patients with a strong likelihood of mitochondrial disease because of a mtDNA mutation and negative testing in blood, should have mtDNA assessed in another tissue to avoid the possibility of missing tissue-specific mutations or low levels of heteroplasmy in blood; tissue-based testing also
helps assess the risk of other organ involvement and heterogeneity in family members and to guide genetic counseling.”

- “Heteroplasmy analysis in urine can selectively be more informative and accurate than testing in blood alone, especially in cases of MELAS due to the common m.3243 A>G mutation.”

- “When considering nuclear gene testing in patients with likely primary mitochondrial disease, NGS methodologies providing complete coverage of known mitochondrial disease gene is preferred. Single-gene testing should usually be avoided because mutations in different genes can produce the same phenotype. If no mutation is identified via known NGS panels, then whole exome sequencing should be considered.”

- The European Federation of Neurological Sciences (2009) provided molecular diagnostic consensus-based guidelines based on literature reviews: “Sequencing of TYMP should be performed only if serum thymidine is elevated.”

- Evidence from peer reviewed journals provide symptoms, clinical findings, imaging, and family history suggestive of MNGIE.8,9

  - Severe gastrointestinal dysmotility, cachexia, ptosis, external ophthalmoparesis/ophthalmoplegia, and sensorimotor neuropathy.
  - Brain MRI that demonstrates abnormal brain white matter (increased FLAIR or T2-weighted signal) consistent with asymptomatic leukoencephalopathy.
  - Family history consistent with autosomal recessive inheritance.

**Criteria**

**TYMP Known Familial Mutation Testing**

- Genetic Counseling
  - Pre and post-test counseling by an appropriate provider(as deemed by the Health Plan policy), AND

- Previous Genetic Testing
  - No previous genetic testing in the individual for MNGIE, and
  - TYMP pathogenic variant(s) identified in parents and/or sibling(s), AND

- Predictive Testing for Asymptomatic Individual:
  - 18 years of age or older, OR
• Diagnostic Testing for Symptomatic Individual:
  o Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of MNGIE, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy

TYMP Sequencing

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Testing:
  o No previous genetic testing for MNGIE, and
  o No known TYMP pathogenic variants in the family, AND
• Diagnostic Testing for Symptomatic Individuals:
  o Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of MNGIE, and
  o Genetic testing is needed to confirm the diagnosis, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy

TYMP Deletion/Duplication

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Criteria for TYMP sequencing is met, AND
• No pathogenic variants or only one pathogenic variant identified in TYMP Sequencing.

References


Molecular Gastrointestinal Pathogen Panel (GIPP) Testing

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gastrointestinal Pathogen (eg, Clostridium difficile, E. coli, Salmonella, Shigella, Norovirus, Giardia), Includes Multiple Reverse Transcription, When Performed, And Multiplex Amplified Probe Technique, Multiple Types Or Subtypes, 3-5 Targets</td>
<td>87505</td>
</tr>
<tr>
<td>Gastrointestinal Pathogen (eg, Clostridium difficile, E. coli, Salmonella, Shigella, Norovirus, Giardia), Includes Multiple Reverse Transcription, When Performed, And Multiplex Amplified Probe Technique, Multiple Types Or Subtypes, 6-11 Targets</td>
<td>87506</td>
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<tr>
<td>Gastrointestinal Pathogen (eg, Clostridium difficile, E. coli, Salmonella, Shigella, Norovirus, Giardia), Includes Multiple Reverse Transcription, When Performed, And Multiplex Amplified Probe Technique, Multiple Types Or Subtypes, 12-25 Targets</td>
<td>87507</td>
</tr>
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</table>

What are nucleic acid amplified probe techniques (NAAT) for the identification of microorganisms via GIPP

Definition

Tests performed by NAAT uses a microorganism’s DNA or RNA to directly identify specific bacteria, viruses, and/or protozoa rather than standard microorganism detection techniques such as bacterial culture, microscopy with and without stains, direct fluorescent antibody testing, rapid antigen testing, qualitative and quantitative immunoassay for identification of antigens or toxins from stool and single-plex PCR assays. Multiplex NAAT tests are included in the larger grouping of culture-independent...
diagnostic tests (CIDT). CIDT includes but is not limited to simplex direct probe and amplified probe techniques.

- This technology offers results in a matter of hours, rather than 2-3 days of time-consuming and labor intensive bacterial cultures and immunoassays for processing stool specimens. CIDT are touted as providing a more comprehensive assessment of disease etiology by increasing the diagnostic yield compared with conventional diagnostic tests permitting earlier initiation of appropriate therapeutic agents targeted to the detected pathogen(s), if any, rather than empirical therapy until culture results are available.

- However, above and beyond microorganism detection, this type of testing does not provide the culture isolates that are needed for antimicrobial susceptibility testing, serotyping, subtyping and whole genome sequencing that are critical for monitoring trends, detecting clusters of illness and investigating outbreaks.¹

Test information

- In addition to CLIA-regulated laboratory developed tests (LDTs) by specialty (e.g. academic) laboratories, several commercial GIPP assays are currently available. For example, Binnicker has evaluated three FDA-cleared GIPP assays,² and all are closed system tests that do not allow random access for physician’s to select likely etiologic agents of diarrhea.

- Please note that the NAAT results of GIPP assays can be inconclusive and non-specific, such as the inability to always distinguish pathogenic from non-pathogenic organisms.³

Guidelines and evidence

- American College of Gastroenterology 2016 Clinical Guidelines⁴ include two relevant sections pertaining to GIPP:
  - “Traditional methods of diagnosis (bacterial culture, microscopy with and without special stains and immunofluorescence, and antigen testing) fail to reveal the etiology of the majority of cases of acute diarrheal infection. If available, the use of Food and Drug Administration-approved culture-independent methods of diagnosis can be recommended at least as an adjunct to traditional methods. (Strong recommendation, low level of evidence).”
“The new diagnostics’ best applicability is for the clinician in practice, seeing one patient at a time rather than in the public health setting, e.g., in outbreak investigations. One potential drawback of molecular technologies is the need to predefine the particular microbes being sought. In addition the significance of an identified organism may not be clear as these molecular technologies, which involve nucleic acid amplification, are limited to our existing knowledge of a microbes’ genome and do not discriminate between viable and non-viable organisms. As a result they can detect microbes at non-pathogenic levels. Given the high rates of asymptomatic carriage of enteropathogens, this can be a considerable problem. To confound matters, further multiplex techniques are more commonly associated with increased detection of mixed infections and the relative importance of each pathogen may be unclear.”

- Infectious Diseases Society of America 2017 Clinical Practice Guidelines for the Diagnosis and Management of Infectious Diarrhea make the following recommendations:

  - “A broad differential diagnosis is recommended in immunocompromised people with diarrhea, especially those with moderate and severe primary or secondary immune deficiencies... Some experts have proposed that these assays may be particularly well suited for making an organism-specific diagnosis in immunocompromised patients.”

  - “Culture-independent, including panel-based multiplex molecular diagnostics from stool and blood specimens, and, when indicated, culture-dependent diagnostic testing should be performed when there is a clinical suspicion of enteric fever or diarrhea with bacteremia (strong, moderate).”

  - “Multipathogen nucleic acid amplification tests can simultaneously detect viral, parasitic, and bacterial agents, including some pathogens that previously could not be easily detected in the clinical setting such as norovirus, and enterotoxigenic E. coli (ETEC), enteropathogenic E. coli (EPEC), and enteroaggregative E. coli (EAEC) in less time than traditional methods. The short time to results could reduce inappropriate use of antimicrobial agents to treat infections that do not require antimicrobial therapy and could shorten the time to targeted management and isolation measures for certain infections such as STEC O157. With these assays, it is common to detect the presence of >1 pathogen that may differ with regard to clinical management.”

  - “Even a positive result for 1 pathogen should be interpreted in the context of the patient’s clinical presentation, because less is known about the clinical significance of tests that detect nucleic acid as compared with traditional assays that generally detect viable organisms. The importance of detection of multiple pathogens in the same specimen is often unclear; it is unknown if all pathogens detected in the specimen are clinically relevant or if one is more strongly associated with the illness.”

- Acute diarrhea, often called gastroenteritis, can be defined as the passage of a greater number of stools of decreased form from the normal lasting < 14 days.
Acute diarrhea is generally associated with clinical features of nausea, vomiting, abdominal pain and cramps, bloating, flatulence, fever, passage of bloody stools, tenesmus and fecal urgency. It is the leading cause of outpatient visits, hospitalizations, and lost quality of life occurring domestically and those traveling abroad.

- Many episodes of acute diarrhea are self-limited and require fluid replacement and supportive care. Oral rehydration is indicated for patients who are mildly to moderately dehydrated. IV fluids may be required for more severe dehydration. Routine use of antidiarrheal agents is not recommended because many of these agents have potentially serious adverse effects, particularly in infants and young children. Antimicrobial therapy is typically warranted for adult and pediatric patients with immune systems which are severely weakened from medications, age and other primary/secondary immunocompromising illnesses/conditions.6-7

- Laboratory testing algorithms for infectious causes of diarrhea generally agree that testing is NOT warranted for community-acquired diarrhea of <7 days duration without signs or symptoms of severe (fever, bloody diarrhea, dysentery, severe abdominal pain, dehydration, hospitalization and immunocompromised state) disease. In general, when community-acquired diarrhea persists for ≥7 days, or the diarrhea is travel-related, or there are signs/symptoms of severe disease, GIPP testing may be warranted. Additional directed testing may be indicated if the GIPP results are negative and diarrhea persists. No additional testing is indicated for GIPP-positive result unless the clinical pictures changes. Clostridium difficile molecular testing is warranted on health-care associated diarrhea with onset after the 3rd inpatient day or after recent antibiotic use.

- Whereas a majority of microorganisms can be identified with up to 5 targets, typically including Salmonella, Campylobacter, Shigella, Cryptosporidium, and Shiga toxin producing E.coli, additional agents may be in the working differential diagnosis, such as (but not limited to) Clostridium difficile, additional E. coli variants, Yersinia enterocolitica, Vibrio parahaemolyticus, Giardia, Cryptosporidium, and viruses including norovirus, rotavirus, and enteric adenoviruses.

- Salient illustrations8-10 of the literature have annotated a diverse set of offending infectious agents (bacterial, parasitic and viral) in patients presenting with acute diarrhea. However, it must be emphasized that such original study recruitment criteria were not designed to stratify probability/incidence distributions of causative organisms, according to more carefully specified patient presentation categories. Furthermore, the molecular predilection for mixed infectious agent identification is a confounding factor when clinicians are trying to pinpoint the precise etiology of acute diarrhea, given the dilemma between pathogenicity and non-pathogenicity, which was briefly cited above.
• As a result, when the patient history, clinical presentation and symptoms, etc. suggest a specific microbial etiology and/or therapy, a broad GIPP consisting of >5 infectious targets is not indicated. However, broader GIPP molecular panels (e.g. 6-25 targets) might occasionally be indicated when a patient presents with a clinical scenario and overlapping symptoms consistent with multiple possible microbiological etiologies, where both diagnosis and treatment are particularly challenging (e.g., as noted above for immunocompromised patients).

Criteria
• The following clinical indications can support the use of molecular GIPP testing.
  a) Individuals with acute diarrhea with moderate-to-severe symptoms (such as fever, dysentery, severe dehydration).
  b) Individuals with community-acquired diarrhea that persists for more than seven days, or individuals with travel-associated diarrhea of uncertain etiology.
  c) Immunocompromised individuals with acute diarrhea. Immunocompromise may support the use of a relatively large number of testing targets, in concert with other supporting clinical documentation in the medical record.
• The following are contraindications to GIPP testing, with any number of targets:
  a) Immunocompetent individuals with mild diarrhea, particularly of ≤ 7 days' duration.
  b) Individuals in whom the clinical presentation of acute diarrhea suggests a specific infectious etiology, unless first-line laboratory testing should fail to detect the suspected organism, and there is still a high clinical suspicion of infectious etiology.
• Molecular GIPP testing is limited to the minimum number of targets needed for therapeutic decision making. When ordering any configuration of infectious disease targets, whether using GIPP or conventional culture, the medical record should clearly indicate the differential diagnosis of possible microorganisms based upon patient history and presenting signs/symptoms.

Billing and reimbursement

If any molecular test panels are ordered, which have variable numbers and configurations of infectious agent targets, then the following guidelines apply to the reimbursement of CPT codes 87505, 87506, and 87507, respectively:
• 87505 (3-5 targets) or 87506 (6-11 targets) is supported by ICD-10-CM R19.7 (Diarrhea, unspecified).
• 87507 (12-25 targets) is supported by ICD-10-CM R19.7 (Diarrhea, unspecified), plus at least one of the following immunodeficiency-related codes in the table: ICD10 Codes Indicating Immunocompromise

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400 Buckwalter Place Boulevard, Bluffton, SC 29910 (800) 918-8924
www.eviCore.com
• If the laboratory’s testing platform consists solely of a multiplexed panel of 12 or more targets, yet only a subset of the organisms are considered medically necessary based on the above criteria, the lab may request reimbursement for that subset of organisms using a procedure code that does not represent all organisms included on the panel.

Exclusions and other considerations

Although outbreak investigations may sometimes require use of GIPP testing, the public health evaluations of such outbreaks are beyond the scope and domain of this guideline.

ICD10 codes

ICD10 codes in this section may be used to support medical necessity as described in the above criteria.

ICD10 Codes Indicating Immunocompromise

<table>
<thead>
<tr>
<th>ICD10 code or range</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>B20</td>
<td>Human immunodeficiency virus [HIV] disease</td>
</tr>
<tr>
<td>D80.X</td>
<td>Immunodeficiency with predominantly antibody defects</td>
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<tr>
<td>D81.0</td>
<td>Combined immunodeficiencies</td>
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<tr>
<td>D82.X</td>
<td>Immunodeficiency associated with other major defects</td>
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<td>D83.X</td>
<td>Common variable immunodeficiency</td>
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<td>D84.X</td>
<td>Other immunodeficiencies</td>
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<tr>
<td>D89.X</td>
<td>Other disorders involving the immune mechanism, not elsewhere classified</td>
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<td>Y92.238</td>
<td>Other place in hospital as the place of occurrence of the external cause</td>
</tr>
<tr>
<td>Y92.239</td>
<td>Unspecified place in hospital as the place of occurrence of the external cause</td>
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</table>

References


MTHFR Variant Analysis for Hyperhomocysteinemia

Procedures addressed

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<thead>
<tr>
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</thead>
<tbody>
<tr>
<td>MTHFR Genotyping</td>
<td>81291</td>
</tr>
</tbody>
</table>

What is hyperhomocysteinemia

Definition

Hyperhomocysteinemia generally refers to mild to moderate elevations of plasma homocysteine levels, which may be defined as 15 to 40 µmol/L.¹

- Hyperhomocysteinemia may be caused by nutritional deficiencies, various medical conditions, certain drugs, smoking, and inherited factors — such as MTHFR gene variants.¹
- The MTHFR gene encodes the 5, 10-methylenetetrahydrofolate reductase (MTHFR) enzyme. MTHFR is involved in folate metabolism. The major circulating form of folate is key to converting homocysteine into methionine. Therefore, MTHFR gene variants that reduce MTHFR enzyme function may predispose one to impaired folate metabolism and ultimately mild to moderate hyperhomocysteinemia. However, homocysteine levels are usually normal if folate intake is sufficient.¹
- Both hyperhomocysteinemia in general, and MTHFR variants specifically, have been reported in association with cardiovascular disease, venous thromboembolism, pregnancy complications, and certain birth defects, such as neural tube defects.¹² However, data is inconsistent and associated risks generally small.

Test information

- MTHFR genetic testing looks for two very common gene variants: C677T and A1298C.²
- Individuals who have two variants, including at least one C677T, may have an increased risk for hyperhomocysteinemia. However, the connection between these MTHFR variants, hyperhomocysteinemia itself, and ultimate disease risk remains unclear.³⁴
• Many experts suggest that measuring homocysteine levels directly is more informative than MTHFR variant testing.⁵

• Note that serious mutations in the MTHFR gene (not the common variants discussed here) are rarely associated with a genetic disorder called homocystinuria.² MTHFR variant testing will not find the mutations that cause homocystinuria.

• MTHFR gene testing may be a component of panels for thrombophilia, cardiovascular disease risk, psychiatric conditions, or preeclampsia. There is insufficient evidence in the peer-reviewed literature to establish clinical utility for any of these indications for testing.

Guidelines and evidence

• As part of the Choosing Wisely campaign, the American College of Medical Genetics and Genomics (2015) released “Five Things Physicians and Patients Should Question,” which states:⁶
  o “Don’t order MTHFR genetic testing for the risk assessment of hereditary thrombophilia. The common MTHFR gene variants, 677C>T and 1298A>G, are prevalent in the general population. Recent meta-analyses have disproven an association between the presence of these variants and venous thromboembolism.”

• Also as part of the Choosing Wisely campaign, the Society for Maternal Fetal Medicine (2014) released “Five Things Physicians and Patients Should Question,” which states:⁷
  o “Don’t do an inherited thrombophilia evaluation for women with histories of pregnancy loss, intrauterine growth restriction (IUGR), preeclampsia and abruption. Scientific data supporting a causal association between either methylenetetrahydrofolate reductase (MTHFR) polymorphisms or other common inherited thrombophilias and adverse pregnancy outcomes, such as recurrent pregnancy loss, severe preeclampsia and IUGR, are lacking.”

• The American College of Medical Genetics and Genomics (ACMG, 2013) states:⁸
  o “It was previously hypothesized that reduced enzyme activity of MTHFR led to mild hyperhomocysteinemia which led to an increased risk for venous thromboembolism, coronary heart disease, and recurrent pregnancy loss. Recent meta-analyses have disproven an association between hyperhomocysteinemia and risk for coronary heart disease and between MTHFR polymorphism status and risk for venous thromboembolism. There is growing evidence that MTHFR polymorphism testing has minimal clinical utility and, therefore should not be ordered as a part of a routine evaluation for thrombophilia.”

• The American College of Obstetricians and Gynecologists (ACOG, 2013) states:⁹
Because of the lack of association between either heterozygosity or homozygosity for the methylenetetrahydrofolate reductase (MTHFR) C677T polymorphism and any negative pregnancy outcomes, including any increased risk for venous thromboembolism, screening with either MTHFR mutation analyses or fasting homocysteine levels is not recommended.

- The National Society of Genetic Counselors (NSGC, 2005) state that MTHFR variant testing is specifically not justified in the case of recurrent pregnancy loss based on available studies.

Criteria

This test is considered investigational and/or experimental.

- Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.

- In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References


MUTYH Associated Polyposis Testing

Procedures addressed

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What is MUTYH-associated polyposis

Definition

MUTYH-associated polyposis (MAP) is an inherited colorectal cancer syndrome caused by mutations in the MUTYH gene (also called MYH). MAP is estimated to account for 0.7% of all colorectal cancer.\(^1\)

- MAP clinical findings overlap those of familial adenomatous polyposis (FAP) and attenuated FAP (AFAP). Affected patients most often have fewer than 100 adenomas, but cases of hundreds and occasionally over 1000 polyps have been reported.\(^1,2\) Hyperplastic and sessile serrated, and traditional serrated adenomatous polyps have also been seen individuals with MAP, although adenomas remain the most common polyp type in MAP.\(^1,3\) Duodenal adenomas occur in 17-25% of individuals with MAP and gastric polyps have been reported in about 11%.\(^1,3,4\) Additionally, approximately one third of individuals with MAP have been described with colorectal cancer and no polyps or only a few polyps.\(^1\)

- Up to 26% of people who meet clinical diagnostic criteria for classic or attenuated FAP, but have normal FAP genetic test results, will have a MAP mutation.\(^1\)

- Because MAP is not clinically distinguishable from FAP or AFAP, the identification of two MUTYH mutations is required to make a MAP diagnosis.\(^1,5\)

- Adenomas and colorectal cancer tend to present later than FAP. The diagnosis of colorectal cancer is often 50 years (range of 45-59 years).\(^1,2\) The lifetime risk for colorectal cancer in individuals with MAP is 43 to 100% in the absence of timely surveillance.\(^1\) There is also an estimated 4-5% lifetime risk for duodenal cancer.\(^1,3\)
• Unlike FAP, MAP is inherited in an autosomal recessive manner — both copies of the MUTYH gene must have a mutation to be affected. This means that siblings are the only relatives likely to be affected in the family history (i.e., you do not see inheritance from parent to child as with FAP).

Test information

• MUTYH Targeted Mutation Analysis: Two MUTYH mutations are particularly common (Y165C and G382D) and account for over 80% of MUTYH mutations in Caucasians of Northern European descent. It is estimated that 1%-2% of the general northern European population is a carrier for a MUTYH mutation. Some laboratories test for only these two mutations or offer reflex options that begin with these two mutations and proceed to full gene sequencing if two mutations are not found.

• MUTYH Sequencing Analysis: MUTYH full sequencing analysis analyzes the entire gene for mutations. It is typically done in reflex to negative results from targeted mutation analysis.

• MUTYH Deletion/Duplication Analysis: If sequencing does not find two mutations, large gene deletion/duplication analysis can be performed. It remains unknown what percentage of MAP is due to large deletions/duplications/rearrangements in the gene and thus are detectable only with this technology. However, large deletions have been reported.

• MUTYH Known Familial Mutation Analysis: Once the mutations that run in the family are known, other relatives can have testing for only those mutations. This is more accurate and cost-effective.

• Multi-gene Panel Test: A multi-gene panel that includes MUTYH and other polyposis genes may also be considered.

Guidelines and evidence

• Guidelines from the National Comprehensive Cancer Network (NCCN, 2017) on High-Risk Colorectal Assessment states the following:
  o MUTYH testing criteria:
    ▪ “Personal history of >10 adenomas”
    ▪ “Individual meeting criteria 1 or 3 (NCCN, 2017) for Serrated Polyposis Syndrome (SPS) [formerly known as hyperplastic polyposis] with at least some adenomas.”
    ▪ “Known deleterious MUTYH mutation(s) in the family”
  o SPS clinical diagnostic criteria:
    ▪ “At least 5 serrated polyps (includes hyperplastic polyps, sessile serrated adenomas/polyps, and traditional serrated adenomas) proximal to the sigmoid colon with 2 or more of these being >10mm”
“Any number of serrated polyps proximal to the sigmoid colon in an individual who has a first-degree relative with serrated polyposis.”

“Greater than 20 serrated polyps of any size, but distributed throughout the colon”

Footnotes:

“When colonic polyposis is present in a single person with a negative family history, consider testing for a de novo APC mutation; if negative, follow with testing of MUTYH (targeted testing for the two common northern European founder mutations c.536A>G and c.1187G>A may be considered first followed by full sequencing if biallelic mutations are not found). When colonic polyposis is present only in siblings, consider recessive inheritance and test for MUTYH first. Order of testing for APC and MUTYH is at the discretion of the clinician. ”

“MUTYH genetic testing is not indicated based on a personal history of desmoid tumor, hepatoblastoma, or cribriform-morular variant of papillary thyroid cancer.”

“Siblings of a patient with MAP are recommended to have site-specific genetic testing for the familial biallelic mutations. Children of an affected parent with MAP are recommended to have site-specific genetic testing for the familial mutation/s. If positive for one MUTYH mutation, full sequencing of MUTYH is recommended. Full sequencing of MUTYH also may be considered in an unaffected parent when the other parent has MAP. If the unaffected parent is found to not be heterozygous for a MUTYH mutation, genetic testing in children is not necessary. If he or she is found to have a MUTYH mutation, testing for the familial mutations in the children is recommended.”
“It is important to note that de novo mutations can occur in APC or MUTYH. Thus, when colonic polyposis is present in an individual with a negative family history, consideration should be given to genetic testing of APC, followed by testing of MUTYH if no APC mutation is found.”

- All recommendations are category 2A.

- Evidence-based guidelines from the American College of Gastroenterology (ACG, 2009) state:9 “Patients with classic FAP, in whom genetic testing is negative, should undergo genetic testing for bi-allelic MUTYH mutations. Patients with 10 - 100 adenomas can be considered for genetic testing for attenuated FAP and if negative, MUTYH associated polyposis” [Grade 2C: Weak recommendation, low-quality or very low-quality evidence].

Criteria
MUTYH Known Familial Mutation Analysis

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  - No previous genetic testing for known MUTYH family mutation(s), AND

- Diagnostic or Predisposition Testing:1,2
  - Two known MUTYH mutations in a sibling, or
  - Both parents with one or two known MUTYH mutations, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

MUTYH Targeted Mutation Analysis for Y179C and G396D Mutations

- Genetic Counseling
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  - No previous MUTYH testing, and
  - No mutation detected on APC gene testing, if performed, AND

- Individual is of Northern European descent, AND

- Diagnostic Testing for Symptomatic Individuals:2,10
  - Clinical findings:
    - > 10 cumulative adenomas, or
At least two adenomas, AND

- At least 5 serrated polyps proximal to the sigmoid colon (2 or more of >10mm), or
- > 20 serrated polyps of any size, but distributed throughout the colon, AND
  - Recessive pattern of inheritance (e.g. family history positive for only an affected sibling), OR

Testing for Presymptomatic/Asymptomatic Individuals: 1,2

- Reproductive partner of a person with MAP (to determine if children at risk), AND
- Rendering laboratory is a qualified provider of service per the Health Plan policy.

### MUTYH Sequencing

- Genetic Counseling
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Testing:
  - No previous MUTYH full sequencing, and
  - Two mutations NOT identified through MUTYH targeted mutation analysis (Y179C and G396D) if performed, and
  - No mutation detected on APC gene testing, if performed, AND
- Diagnostic Testing for Symptomatic Individuals: 2,10
  - Clinical findings:
    - > 10 cumulative adenomas, or
    - At least two adenomas, AND
      - At least 5 serrated polyps proximal to the sigmoid colon (2 or more of >10mm), or
      - > 20 serrated polyps of any size, but distributed throughout the colon, AND
    - Recessive pattern of inheritance (e.g. family history positive for only an affected sibling), OR
- Testing for Presymptomatic/Asymptomatic Individuals: 1,2
o Reproductive partner of a person with MAP (to determine if children at risk), AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

MUTYH Deletion/Duplication Analysis

• Genetic Counseling
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o MUTYH full sequencing performed, and
  o No mutations or only one mutation detected in MUTYH through any previous testing (founder mutation panel or full gene sequencing), and
  o No mutation detected on APC gene testing, if performed, AND

• Diagnostic Testing for Symptomatic Individuals:2,10
  o Clinical findings:
    ▪ > 10 cumulative adenomas, or
    ▪ At least two adenomas, AND
      • At least 5 serrated polyps proximal to the sigmoid colon (2 or more of >10mm), or
      • > 20 serrated polyps of any size, but distributed throughout the colon, AND
  o Recessive pattern of inheritance (e.g. family history positive for only an affected sibling), OR

• Testing for Presymptomatic/Asymptomatic Individuals:1,2
  o Reproductive partner of a person with MAP (to determine if children at risk), AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


Myeloma Prognostic Risk Signature (MyPRS)

MOL.TS.237.A
v2.0.2018

Procedures addressed

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What is MyPRS

Definition

The Myeloma Prognostic Risk Signature (MyPRS®) (Signal Genetics™) has been developed to estimate the underlying activity of disease progression, in patients diagnosed with active MM. The test may be used as a potentially useful risk stratification tool to predict treatment response to chemotherapy, predict risk of survival and relapse, and tailor therapy selection. Specifically, MyPRS may identify a high-risk patient group for disease progression based on the expression levels of 70 selected genes measured at baseline. It may be helpful to stratify patients into high-, high risk-borderline, low-risk borderline, and low-risk categories to optimize individual treatment.  

- Multiple myeloma (MM) is a malignant and often incurable hematological cancer, characterized by the abnormal and uncontrolled proliferation of plasma cells in bone marrow, leading to impaired hematopoiesis and production of monoclonal immunoglobulin (Ig). The disease is responsible for about 1% of all cancers worldwide and 10 to 15% of all hematological cancers. MM usually affects older adults (median age of onset is 71 and 74 years for men and women, respectively). For the period between 2009 and 2010, the relative world-wide 5-year survival rate was approximately 45%.

- Clinical features of MM include anemia (73%), bone pain (58%), fatigue (32%), and unusual weight loss (25%). Diagnostic laboratory and clinical assessments include hypercalcemia, kidney dysfunction, anemia, and bone lesions. In general, patients are treated with autologous stem-cell transplantation (ASCT), along with supportive measures, such as pain therapy, administration of bisphosphonates, and irradiation of skeletal/extramedullary lesions.  

- A growing body of research suggests specific genetic lesions play an important role in the tumor biology of MM. Furthermore, the high number of chromosomal aberrations and multiple changes in gene expression of these lesions has
demonstrated that the underlying genetic features of MM tumor cells are responsible for the significant degree of clinical heterogeneity typically observed in this disease. Several molecular subtypes, each with a unique path of pathogenesis and clinical presentation, have also been identified. The inherent molecular heterogeneity of the disease is believed to translate into highly variable treatment responses and survival times (ranging from a few months to 15 years or more). Given the considerable heterogeneity of associated outcomes, various prognostic risk factors specific to MM have been identified to predict the course of disease, define individualized treatment strategies, predict survival, and enhance overall therapeutic decision making.

- Conventional cytogenetic methods, such as karyotyping and fluorescence in situ hybridization (FISH), are used in clinical practice to assess disease prognosis and stratify MM patients based on recurrent chromosomal changes. Risk stratification is intended to ensure patients receive proper treatment, depending on disease severity. One available risk stratification strategy is the evidence-based algorithm, the Mayo Stratification of Myeloma and Risk-Adapted Therapy (mSMART), used to inform treatment decisions for patients with newly diagnosed disease. However, given the heterogeneity of MM, conventional prognostic methods may not accurately estimate risk.

**Test information**

- According to Signal Genetics, the MyPRS test uses the Affymetrix GeneChip® 3000Dx v.2 System, a whole-genome microarray platform, and requires at least 20,000 CD138+ plasma cells in order to obtain sufficient genetic material for gene expression analysis.

- The MyPRS gene expression profiling model consists of a continuous gene score that is a linear combination of the 70 genes along with a cutoff, such that patients with a score greater than the cutoff are categorized as high risk and otherwise low risk for disease progression.

- The MyPRS prognostic score has the ability to predict a patient’s likely event-free survival (EFS) and overall survival (OS) at the time of diagnosis or after relapse. The algorithm used to develop this prognostic score was based on mathematical models using microarray technology and multivariate analysis of independent patient cohorts over 8 years of follow-up. Results of the model indicate that on a risk score of 0 (lowest likelihood of risk; good prognosis) to 100 (highest likelihood of risk; poor prognosis), a cut-off point of 45.2 discriminates between low and high risk patients.

- The test also provides results of a molecular subtype (7-class molecular subtype taxonomy), each associated with unique genetic lesions, altered genes, and outcome variation.

- Patients are provided results of virtual karyotyping to predict cytogenic abnormalities associated with MM, which is based on an 816-gene algorithm using
gene expression data, and validated against multiple traditional cytogenic techniques.\textsuperscript{6}

- In November 2016, Quest Diagnostics purchased MyPRS assets from Signal Genetics.\textsuperscript{10}

### Guidelines and evidence

- The National Comprehensive Cancer Network (NCCN, 2017) Clinical Practice Guidelines stated the following regarding gene expression profiling (GEP):
  - “GEP is a powerful and fast tool with the potential to provide additional prognostic value to further refine risk stratification, help therapeutic decisions, and inform novel drug design and development.”\textsuperscript{11}
  - “The NCCN Panel unanimously agreed that although GEP is not currently routinely used in clinical practice during diagnostic workup, GEP is a useful tool and may be helpful in selected patients to estimate the aggressiveness of the disease and individualize treatment.”
  - The NCCN Panel does not make any explicit recommendations for its use in its diagnostic and treatment pathways for cases of MM.

- There is insufficient evidence in the peer-reviewed literature to draw definitive conclusions regarding the analytical validity, clinical validity, and clinical utility of the MyPRS test to accurately provide prognostic risk stratification among patients who are newly diagnosed with MM or who have relapsed following treatment.\textsuperscript{12-15}

  - The evidence base mostly consists of retrospective studies evaluating small numbers of patients that evaluated the strength of the association between the MyPRS score with various survival measures, including post relapse survival, overall survival, and progression-free survival. Although the available studies reported significant associations between MyPRS and survival measures (patients with high MyPRS scores may be at increased risk of relapse and death), with study authors concluding that MyPRS has value as a risk stratification tool, the quality of the overall evidence is low given the retrospective study designs across the evidence base, and the lack of reported accuracy measures, including sensitivity, specificity, PPV, NPV, and clinical utility values. Furthermore, there is little to no evidence regarding the comparative accuracy of MyPRS with FISH testing or MyPRS with karyotyping. It is unknown if MyPRS can be an adequate substitute for FISH testing in patients with MM as part of the routine workup of the disease.
Future prospective studies, allocating patients to therapies determined to be most effective based on MyPRS score, with adequate sample sizes, using gold standard diagnostic and/or prognostic measures, are necessary to elucidate its role as an adjunct to existing risk stratification measures or as a stand-alone test. Well-designed clinical utility studies are also needed to assess whether the MyPRS test leads to improved therapeutic clinical decision-making and improved patient outcomes.

Criteria

- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  - In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

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**Myoclonic Epilepsy with Ragged Red Fibers (MERRF)**

**Procedures addressed**

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**What is MERRF**

**Definition**

Myoclonic Epilepsy with Ragged Red Fibers (MERRF) is a multisystem mitochondrial disease.¹

- MERRF typically presents with myoclonus (brief, involuntary twitching of a muscle or a group of muscles), followed by generalized epilepsy, ataxia (lack of coordination of muscle movements), weakness, and dementia.¹ Ragged red fibers (RRF) are identified on muscle biopsy pathology.¹
  - Other common findings include hearing loss, short stature, optic atrophy, and cardiomyopathy with Wolff-Parkinson-White syndrome (a syndrome in which there is extra electrical connection in the heart at birth causing rapid heartbeat). Occasionally pigmentary retinopathy and lipomatosis are observed.¹
  - Most cases present in childhood after normal early development.¹

- MERRF is caused by mutations in the mitochondrial DNA (mtDNA) and follows maternal inheritance. This means that a female who carries the mtDNA point mutation will pass it on to all of her children. A male who carries the mtDNA mutation cannot pass it on to his children.¹,²

- For all mtDNA mutations, clinical expressivity depends on the three following factors:
  - The relative abundance of mutant mtDNA, or mutational load (heteroplasmy)
  - The organs and tissues in which the mutant mtDNA is found (tissue distribution), and

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The vulnerability of each tissue to impaired oxidative metabolism (threshold effect).

- The estimated prevalence of MERRF is approximately 0.25-1.5/100,000 individuals.\(^1\)
- Management is usually palliative. Certain antiepileptic drugs, such as valproic acid, should be avoided as they may cause secondary carnitine deficiency or can be used with L-carnitine supplementation.\(^1\)
- At-risk individuals may also benefit from clinical assessment to initiate baseline evaluations (neurology, cardiology, ophthalmology, and audiology) and potential intervention prior to exhibiting clinical manifestations.\(^1\)

**Test information**

- The investigation and diagnosis of patients with mitochondrial disease often necessitates a combination of techniques including muscle histocytochemistry, biochemical assessment and molecular genetic studies along with clinical assessment. Any molecular genetic test for a mtDNA mutation should ideally be directed by the clinical phenotype and results of these other investigations.\(^3\)
- MERRF Mutation Panel: Heteroplasmic mutations in the mtDNA genes, MT-TK, MT-TL1, MT-TF, MT-TI, and MT-TP cause MERRF. Mutations in the mtDNA genes MT-TH, MT-TS1, MT-TS2, cause MELAS/MERRF overlap syndrome.
  - Approximately 90% of cases of MERRF are due to MT-TK mutations. 80% of MERRF cases are the result of a specific genetic change, m.8344A>G (formerly A8344G) in MT-TK.\(^1,2,4\)
    - Three additional MT-TK mutations, m.8356T>C, m.8363G>A, and m.8361G>A, are present in an additional 10% of affected individuals. These three mutations can also be associated with other mitochondrial or genetic conditions.\(^1\)
  - Detection rate of the four-mutation panel is about 90%.\(^1\)
  - “Sequence analysis / scanning for pathogenic variants is used to detect pathogenic variants throughout mtDNA and is not specific for MERRF. The overall variant detection rate for MERRF by scanning/sequence analysis of mtDNA is 90%-95%.”\(^1\)
- Due to its ability to simultaneously sequence the entire mtDNA and measure heteroplasmy at each position, next generation sequencing (NGS) is an option for assessing MERRF and overlapping syndromes. However, certain targeted mutation analyses can estimate heteroplasmy. Typically, Sanger sequence analysis will miss heteroplasmy below 20%.
If genetic testing is negative in a blood sample in a person with symptoms of MERRF, testing can be done on other specimens. Typically this is done when the phenotype is highly suggestive of presence of a MERRF mutation or when there is a need to assess reproductive risk.

- Muscle may be considered as a secondary tissue since it is clinically involved as evidenced by Ragged Red Fibers. Muscle biopsy allows enzymatic analysis of the electron transport chain, light and ultra structural microscopy, and mtDNA copy number analysis—all of which may provide highly useful information. However, the invasiveness and procedural costs associated with a muscle biopsy are factors to consider.

- Genetic testing can also be done on skin fibroblasts, urinary sediment, or buccal mucosa. If cultured fibroblasts are used, measures such as limited passaging and uridine supplementation should be taken to reduce selection against mutant genotypes that may lead to skewed heteroplasmy.

Guidelines and evidence

- No specific evidence-based U.S. testing guidelines were identified.
- Case reports and a limited number of case series are the primary evidence base available for the diagnosis of mitochondrial disease.5-7
- The Mitochondrial Medicine Society developed consensus recommendations using the Delphi method and published them in 2015.8

- Recommendations for DNA testing
  - “Massively parallel sequencing/NGS of the mtDNA genome is the preferred methodology when testing mtDNA and should be performed in cases of suspected mitochondrial disease instead of testing for a limited number of pathogenic point mutations.”
  - “Patients with a strong likelihood of mitochondrial disease because of a mtDNA mutation and negative testing in blood, should have mtDNA assessed in another tissue to avoid the possibility of missing tissue-specific mutations or low levels of heteroplasmy in blood; tissue-based testing also helps assess the risk of other organ involvement and heterogeneity in family members and to guide genetic counseling.”
  - “Heteroplasmy analysis in urine can selectively be more informative and accurate than testing in blood alone, especially in cases of MELAS due to the common m.3243 A>G mutation.”
“When considering nuclear gene testing in patients with likely primary mitochondrial disease, NGS methodologies providing complete coverage of known mitochondrial disease gene is preferred. Single-gene testing should usually be avoided because mutations in different genes can produce the same phenotype. If no mutation is identified via known NGS panels, then whole exome sequencing should be considered.”

- The European Federation of Neurological Sciences (2009) provided molecular diagnostic consensus-based guidelines based on literature reviews: “If the phenotype suggests syndromic mitochondrial disease due to mtDNA point mutations (MELAS, MERRF, NARP, LHON) DNA-microarrays using allele-specific oligonucleotide hybridisation, real-time-PCR or single-gene sequencing are indicated.”

- The clinical utility of genetic testing for MERRF was described by a workshop of the National Institute of Neurological Disorders and Stroke (2008):
  - “The diagnosis of mitochondrial diseases is complicated by their heterogeneous presentations and by the lack of screening procedures or diagnostic biomarkers that are both sensitive and specific. The workshop panelists explained that diagnosis is often a lengthy process beginning with a general clinical evaluation followed by metabolic screening and imaging and finally by genetic tests and more invasive biochemical and histological analyses. The identification of known mitochondrial mutations in tissue has greatly aided diagnosis. However, even when clinical features and family history strongly suggest mitochondrial disease, the underlying genetic mutation can elude detection, and there is no current screening procedure that would be practical for all cases of suspected mitochondrial disease.”
  - Initial screening includes testing lactate and CSF protein levels, muscle biopsy, EEG, ECG, and MRI. “It is important to note that biochemical abnormalities may not be present during periods when the mitochondrial disease is quiescent/dormant.”

- The Clinical Molecular Genetics Society of UK (2008) provided practice-based guidelines for the molecular diagnosis of mitochondrial disease: “In cases with strong clinical evidence, testing should begin with checking for the common mutation, m.8344A>G. Subsequent testing for other mutations, such as m.8356T>C, may be indicated in cases with a strong clinical indication of MERRF.”

Criteria

Known MERRF Familial Mutation Testing

- Genetic Counseling
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing
  o No previous genetic testing in the individual for MERRF,** and
  o MERRF pathogenic variant identified in matrilineal relative, AND

• Predictive Testing for Asymptomatic Individual:
  o 18 years of age or older, or
  o Under the age of 18 years, and
    ▪ Presymptomatic screening for Wolff-Parkinson-White is being considered, OR

• Diagnostic Testing for Symptomatic Individual:
  o Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of MERRF, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

MERRF Targeted Mutation Analysis

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous genetic testing for MERRF,** and
  o No known MERRF pathogenic variants in the family, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of MERRF, and
  o Genetic testing is needed to confirm the diagnosis, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

Whole mtDNA Sequencing

• Genetic Counseling
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Criteria for MERRF targeted mutation analysis is met, AND

• No pathogenic variants identified in the MERRF targeted mutation analysis, AND
- Paternal transmission has been ruled out

** Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

References


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What is NETest

Definition

Neuroendocrine tumors (NETs) are a group of tumors that originate from epithelial cells with neuroendocrine variances; gastroenteropancreatic NETs are a subgroup of NETs that develop from the gastrointestinal tract.¹

- Detection of these lesions is often delayed due to the heterogeneous cellular make-up and inconspicuous symptomology.¹
- The prevalence and incidence of gastroenteropancreatic neuroendocrine tumors (NETs) have been increasing.¹
- Currently, there is a lack of specific blood markers for NET detection. Measurement of the neuroendocrine secretory peptide Chromogranin A (CgA) is often used, but is characterized by flaws since it is a single value, non-specific, and assay data are highly variable.
- As a result, there is greater interest in the discovery of effective biomarkers, such as the NETest, to evaluate disease risk and new therapies targeting gastroenteropancreatic NET.²⁻⁶

Test information

- NETest is a noninvasive blood test designed to assist in identifying activity of neuroendocrine tumor disease.
- This test examines the expression of 52 genes, which can be used to identify active disease and provide information about the biology of the tumor cell.
- As an adjunct to standard clinical assessment, the NETest provides an assessment of treatment responses in patients with NETs.²⁻⁶
• The algorithm measures the activity of RNA gene expression and calculates a risk score. Risk scores range from 0-100%. The higher the score, the higher the risk of active disease at the time of testing. The following categories have a sensitivity of 95.7%:2-6
  o Very low (≤13.4%) exhibit minimal risk for disease activity.
  o Low (13.4% - 43.4%) are classified as low active or stable disease
  o High (>43.4%) are classified as highly active disease.

Guidelines and evidence
• There are currently no national guidelines that recommend the use of NETest in the screening of diagnosis of neuroendocrine tumors.

• Preliminary evidence of studies evaluating its clinical validity suggests that the NETest may be comparable to currently available alternatives, demonstrating relatively good measures of test accuracy. In the studies reporting test accuracy metrics, the NETest demonstrated consistent and reasonably high sensitivity, specificity, positive predictive value and negative predictive values. However, studies are hampered by several limitations, which weaken the overall quality of the evidence base. These include few numbers of studies, small patient populations, lack of generalizability to broader real-world patient populations in clinical practice, lack of blinding, and heterogeneity in patient clinical characteristics across comparator groups. In addition, studies regarding clinical validity alone are insufficient to provide support for a recommendation that the NETest can be used to guide clinician decisions regarding treatment responses among patients with NET. Furthermore, given the lack of trials evaluating clinical utility, the evidence is insufficient to determine if the NETest demonstrates clinical utility. It remains uncertain if use of the NETest results in clinically meaningful benefits measured by patient-relevant outcomes, beyond that being provided by standard clinical assessments. Additional studies definitively demonstrating clinical utility in well-designed clinical trials with proper patient-relevant outcomes are necessary to confirm that the NETest may help clinicians to make better informed treatment decisions.7-11

Criteria
• This test is considered investigational and/or experimental.
  o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical
management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.

- In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References


2. Understanding the NETest results. NETest website. Available at: http://www.wrenlaboratories.com/patient/understanding-the-netest-results


5. Testing overview. NETest website. Available at: http://wrenlaboratories.com/provider/testing-overview/


Neurogenic Muscle Weakness, Ataxia, and Retinitis Pigmentosa (NARP)

Procedures addressed

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What is NARP

Definition

Neurogenic muscle weakness, Ataxia, and Retinitis Pigmentosa is a multisystem mitochondrial disease. NARP is characterized by proximal neurogenic muscle weakness with sensory neuropathy, ataxia, learning difficulties, and pigmentary retinopathy. Most cases present in childhood with ataxia and learning difficulties. Seizures may also be present.

- NARP is caused by mutations in the mitochondrial DNA (mtDNA) and follows maternal inheritance. This means that a female who carries the mtDNA mutation at high mutation load will typically pass it on to all of her children. A male who carries the mtDNA mutation cannot pass it on to his children.
- For all mtDNA mutations, clinical expressivity depends on the three following factors:
  - The relative abundance of mutant mtDNA, mutational load (heteroplasm) 
  - The organs and tissues in which the mutant mtDNA is found (tissue distribution), and
  - The vulnerability of each tissue to impaired oxidative metabolism (threshold effect).
- The exact prevalence of NARP is unknown.
- Management of NARP is generally supportive. Regular neurologic, ophthalmologic, and cardiologic screenings are recommended for affected individuals. Anti-epileptic drugs that affect the mitochondrial respiratory chain should be avoided, as they may
cause secondary carnitine deficiency or can be used with L-carnitine supplementation.¹

- At-risk individuals may also benefit from clinical assessment to initiate baseline evaluations (neurology, cardiology, ophthalmology, and audiology) and potential intervention prior to exhibiting clinical manifestations.¹

**Test information**

- The investigation and diagnosis of patients with mitochondrial disease often necessitates a combination of techniques including muscle histocytochemistry, biochemical assessment, and molecular genetic studies along with clinical assessment. Any molecular genetic test for a mtDNA mutation should ideally be directed by the clinical phenotype and results of these other investigations.²

- **NARP Targeted Mutation Analysis**
  - m.8993T>G (T8993G) and m.8993T>C (T8993C) in MT-ATP6 cause ~50% of cases of NARP.¹
  - If negative, whole genome sequencing of mitochondrial DNA can detect more rare mutations associated with NARP, but does not significantly increase the detection rate over testing for the common two mutations¹

- While genetic test results alone cannot predict the exact course or phenotype of the disease, severity does correlate with mutation load.¹,⁴ The clinical course for mitochondrial diseases is subject to the concepts of heteroplasmy, tissue distribution, and threshold effect.¹,³

- Due to its ability to simultaneously sequence the entire mtDNA and measure heteroplasmy at each position, next generation sequencing (NGS) is an attractive option for assessing NARP and overlapping syndromes. However, certain targeted mutation analyses can estimate heteroplasmy. Typically, Sanger sequence analysis will miss heteroplasmy below 20%.

- Genetic testing can also be done on skin fibroblasts, urinary sediment, or buccal mucosa.¹ If cultured fibroblasts are used, measures such as limited passaging and uridine supplementation should be taken to reduce selection against mutant genotypes that may lead to skewed heteroplasmy.

- If genetic testing is negative in a blood sample in a person with symptoms of NARP, testing can be done on other specimens. Typically this is done when the phenotype is highly suggestive of presence of a NARP mutation or when there is a need to assess reproductive risk for offspring with higher mutant load and risk for developing Leigh disease.

  - Muscle may be considered as a secondary tissue. Muscle biopsy allows enzymatic analysis of the electron transport chain, light and ultrastructural microscopy, and mtDNA copy number analysis—all of which may provide highly useful information.
• However, muscle (and/or liver) biopsies are often not necessary and should be avoided when possible due to their invasive nature. Biopsies should only be considered when the diagnosis cannot be confirmed with DNA testing of other more accessible tissues.

Guidelines and evidence

• No specific evidence-based U.S. testing guidelines were identified.
• Case reports and a limited number of case series are the primary evidence base available for the diagnosis of mitochondrial disease.4-6
• The Mitochondrial Medicine Society developed consensus recommendations using the Delphi method and published them in 2015.7

  o Recommendations for DNA testing
    ▪ “Massively parallel sequencing/NGS of the mtDNA genome is the preferred methodology when testing mtDNA and should be performed in cases of suspected mitochondrial disease instead of testing for a limited number of pathogenic point mutations.”
    ▪ “Patients with a strong likelihood of mitochondrial disease because of a mtDNA mutation and negative testing in blood, should have mtDNA assessed in another tissue to avoid the possibility of missing tissue-specific mutations or low levels of heteroplasmy in blood; tissue-based testing also helps assess the risk of other organ involvement and heterogeneity in family members and to guide genetic counseling.”
    ▪ “Heteroplasmy analysis in urine can selectively be more informative and accurate than testing in blood alone, especially in cases of MELAS due to the common m.3243 A>G mutation.”
    ▪ “When considering nuclear gene testing in patients with likely primary mitochondrial disease, NGS methodologies providing complete coverage of known mitochondrial disease gene is preferred. Single-gene testing should usually be avoided because mutations in different genes can produce the same phenotype. If no mutation is identified via known NGS panels, then whole exome sequencing should be considered.”

• A workshop of the National Institute of Neurological Disorders and Stroke (2008)3 summarizes:
  o “The diagnosis of mitochondrial diseases is complicated by their heterogeneous presentations and by the lack of screening procedures or diagnostic biomarkers that are both sensitive and specific. The workshop panelists explained that diagnosis is often a lengthy process beginning with a general clinical evaluation followed by metabolic screening and imaging and finally by genetic tests and more invasive biochemical and histological analyses. The identification of known mitochondrial mutations in tissue has greatly aided diagnosis. However, even
when clinical features and family history strongly suggest mitochondrial disease, the underlying genetic mutation can elude detection, and there is no current screening procedure that would be practical for all cases of suspected mitochondrial disease."

- The Clinical Molecular Genetics Society (CMGS) of the United Kingdom (2008)\(^2\) practice-based guidelines for the molecular diagnosis of mitochondrial disease state that: “For routine referrals for NARP, presence of T8993G and T8993C mutations should be investigated.”

- The European Federation of Neurological Sciences (2009)\(^8\) evidence-based molecular diagnostic guidelines state: “If the phenotype suggests syndromic mitochondrial disease due to mtDNA point mutations (MELAS, MERRF, NARP, LHON) DNA-microarrays using allele-specific oligonucleotide hybridisation, real-time-PCR or single-gene sequencing are indicated.”

**Criteria**

**Known NARP Familial Mutation Testing**

- Genetic Counseling
  - Pre- and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing
  - No previous genetic testing in the individual for NARP\(^*\), and
  - NARP pathogenic variant identified in matrilineal relative, AND

- Predictive Testing for Asymptomatic Individual:
  - 18 years of age or older, or
  - Under the age of 18 years, and
    - Screening for learning disabilities, retinitis pigmentosa, and/or ataxia is being considered, OR

- Diagnostic Testing for Symptomatic Individual:
  - Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of NARP, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

**NARP Targeted Mutation Analysis**

- Genetic Counseling
• Pre- and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous genetic testing for NARP**, and
  o No known NARP pathogenic variants in the family, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Clinical exam and/or biochemical testing suggestive, but not confirmatory, of a diagnosis of NARP, and
  o Genetic testing is needed to confirm the diagnosis, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

** Whole mtDNA Sequencing

• Genetic Counseling
  o Pre- and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Criteria for NARP targeted mutation analysis is met, AND

• No pathogenic variants identified in the NARP targeted mutation analysis, AND

• Paternal transmission has been ruled out

** Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

References


Niemann-Pick Disease Types A and B Testing

Procedures addressed

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What is Niemann-Pick disease types A and B

Definition

Niemann-Pick disease is a genetic disorder caused by an inability to process lipids (fats), which results in a toxic buildup of lipids in some organs.¹-³

- Two types of Niemann-Pick disease are caused by a deficiency of the acid sphingomyelinase enzyme:
  - Type A, also called the "neurological" or "neuronopathic" type, causes symptoms beginning in infancy. These include an enlarged liver and spleen (hepatosplenomegaly), psychomotor impairment with neurologic deterioration, interstitial lung disease, and eventually a classic cherry-red spot of the retina. Affected individuals usually do not survive beyond childhood.¹-³
  - Type B, also called the "non-neurological" or "non-neuronopathic" type, causes some symptoms similar to type A, but symptoms are usually milder and begin later. Additional symptoms include hyperlipidemia (high fat levels in blood) and thrombocytopenia (low platelets). Affected individuals can survive to adulthood.¹-³
- The SMPD1 gene encodes the acid sphingomyelinase (ASM) enzyme. Gene mutations in the SMPD1 gene lead to reduced or absent sphingomyelinase enzyme activity, causing the symptoms of Niemann-Pick disease.¹-³
Niemann-Pick disease is suspected when a patient presents with hepatosplenomegaly, interstitial lung disease, and depending on the subtype, neurological symptoms in infancy or abnormal blood findings. However, a diagnosis cannot be made clinically.

When Niemann-Pick disease is suspected, acid sphingomyelinase enzyme activity testing should be performed first. People with Niemann-Pick disease type A or B usually have less than 10% of normal ASM activity compared to healthy individuals. Measuring ASM enzyme activity in peripheral blood lymphocytes or cultured skin fibroblasts is a reliable way to confirm a suspected case of Niemann-Pick disease. However, false-negative and inconclusive results are possible. In such cases, genetic testing may be useful to resolve a diagnosis.

About 1 in 250,000 people have Niemann-Pick disease. Type A is more common in persons of Ashkenazi Jewish descent than in the general population. In the Ashkenazi Jewish population, the frequency of Niemann-Pick disease is 1 in 40,000.

Niemann-Pick disease is an autosomal recessive disorder. An affected individual must inherit SMPD1 gene mutations from both parents.

- Individuals who inherit only one mutation are called carriers. Carriers do not show symptoms of Niemann-Pick disease, but have a 50% chance of passing on the mutation to their children.
- Two carriers of Niemann-Pick disease have a 25% chance of having a child with the disorder.
- Prenatal diagnosis for at-risk pregnancies can be performed by molecular genetic testing (if the mutations in both parents are known).

Individuals at increased risk to have a child with Niemann-Pick disease should routinely be offered carrier screening. This includes those with:

- Ashkenazi Jewish ancestry (1 in 90 carrier risk)
- A family history of Niemann-Pick disease (regardless of ethnicity)
- A partner who is a known carrier of Niemann-Pick disease (or affected with the milder type)

Test information

- **SMPD1 Mutation Analysis** tests for four of the most common SMPD1 gene mutations.
  - Three mutations - R496L, L302P, fsP330 - account for 97% of all cases of Niemann-Pick disease type A in Ashkenazi Jewish people.
The fourth mutation - deltaR608 - is a common cause of Niemann-Pick disease type B in people of North African descent.\(^3\)

Carrier screening by SMPD1 mutation panel for Niemann-Pick disease is widely available as part of an “Ashkenazi Jewish Panel” that includes several other genetic disease that are more common in this population. (See Ashkenazi Jewish Carrier Screening.)

- **SMPD1 Sequencing** analyzes the entire coding region of the SMPD1 is available to detect less common mutations that cannot be detected on a common mutation analysis panel. SMPD1 sequencing detects more than 95\% of all SMPD1 mutations.\(^3\)

- **SMPD1 Deletion/Duplication Analysis** is available to detect large gene rearrangements that cannot be detected by sequencing. However, the frequency of such mutations is unknown.\(^3\)

- **SMPD1 Known Familial Mutation Testing** can be performed for at-risk relatives when the familial mutation is known and is not one of the common mutations.\(^3\)

### Guidelines and evidence

- Professional guidelines generally support Niemann-Pick disease carrier screening for those at increased risk.\(^4,5\)

- Consensus guidelines from the American College of Obstetricians and Gynecologists (ACOG, 2009) address carrier screening and prenatal diagnosis for Niemann-Pick disease:
  
  o “Individuals with a positive family history of one of these disorders [including Niemann-Pick disease] should be offered carrier screening for the specific disorder and may benefit from genetic counseling.”

  o Carrier screening for Ashkenazi Jewish people is routinely recommended for some disorders (i.e., Tay-Sachs, Canavan, cystic fibrosis, familial dysautonomia). However, for testing of a group of other disorders more common in this population (including Niemann-Pick disease), ACOG simply states: “Individuals of Ashkenazi Jewish descent may inquire about the availability of carrier screening for other disorders.”

  o “If it is determined that this individual [an Ashkenazi Jewish descent partner] is a carrier, the other partner should be offered screening.”

  o “When both partners are carriers of one of these disorders, they should be referred for genetic counseling and offered prenatal diagnosis.”

- Consensus guidelines from the American College of Medical Genetics (2008) recommend routine carrier screening for a group of disorders that includes Niemann-Pick when at least one member of the couple is Ashkenazi Jewish and that couple is pregnant or planning pregnancy.\(^5\)
• No evidence-based US diagnostic testing guidelines have been identified.
• A 2015 expert-authored review recommends the following testing strategy for diagnosis of an affected person:\(^3\)

  o “The diagnosis of ASM deficiency is established by detection of either biallelic pathogenic variants in SMPD1 on molecular genetic testing or residual ASM enzyme activity that is less than 10% of controls (in peripheral blood lymphocytes or cultured skin fibroblasts).”

  o Molecular testing approaches include single-gene testing and use of a multi-gene panel.

  o For individuals from populations in which common SMPD1 pathogenic variants occur (e.g., individuals of Ashkenazi Jewish background with a severe neurodegenerative form of the disease suggestive of NPD-A, individuals of North African descent with NPD-B, or individuals from Chile, Saudi Arabia, and Turkey):
    ▪ Perform targeted analysis for pathogenic variants.
    ▪ If targeted analysis does not identify both pathogenic variants in individuals from these populations, sequence analysis of SMPD1 is appropriate.

  o For individuals who are not in the populations discussed above:
    ▪ Perform sequence analysis.
    ▪ “If no or only one pathogenic variant is identified, consider gene-targeted deletion/duplication analysis.”

Criteria

Niemann Pick Type A or B Known Familial Mutation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous genetic testing for Niemann Pick A or B, AND

• Diagnostic and Predisposition Testing:
  o Niemann Pick A or B family mutation identified in biologic relative(s), OR

• Prenatal Testing:
  o Niemann Pick A or B mutation identified in both biologic parents, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Niemann Pick A or B Targeted Mutation Analysis

• Genetic Counseling
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous genetic testing for Niemann Pick A or B
• Diagnostic Testing for Symptomatic Individuals:
  o Measurement of acid sphingomyelinase (ASM) enzyme activity in peripheral blood lymphocytes or cultured skin fibroblasts (in symptomatic individuals) with negative or equivocal result where suspicion of clinical diagnosis remains high, and
  o Hepatosplenomegaly, and/or
  o Evidence of interstitial lung disease on chest radiograph, and/or
  o Developmental Delay, and/or
  o Cherry Red Maculae, and/or
  o Hyperlipidemia, and/or
  o Thrombocytopenia, OR
• Predisposition/Carrier Testing for Presymptomatic/Asymptomatic Individuals:
  o Biologic relative(s) (1st degree) diagnosed with Niemann Pick A or B clinically, and no family mutation identified, or
  o Ashkenazi Jewish ancestry and intention to reproduce, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Niemann Pick A or B Sequencing

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o If Ashkenazi Jewish, common mutations have been tested and resulted negative
• Diagnostic Testing for Symptomatic Individuals:
• Measurement of acid sphingomyelinase (ASM) enzyme activity in peripheral blood lymphocytes or cultured skin fibroblasts (in symptomatic individuals) with negative or equivocal result where suspicion of clinical diagnosis remains high, and
  o Hepatosplenomegaly, and/or
  o Evidence of interstitial lung disease on chest radiograph, and/or
  o Developmental Delay, and/or
  o Cherry Red Maculae, and/or
  o Hyperlipidemia, and/or
  o Thrombocytopenia, OR

• Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  o Biologic relative(s) (1st degree) diagnosed with Niemann Pick A or B clinically, and no family mutation identified, and
  o If Ashkenazi Jewish, common mutations have been tested and resulted negative, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Niemann Pick A or B Deletion/Duplication Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous large rearrangement testing, and
  o Previous SMPD1 sequencing performed and no mutations found, and
  o No known familial mutation, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

References


Niemann-Pick Type C Testing

Procedures addressed

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What is Niemann-Pick Disease type C

Definition

Niemann-Pick Disease, type C (NPC) is a lipid storage condition that can present at any age, though the classic presentation is in mid-to-late childhood. Symptoms fall into one of three categories: visceral, neurological and psychological.¹

- The presentation of clinical symptoms at each stage is different: ²,³
  - Infants typically present with hypotonia and developmental delay, with or without lung and liver disease. Liver disease can be severe, resulting in the death of an infant in a few days to a few months.
  - Children with NPC exhibit progressive ataxia, vertical supranuclear gaze palsy (VSGP) and dementia.
  - Adults who develop NPC usually have an onset of progressive cognitive impairment or other psychiatric symptoms.
- There is wide variability with disease progression and survival rate, which can range from just a few days to, in rare circumstances, 60 years. Most individuals survive between 10-25 years.⁴
- Two genes have been associated with NPC: NPC1 and NPC2. The proteins of these genes are thought to work together in the cellular transport of cholesterol and other molecules. Most (90-95%) individuals with NPC have at least one identifiable
gene mutation in NPC1.\textsuperscript{5,6} Only 30 families have been found to have mutations in the NPC2 gene, making mutations in this gene rare (about 4\% of NPC cases).\textsuperscript{1,5,7}

- There have been over 200 mutations described that cause NPC.\textsuperscript{6} Genotype-phenotype correlation is difficult to determine as most individuals are compound heterozygotes; however, there has been observation of some alleles being associated with mild or severe disease.\textsuperscript{8-10}

- NPC is thought to have a prevalence of 1 in 120,000 livebirths.\textsuperscript{1} There are a few populations that have a founder effect, including French Acadians of Nova Scotia, Canada originally from Normandy France\textsuperscript{7}; individuals of Hispanic descent in the Upper Rio Grande valley of the United States\textsuperscript{7}; and a Bedouin group in Israel.

- NPC is inherited in an autosomal recessive inheritance pattern. Because NPC is recessive, individuals usually do not have other affected family members. Males and females are equally likely to be affected. When both parents are known carriers, there is a 1/4 (25\%) chance for each pregnancy to be affected. Preimplantation and prenatal genetic diagnosis are available for at-risk pregnancies.

- Recently, an NPC suspicion index has been presented as a way to identify individuals with a strong suspicion of NPC, versus those who may need further evaluation and those whose suspicion is low.\textsuperscript{11} This index comprises ranked assessments of visceral, neurological and psychiatric signs and symptoms that are specific to NPC, taking family history into account, to provide an NPC risk prediction score. Patients scoring ≥70 should be referred for immediate testing. Those scoring from 40-69 should be evaluated for further signs and symptoms of a differential diagnosis. Scores below 40 have a low suspicion of NPC.\textsuperscript{1,11}

- Once a diagnosis of NPC is suspected, diagnosis may include biochemical and/or genetic testing.

- Healthcare management after diagnosis includes treatment for current symptoms. This generally includes medications to prevent the onset of seizures, although treatment of liver disease, sleeping dysfunction or other symptoms should be considered as well. There is no definitive therapy available for NPC. Bone marrow transplantation (BMT), liver transplantation or the use of cholesterol lowering drugs did not prevent the progression of neurological disease.

**Test information**

- *Filipin biochemical testing for Niemann-Pick type C* involves demonstration of abnormal intracellular cholesterol homeostasis in cultured fibroblasts.\textsuperscript{7,12} Fibroblasts are cultured in an LDL-enriched medium, and then fixed and stained with a compound called “filipin”. To perform biochemical testing, filipin interacts with unesterified cholesterol to make specific cholesterol-filled complexes in ~80-85\% of cases.
  - When this testing indicates an individual is affected, sequence/mutation analysis should be considered.
Carrier testing is not available through biochemical testing, as there is overlap of enzyme activity between carriers and non-carriers.

The biochemical assay can be used for prenatal diagnosis if both mutations are not known.\(^7\)

- **NPC1 sequence analysis** can identify ~80-90% of mutations in the NPC1 gene.\(^13\)
- **NPC2 sequence analysis** identifies virtually 100% of mutations in the NPC2 gene.\(^13\)
- **NPC1 and NPC2 deletion/duplication analysis** is available clinically for individuals who test negative on sequence analysis.
- **NPC1 and NPC2 known familial mutations**: Once a disease-causing mutation has been identified, relatives of affected individuals can be tested. Because of the variability of age of onset and presenting symptoms, individuals undergoing carrier testing should be aware that they could be identified as carrying two mutant alleles, and thus affected. Individuals identified as carriers for NPC can have preimplantation or prenatal testing. Prenatal testing can be performed through mutation analysis on CVS or amniocytes if both parental mutations are known.\(^13\)

Guidelines and evidence

- Consensus-based diagnostic recommendations are available from the NP-C Guidelines Working Group (2012), an international, collaborative group of disease experts:\(^1\)
  - “Laboratory diagnostic tests for NP-C are complex and can be difficult to interpret due to a variety of methodological factors. Diagnostic testing to confirm NP-C, following screening and differential diagnosis, should therefore be conducted by, or in consultation with, regional or national care centers specializing in the diagnosis of inherited metabolic disorders.”
  - “The demonstration of impaired intracellular cholesterol transport by filipin staining in fibroblasts cultured from patient skin biopsies remains a key diagnostic test for NP-C.”

  - “In 80–85% of cases, fluorescence microscopic examination of NP-C positive cells typically reveals strongly fluorescent, cholesterol-filled perinuclear vesicles — the “classical” cholesterol storage pattern. Most other cases with a “variant biochemical phenotype” show a less pronounced, more variable cholesterol storage.”
  - “LDL-induced cholesteryl ester formation assays are no longer systematically used as a secondary biochemical test, as they are technically challenging (particularly in variant cases), costly and time-consuming.”
“Biochemical tests cannot be relied upon to identify heterozygote carriers of NP-C in whom filipin test findings may either appear normal or display mild abnormalities, with changes similar to those seen in “variant” cell lines.”

- Regarding genetic testing:
  - “NP-C is caused by autosomal recessive mutations in either of two genes, NPC1 (located to chromosome 18, q11–q12) or NPC2 (located to chromosome 14; q24.3).”
  - “Over 95% of NP-C patients have pathological NPC1 mutations, with approximately 4% of patients expressing disease-causing mutations in NPC2; the remaining patients appear to possess as yet unidentified gene mutations.”
  - “DNA sequencing should ideally be performed in parallel with filipin staining examinations, where possible. Significant advances have been made in genetic sequencing of NPC1 and/or NPC2 gene mutations, but it is not yet possible to replace filipin staining with DNA sequencing as the primary diagnostic method.”
  - “Gene testing should be undertaken in all newly diagnosed patients to:”
    - “allow safe prenatal diagnosis”
    - “expedite identification of eventual affected siblings”
    - “allow detection of carriers in blood relative”
    - “identify NPC2 patients who may be candidates for hematopoietic stem cell transplantation.”

### Criteria

**Niemann-Pick Disease Type C Known Familial Mutation Analysis**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Testing:
  - No previous genetic testing for Niemann-Pick C, AND
- Diagnostic and Predisposition Testing:
  - Niemann-Pick C family mutation identified in biologic relative(s), OR
- Carrier Testing:
  - Niemann-Pick C family mutation identified in biologic relative(s), OR
• Prenatal Testing:
  o Niemann-Pick C mutation identified in both biologic parents AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Niemann-Pick C Disease Sequencing

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o Biochemical testing performed on cultured skin fibroblasts showing abnormal intracellular cholesterol homeostasis, and
  o No previous genetic testing for Niemann-Pick C, AND
• Diagnostic Testing for Symptomatic Individuals:
  o Hepatosplenomegaly and/or liver failure, or
  o Central hypotonia or low muscle tone characterized by frequent falls and clumsiness, or
  o Ocular motor abnormalities, especially saccadic eye movements (SEM) and vertical supranuclear gaze palsy, or
  o Delayed or arrested speech development with or without cognitive impairment, or
  o Cerebellar ataxia, or
  o Seizures, or
  o Dystonia, or
  o Dysphagia, OR
• Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  o Biologic relative(s) (1\textsuperscript{st}, 2\textsuperscript{nd}, or 3\textsuperscript{rd} degree) diagnosed with NPC clinically, and no family mutation identified, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Niemann-Pick C Disease Deletion/Duplication Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o Biochemical testing performed on cultured skin fibroblasts showing abnormal intracellular cholesterol homeostasis, and
  o NPC1 and NPC2 sequencing performed and no mutations or only one mutation identified, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Hepatosplenomegaly and/or liver failure, or
  o Central hypotonia or low muscle tone characterized by frequent falls and clumsiness, or
  o Ocular motor abnormalities, especially saccadic eye movements (SEM) and vertical supranuclear gaze palsy, or
  o Delayed or arrested speech development with or without cognitive impairment, or
  o Cerebellar ataxia, or
  o Seizures, or
  o Dystonia, or
  o Dysphagia, OR

• Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
  o Biologic relative(s) (1st, 2nd, or 3rd degree) diagnosed with NPC clinically, and no family mutation identified, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


Non-Invasive Prenatal Testing

Procedures addressed

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<thead>
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<tr>
<td>Non-Invasive Prenatal Testing for Fetal Aneuploidy</td>
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<td>Non-Invasive Prenatal Testing for Fetal Aneuploidy with Risk Score</td>
<td>81507</td>
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<td>Non-Invasive Prenatal Testing for Fetal Aneuploidy with Risk Score</td>
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<tr>
<td>Non-Invasive Prenatal Testing for Fetal Chromosomal Microdeletions</td>
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What is a chromosome abnormality

Definition

A chromosome abnormality is any difference in the structure, arrangement, or amount of genetic material packaged into the chromosomes.¹

- Humans usually have 23 pairs of chromosomes. Each chromosome has a characteristic appearance that should be the same in each person.
- Chromosome abnormalities can lead to a variety of developmental and reproductive disorders. Common chromosome abnormalities that affect development include Down syndrome (trisomy 21), trisomy 18, trisomy 13, Turner syndrome, and Klinefelter syndrome.
- About 1 in 200 newborns has some type of chromosome abnormality and a higher percentage of pregnancies are affected but lost during pregnancy. About 6%-11% of stillbirths or neonatal deaths are associated with a chromosome abnormality.²³
• The risk of having a child with an extra chromosome, notably Down syndrome, increases as a woman gets older. Historically, invasive prenatal diagnosis was only offered to women over the age of 35. However, many babies with Down syndrome are born to women under 35. Prenatal screening for Down syndrome and certain other chromosome abnormalities is now routinely offered to all pregnant women. As a result, prenatal diagnosis is now an option for most pregnant women.

**Test information**

• Non-invasive prenatal testing (NIPT) is performed on a maternal plasma sample generally collected between 9-20 weeks’ gestation.

• Testing methodology relies on the presence of cell-free fetal DNA in maternal circulation. Approximately 10% of DNA in maternal circulation is of fetal origin.

• Analysis of cell-free fetal DNA is performed to identify pregnancies at high risk for chromosomal aneuploidy. Detection rates for trisomies 21, 18, and 13 are greater than 98%, with false positive rates of less than 0.5%.

• Some laboratories also test for sex chromosome aneuploidies (such as Turner syndrome or Klinefelter syndrome) as well as rare chromosome microdeletion syndromes, with variable performance.

• Each commercial laboratory offering NIPT has a proprietary platform and bioinformatics pipeline.

• Chromosome analysis on invasive diagnostic testing (CVS and amniocentesis) is also routinely available for assessment of fetal chromosome abnormalities in pregnancy.

**Guidelines and evidence**

• The American College of Medical Genetics and Genomics (ACMG, 2016) published a position statement regarding Non Invasive Prenatal Screening (NIPS), recommending the following:

  o “Informing all pregnant women that NIPS is the most sensitive screening option for traditionally screened aneuploidies (i.e., Patau, Edwards, and Down syndrome).”

  o “Informing all pregnant women of the availability of the expanded use of NIPS to screen for clinically relevant copy number variations (CNV’s) when the following conditions can also be met:”

    ▪ “Obstetric care providers should discuss with their patients the desire for prenatal screening as opposed to diagnostic testing (i.e., CVS or amniocentesis).”

    ▪ “Obstetric care providers should discuss with their patients the desire for maximum fetal genomic information through prenatal screening.”
“Obstetric care providers should inform their patients of the higher likelihood of false-positive and false-negative results for these conditions as compared to results obtained when NIPS is limited to common aneuploidy screening.”

“Obstetric care providers should inform their patients of the potential for results of conditions that, once confirmed, may have an uncertain prognosis.”

- “Offering diagnostic testing when a positive screening test result is reported after NIPS.”
- “Offering diagnostic testing for a no-call NIPS result due to low fetal fraction if maternal blood for NIPS was drawn at an appropriate gestational age. A repeat blood draw is NOT appropriate.”
- “Informing all pregnant women, as part of pretest counseling for NIPS, of the availability of the expanded use of screening for sex chromosome aneuploidies.”

- “Offering aneuploidy screening other than NIPS in cases of significant obesity.”

The ACMG specifically recommended against the following:

- “NIPS to screen for genome-wide CNVs. If this level of information is desired, then diagnostic testing (e.g., chorionic villous sampling or amniocentesis) followed by CMA is recommended.”
- “NIPS to screen for autosomal aneuploidies other than those involving chromosomes 13, 18, and 21.”

In 2016, The American College of Obstetricians and Gynecologists (ACOG) and the Society for Maternal Fetal Medicine (SMFM) published a joint practice bulletin stating the following:⁶

- “All women should be offered the option of aneuploidy screening or diagnostic testing for fetal genetic disorders, regardless of maternal age.”
- “Cell-free DNA screening tests for microdeletions have not been validated clinically and are not recommended at this time.”

In 2015, The American College of Obstetricians and Gynecologists (ACOG) and the Society for Maternal Fetal Medicine (SMFM) published a joint committee opinion stating the following:⁷

- “Given the performance of conventional screening methods, the limitations of cell-free DNA screening performance, and the limited data on cost-effectiveness in the low-risk obstetrics population, conventional screening methods remain the most appropriate choice for first-line screening for most women in the general obstetrics population.”
“Although any patient may choose cell-free DNA analysis as a screening strategy for common aneuploidies regardless of her risk status, the patient choosing this testing should understand the limitations and benefits of this screening paradigm in the context of alternative screening and diagnostic options.”

- The International Society for Prenatal Diagnosis (ISPD) first issued a position statement on NIPT in January 2011 and then updated its recommendations in April 2013 and again in April 2015. ISPD summarizes that:

  - “The following protocol options are currently considered appropriate:”
    - “cfDNA screening as a primary test offered to all pregnant women.”
    - “cfDNA secondary to a high risk assessment based on serum and ultrasound screening protocols (options 4-9 below).”

- The Society for Maternal Fetal Medicine (SMFM, 2015) published an expert opinion on NIPT, reiterating its position regarding the use of NIPT in high-risk pregnancies, while also clearly stating that “routine screening for microdeletions with cfDNA is not recommended.”

- The National Society of Genetic Counselors (NSGC, 2013; currently being updated 2016) practice guideline includes NIPT as an option for patients at increased risk for chromosome aneuploidy:

  - “Patients who desire screening information may be offered NIPT due to the high detection rates and low false positive rates. NIPT should only be offered in the context of informed consent, education, and counseling by a qualified provider, such as a genetic counselor. Standard confirmatory diagnostic testing should be offered as follow-up to positive NIPT results. High risk patients who decline NIPT but remain interested in screening should be made aware of alternate screening options as appropriate based on gestational age and screening availability.”

Criteria

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Prenatal Screening:
  - Cell-free fetal DNA-based prenatal screening for fetal aneuploidy (trisomy 13, 18, and 21) is considered medically necessary when all of the following criteria are met:
    - Singleton pregnancy, AND
    - Gestational age within the window validated by the selected testing laboratory, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

• Cell-free fetal DNA testing is not considered medically necessary in the following circumstances:
  o Singleton pregnancies in which the demise of a twin has occurred.
  o Multiple gestation pregnancies, which may be defined by the presence of one of the following ICD codes: O30.X, O31.X
  o More than one cell-free fetal DNA test performed per pregnancy defined as no more than one paid cell-free fetal DNA procedure code within 10 weeks
  o When karyotyping, aneuploidy FISH, and/or array CGH have already been performed on the pregnancy defined as any of these procedure codes paid within 10 weeks of the cell-free fetal DNA test

• Additional testing:
  o This policy applies to only cell-free fetal DNA-based prenatal screening for common aneuploidies of chromosomes 13, 18, and 21.
  o Screening for aneuploidy of the X and Y chromosomes and/or detection of less common trisomies, are not separately reimbursable under these coverage guidelines. Additional procedure codes billed with cell-free fetal DNA testing for this purpose are not eligible for reimbursement.
  o Detection of microdeletions by cell-free fetal DNA based screening is not eligible for reimbursement. Additional procedure codes billed with cell-free fetal DNA testing for this purpose are not eligible for reimbursement.

• Additional prenatal diagnostic testing:
  o Prenatal diagnosis by amniocentesis or CVS following NIPT is generally only indicated when NIPT results are abnormal or additional information becomes available throughout the pregnancy that suggests additional risk factors. Amniocentesis or CVS billed after NIPT is subject to medical necessity review.

Billing and reimbursement considerations

• Non-specific procedure codes (e.g. 81479, 81599, 84999) or any procedure codes that do not accurately describe the test methodology performed (e.g. 88271) are not eligible for reimbursement.

References


Oncotype DX Breast DCIS

Procedures addressed

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<tr>
<td>Oncotype DX Breast DCIS Assay</td>
<td>81479, 0045U</td>
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</table>

What is Oncotype DX for breast cancer prognosis

Definition

Oncotype DX® is a gene expression assay designed to determine the risk of a breast cancer recurrence within 10 years of the original diagnosis.¹

- It is intended for early stage, hormone receptor-positive, lymph node-negative breast cancer.¹-⁴
- Oncotype DX should be used with other standard methods of breast cancer assessment such as disease staging, grading, and other tumor markers.¹,²
- Oncotype DX results appear to correlate with chemotherapy benefit, which may help with the decision between tamoxifen only and adjuvant chemotherapy.⁵,⁶ Studies have demonstrated that the addition of Oncotype DX results changed treatment recommendations and decisions in 25% to 44% of patients, with the majority of recommendations changing from chemotherapy plus tamoxifen to tamoxifen only.⁷-⁹
- OncotypeDX can be used in individuals with ductal carcinoma in situ (DCIS) in addition to individuals with invasive carcinoma.

Test information

- Depending on the risk being calculated (local or distant metastasis), either a DCIS Breast Score® (DCIS or invasive carcinoma) or a Breast Recurrence Score® (invasive carcinoma) is calculated.¹⁰-¹³
- The Oncotype DX DCIS Breast Score® algorithm is intended for use in women with DCIS treated by local excision, with or without tamoxifen treatment. The score result is reported as a number between 0 and 100, with lower scores representing a low chance of recurrence and a higher score representing a high chance of recurrence within 10 years.¹⁰-¹³
Oncotype DX measures the expression level of 21 genes (16 cancer and 5 reference) from paraffin-embedded breast tumor tissue. These sixteen genes consistently correlated with distant recurrence-free survival in three studies that explored the expression of 250 genes in breast tumor samples.

The Oncotype DX DCIS score is calculated using a subset of 12 of the 21 gene Oncotype DX panel, including 7 cancer-related and 5 reference genes. On the patient report, average 10 year rates for any local/same breast recurrence (DCIS and invasive) as well as local invasive rate only are reported for a given DCIS Breast Score. Results of the DCIS Breast Score have the potential to change the treatment decision based on risk of local recurrence.

The results are provided as a Recurrence Score® (RS, 0-100) with higher scores reflecting higher risk of recurrence. Three risk categories help characterize prognosis:

- **Low risk (RS<18), ~50% of patients tested**
  - Least aggressive tumors
  - Metastasis unlikely
  - 7% recurrence by 10 yrs

- **Intermediate risk (RS 18-30), ~25% of patients tested**
  - More aggressive tumors
  - Metastasis more likely
  - 14% recurrence by 10 yrs

- **High risk (RS 31 or higher), ~25% of patients tested**
  - Most aggressive tumors
  - Metastasis most likely
  - 31% recurrence by 10 yrs

Patients with high scores benefit the most from chemotherapy, showing a substantial reduction in 10 year recurrence. Patients with intermediate scores show questionable benefit from chemotherapy, whereas those with low scores benefit the least from chemotherapy.

**Guidelines and evidence**

The National Comprehensive Cancer Network (NCCN, 2016) breast cancer treatment guidelines include Oncotype DX Breast ("21-gene RT-PCR assay") in their treatment algorithm for hormone receptor-positive, HER2-negative breast cancer.
They recommend considering the Oncotype DX assay in the following circumstances:

- Histology: Ductal, Lobular, Mixed, Metaplastic
- Tumor >0.5 cm (T1b-T3)
- pN0 or pN1mi (<2mm axillary node metastasis)

In the discussion, NCCN guidelines state: “Pending the results of prospective trials, the Panel considers the 21-gene RT-PCR assay [Oncotype DX] as an option when evaluating patients with primary tumors characterized as 0.6-1.0cm with unfavorable features or >1cm, and node-negative, hormone receptor positive and HER2-negative (category 2A). In this circumstance, the recurrence score may be determined to assist in estimating likelihood of recurrence and benefit from chemotherapy.”

The 14th St Gallen International Breast Cancer Conference (2015) Expert Panel confirmed previously published recommendations:

- Regarding Oncotype DX, the 2011 recommendations stated: “Several tests are available which define prognosis. These may indicate a prognosis so good that the doctor and patient decide that chemotherapy is not required. A strong majority of the Panel agreed that the 21-gene signature (Oncotype DX) may also be used where available to predict chemotherapy responsiveness in an endocrine responsive cohort where uncertainty remains after consideration of other tests…”

In 2015, the Panel “considered the role of multiparameter molecular marker assays for prognosis separately in years 1-5 and beyond 5 years and their value in selecting patients who require chemotherapy.” The Panel concluded that “only Oncotype DX commanded a majority in favor of its value in predicting the usefulness of chemotherapy.”

The Evaluation of Genomic Applications in Practice and Prevention Working Group (EGAPP, 2009 and updated in 2016) found:

- “Insufficient evidence to make a recommendation for or against the use of tumor gene expression profiles to improve outcomes in defined populations of women with breast cancer. In the updated 2016 publication, “evidence of clinical validity for Oncotype DX was confirmed as adequate. With regard to clinical utility, although there was evidence from prospective retrospective studies that the Oncotype DX test predicts benefit from chemotherapy, and there was adequate evidence that the use of Oncotype DX gene expression profiling in clinical practice changes treatment decisions regarding chemotherapy, no direct evidence was found that the use of Oncotype DX testing leads to improved clinical outcomes. Until definitive evidence for clinical utility is available,
clinicians must decide on a case-by-case basis whether to offer the test to patients."\textsuperscript{17,18}

- The 2007 evidence-based guidelines from the American Society of Clinical Oncology (ASCO) about breast cancer tumor marker use state:
  
  - "In newly diagnosed patients with node-negative, estrogen-receptor positive breast cancer, the Oncotype DX assay can be used to predict the risk of recurrence in patients treated with tamoxifen. Oncotype DX may be used to identify patients who are predicted to obtain the most therapeutic benefit from adjuvant tamoxifen and may not require adjuvant chemotherapy. In addition, patients with high recurrence scores appear to achieve relatively more benefit from adjuvant chemotherapy (specifically (C)MF) than from tamoxifen. There are insufficient data at present to comment on whether these conclusions generalize to hormonal therapies other than tamoxifen, or whether this assay applies to other chemotherapy regimens."\textsuperscript{3}
  
  - In 2016, the American Society of Clinical Oncology (ASCO), stated: "If a patient has ER/PgR-positive, HER2-negative (node-negative) breast cancer, the clinician may use the 21-gene recurrence score (RS; Oncotype DX; Genomic Health, Redwood City, CA) to guide decisions on adjuvant systemic chemotherapy. Type: evidence based. Evidence quality: high. Strength of recommendation: strong."\textsuperscript{4}

- Literature Review
  
  - Rakovitch et al. (2015) conducted a population cohort study (n=3320 women with DCIS) with a median follow-up period of 9.6 years.\textsuperscript{19} Study authors demonstrated that the DCIS Score independently predicted the risk of local recurrence in women with DCIS treated with breast conserving surgery (HR, 2.15; 95% CI, 1.43-3.22). Patients considered low risk via the DCIS Score (62%) had 10-year local recurrence of 13%; intermediate risk (17%) patients had 10-year local recurrence of 33%; and high risk (21%) patients had 10-year local recurrence of 28%. The DCIS Score is intended to provide a quantified risk score for local recurrence to help clinicians guide treatment decisions and potentially reduced the effects of overtreatment with radiotherapy.

  - Study results of this trial and others indicate that despite the ability of Oncotype DX to reclassify patients into different risk groups, it is not clear if the risk estimation is accurate enough to induce changes in treatment strategies or disease management, or if the 10-year local recurrence of approximately 13% is still low enough for patients to successfully avoid radiation therapy and the risk of its associated complications.\textsuperscript{20}

**Criteria**

- This test is considered investigational and/or experimental.
o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.

o In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References


Oncotype DX for Breast Cancer Prognosis

Procedures addressed

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What is Oncotype DX for breast cancer prognosis

Definition

Oncotype DX® is a gene expression assay designed to determine the risk of a breast cancer recurrence within 10 years of the original diagnosis.¹

- It is intended for early stage, hormone receptor-positive, lymph node-negative breast cancer.¹⁻⁴
- Oncotype DX should be used with other standard methods of breast cancer assessment such as disease staging, grading, and other tumor markers.¹⁻²
- Oncotype DX results appear to correlate with chemotherapy benefit,⁵⁻⁶ which may help with the decision between tamoxifen only and adjuvant chemotherapy. Studies have demonstrated that the addition of Oncotype DX results changed treatment recommendations and decisions in 25% to 44% of patients, with the majority of recommendations changing from chemotherapy plus tamoxifen to tamoxifen only.⁷⁻⁹

Test information

- Oncotype DX measures the expression level of 21 genes (16 cancer and 5 reference) from paraffin-embedded breast tumor tissue.¹ These sixteen genes consistently correlated with distant recurrence-free survival in three studies that explored the expression of 250 genes in breast tumor samples.⁵
- The results are provided as a Recurrence Score® (RS, 0-100) with higher scores reflecting higher risk of recurrence. Three risk categories help characterize prognosis:¹⁻²
  - Low risk (RS<18), ~50% of patients tested
    - Least aggressive tumors
• Metastasis unlikely
• 7% recurrence by 10 yrs
  o Intermediate risk (RS 18-30), ~25% of patients tested
    ▪ More aggressive tumors
    ▪ Metastasis more likely
    ▪ 14% recurrence by 10 yrs
  o High risk (RS 31 or higher), ~25% of patients tested
    ▪ Most aggressive tumors
    ▪ Metastasis most likely
    ▪ 31% recurrence by 10 yrs

Patients with high scores benefit the most from chemotherapy, showing a substantial reduction in 10 year recurrence. Patients with intermediate scores show questionable benefit from chemotherapy, whereas those with low scores benefit the least from chemotherapy.2,5,6

Guidelines and evidence

• The National Comprehensive Cancer Network (NCCN, 2016) breast cancer treatment guidelines include Oncotype DX Breast (“21-gene RT-PCR assay”) in their treatment algorithm for hormone receptor-positive, HER2-negative breast cancer.10
  o They recommend considering the Oncotype DX assay in the following circumstances:
    ▪ Histology: Ductal, Lobular, Mixed, Metaplastic
    ▪ Tumor >0.5 cm (T1b-T3)
    ▪ pN0 or pN1mi (<2mm axillary node metastasis)
  o In the discussion, NCCN guidelines state: “Pending the results of prospective trials, the Panel considers the 21-gene RT-PCR assay [Oncotype DX] as an option when evaluating patients with primary tumors characterized as 0.6-1.0cm with unfavorable features or >1cm, and node-negative, hormone receptor positive and HER2-negative (category 2A). In this circumstance, the recurrence score may be determined to assist in estimating likelihood of recurrence and benefit from chemotherapy.”10 (Category 2B: The recommendation is based on lower level evidence and there is non-uniform NCCN consensus, but no major disagreement).
• In V1.2015 of the NCCN guidelines, a footnote (bb in 2015 guidelines; cc in V2.2016) was added that states “The 21-gene RT-PCR assay recurrence score can be considered in select patients with 1-3 involved ipsilateral axillary lymph nodes to guide the addition of combination chemotherapy to standard hormone therapy. A retrospective analysis of a prospective randomized trial suggests that this test is predictive in this group similar to its performance in node-negative disease.”

• The 14th St Gallen International Breast Cancer Conference (2015) Expert Panel confirmed previously published recommendations:
  o Regarding Oncotype DX, the 2011 recommendations stated: “Several tests are available which define prognosis. These may indicate a prognosis so good that the doctor and patient decide that chemotherapy is not required. A strong majority of the Panel agreed that the 21-gene signature (Oncotype DX) may also be used where available to predict chemotherapy responsiveness in an endocrine responsive cohort where uncertainty remains after consideration of other tests...”
  o In 2015, the Panel “considered the role of multiparameter molecular marker assays for prognosis separately in years 1-5 and beyond 5 years and their value in selecting patients who require chemotherapy.” The Panel concluded that “only Oncotype DX commanded a majority in favor of its value in predicting the usefulness of chemotherapy.”

• The Evaluation of Genomic Applications in Practice and Prevention Working Group (EGAPP, 2009 and updated in 2016) found:
  o “Insufficient evidence to make a recommendation for or against the use of tumor gene expression profiles to improve outcomes in defined populations of women with breast cancer. In the updated 2016 publication, “evidence of clinical validity for Oncotype DX was confirmed as adequate. With regard to clinical utility, although there was evidence from prospective retrospective studies that the Oncotype DX test predicts benefit from chemotherapy, and there was adequate evidence that the use of Oncotype DX gene expression profiling in clinical practice changes treatment decisions regarding chemotherapy, no direct evidence was found that the use of Oncotype DX testing leads to improved clinical outcomes. Until definitive evidence for clinical utility is available, clinicians must decide on a case-by-case basis whether to offer the test to patients.”
  o The 2007 evidence-based guidelines from the American Society of Clinical Oncology (ASCO) about breast cancer tumor marker use state:
    o “In newly diagnosed patients with node-negative, estrogen-receptor positive breast cancer, the Oncotype DX assay can be used to predict the risk of recurrence in patients treated with tamoxifen. Oncotype DX may be used to identify patients who are predicted to obtain the most therapeutic benefit from adjuvant tamoxifen and may not require adjuvant chemotherapy. In addition,
patients with high recurrence scores appear to achieve relatively more benefit from adjuvant chemotherapy (specifically (C)MF) than from tamoxifen. There are insufficient data at present to comment on whether these conclusions generalize to hormonal therapies other than tamoxifen, or whether this assay applies to other chemotherapy regimens.\textsuperscript{3}

- In 2016, the American Society of Clinical Oncology (ASCO) stated “If a patient has ER/PgR-positive, HER2-negative (node-negative) breast cancer, the clinician may use the 21-gene recurrence score (RS; Oncotype DX; Genomic Health, Redwood City, CA) to guide decisions on adjuvant systemic chemotherapy. Type: evidence based. Evidence quality: high. Strength of recommendation: Strong.”\textsuperscript{4}

- **Additional clinical application issues:**
  - **Male gender** — No studies specific to the application of Oncotype DX in men with breast cancer have been identified. In general, the NCCN breast cancer treatment guidelines do not differentiate treatment on the basis of gender\textsuperscript{10}, which suggests Oncotype DX would not be excluded for males who meet NCCN clinical criteria for considering such testing.
  - **Multiple primary breast tumors** — No studies specific to the application of Oncotype DX in those with multiple breast primary cancers have been identified. Guidelines do not address this issue. A single poster summarized data in a study that used the Oncotype DX test to help assess if synchronous breast cancers were independent neoplastic events or spread of a single tumor. Of 11 patients who met criteria, 5 had different risk scores by Oncotype DX testing (with 3 of these patients having tumors assigned to different risk categories). Of these 5 with significantly different scores, 4 involved bilateral tumors and the other involved tumors in different quadrants. Comparing tumors by histology, 4 of 5 had clearly different histology and 1 had equivocal histology. Of the 6 with similar risk scores, 3 had the same histology, 2 equivocal, and in only 1 case was histology clearly different between the two tumors. This very limited data suggests Oncotype DX may be useful in multiple primaries when tumors independently meet criteria. A study published in 2016 noted that “Among women with synchronous bilateral ER-positive HER2-negative breast cancer, Oncotype DX recurrence scores were concordant in 67% of cases. These data suggest that testing of both tumors should be considered in patients who are candidates for adjuvant chemotherapy.”\textsuperscript{15}
  - **Positive lymph nodes** — There is currently insufficient evidence in the peer-reviewed literature regarding the use of Oncotype DX in women with early stage (ER+/HER2-) node-positive breast cancer who are considering adjuvant chemotherapy.
    - Several prospective and retrospective-prospective studies were identified evaluating the use of Oncotype DX in early stage, node-positive breast cancer, and results suggest that use of Oncotype DX allows for prognostic risk stratification.\textsuperscript{16-21} However, without chemotherapy, the risk of recurrence...
for patients with positive nodes appears to be notably higher than patients with negative nodes, and as such, it is not clear if patients with positive nodes can safely avoid chemotherapy treatment regimens based on Oncotype DX test results.

- There is at least one clinical trial underway, RxPonder, to evaluate the utility of the Oncotype DX Breast Cancer assay for women with 1-3 positive lymph nodes (ER/PR-positive, HER2-negative). This trial aims to support chance findings from a retrospective subset analysis of the SWOG-8814 trial data that suggested Oncotype DX high and low risk scores were able to predict chemotherapy benefit regardless of node status. An abstract presented at the European Breast Cancer Conference in 2016 presented the 5-year outcome data from a prospective trial with the conclusion of: WSG PlanB for the first time shows excellent 5-year disease free survival of 94% in a population of high risk node-negative and node-positive (pN1) (41.1% had node-positive disease) early BC patients (HR+ HER2−) who omitted adjuvant CT based on RS ≤11. Together recurrence score and classical clinical-pathological markers, despite of substantial heterogeneity in their assessment, provided independent prognostic information.

These 5-year outcome data from a prospective trial incorporating the RS support the incorporation of the assay in combination with nodal status, grade and tumor size for adjuvant treatment decisions in early HR+ HER2− breast cancer. However, evidence to support use in node-positive disease remains limited and use in this population is controversial. Therefore, while NCCN has addressed this indication in a footnote, use of Oncotype DX Breast Cancer assay in this population is not yet considered standard of care.

- **Ductal Carcinoma In Situ** — There is currently insufficient evidence in the peer-reviewed literature regarding the use of Oncotype DX in women with ductal carcinoma in situ (DCIS) who are considering radiation therapy.

  - Rakovitch et al. (2015) conducted a population cohort study (n=3320 women with DCIS) with a median follow-up period of 9.6 years. Study authors demonstrated that the DCIS Score independently predicted the risk of local recurrence in women with DCIS treated with breast conserving surgery (HR, 2.15; 95% CI, 1.43-3.22). Patients considered low risk via the DCIS Score (62%) had 10-year local recurrence of 13%; intermediate risk (17%) patients had 10-year local recurrence of 33%; and high risk (21%) patients had 10-year local recurrence of 28%. The DCIS Score is intended to provide a quantified risk score for local recurrence to help clinicians guide treatment decisions and potentially reduced the effects of overtreatment with radiotherapy. Study results of this trial and others indicate that despite the ability of Oncotype DX to reclassify patients into different risk groups, it is not clear if the risk estimation is accurate enough to induce changes in treatment strategies or disease management, or if the 10-year local recurrence of approximately 13% is still low enough for patients to successfully avoid radiation therapy and the risk of its associated complications.
Criteria

- Previous Testing:
  - No repeat Oncotype DX® testing on the same sample when a result was successfully obtained, and
  - No previous gene expression assay (e.g. Prosigna) performed on the same sample when a result was successfully obtained, AND

- Testing Multiple Samples:
  - When more than one breast cancer primary is diagnosed:
    - There should be reasonable evidence that the tumors are distinct (e.g., bilateral, different quadrants, different histopathologic features, etc.), and
    - There should be no evidence from either tumor that chemotherapy is indicated with or without knowledge of the Oncotype DX test result (e.g., histopathologic features or previous Oncotype DX result of one tumor suggest chemotherapy is indicated), and
    - If both tumors are to be tested, both tumors must independently meet the required clinical characteristics outlined below.

- Required Clinical Characteristics:
  - Invasive breast cancer meeting all of the following criteria:
    - Tumor size >0.5cm (5mm) in greatest dimension (T1b-T3), and
    - Estrogen receptor positive (ER+), and
    - HER2 negative, and
  - Patient has no regional lymph node metastasis or 1-3 positive ipsilateral axillary lymph nodes, and
  - Chemotherapy is a treatment option for the patient; results from this Oncotype DX test will be used in making chemotherapy treatment decisions, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


Oncotype DX for Colorectal Cancer Recurrence Risk

Procedures addressed

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<td>Oncotype DX Colon Cancer Assay</td>
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What is the Oncotype DX Colon Cancer Assay

Definition

The Oncotype DX® Colon Cancer Assay measures the expression of a panel of genes in stage II colon cancer tumors to predict the risk of future recurrence.¹

- Stage II colon cancer is defined by a primary tumor that has grown into or through the outermost layers of the colon, but has not spread to nearby lymph nodes or more distant metastasis.² At least 12 to 13 lymph nodes should be evaluated.³,⁴

- Stage II colon cancer is often treated with surgery alone with good prognosis.³,⁴ Adjuvant chemotherapy is not routinely recommended because it does not appear to improve 5-year survival rates by more than 5% among all people with stage II disease.³,⁴

- However, up to 25% of people with stage II disease will have a recurrence within 5 years.³ The decision about adjuvant chemotherapy is currently influenced by factors that help predict a higher recurrence risk, including:³,⁴
  - Inadequately sampled lymph nodes
  - Tumor characteristics such as T4 lesion (tumor penetrates to visceral peritoneum or adheres/invaded other organs²), perforation, poorly differentiated histology
  - Microsatellite instability and/or mismatch repair expression test results (particularly if considering 5-FU therapy only)

- These prognostic markers are imperfect and the need for additional validated prognostic markers is recognized.³
• The Oncotype DX Colon Cancer Assay proposes an additional method for stratifying recurrence risk to assist in the adjuvant chemotherapy decision. Genomic Health, who markets the assay, suggests the optimal use may be for people with “standard risk” stage II colon cancer (T3 tumor, mismatch repair proficient/microsatellite stable) following surgery, where other accepted prognostic factors do not make the chemotherapy decision clearer.¹

**Test information**

• The Oncotype DX Colon Cancer Assay quantifies the expression of 12 genes from paraffin-embedded primary colon cancer tissue samples.¹

  o Seven cancer genes associated with recurrence-free interval: Ki-67, C-MYC, MYBL2, FAP, BGN, INHBA, GADD45B
  o Five reference genes (to normalize expression levels): ATP5E, PGK1, GPX1, UBB, VDAC2

• The results are provided as a Recurrence Score, which translates into a percent recurrence risk at three years. Further risk information is provided based on such characteristics as T3/T4 tumor grade and mismatch repair results.¹

**Guidelines and evidence**

• The National Comprehensive Cancer Network (NCCN, 2017) colon cancer guidelines state “There are insufficient data to recommend the use of multi-gene assay panels to determine adjuvant therapy” when outlining the adjuvant therapy and surveillance options by cancer stage.⁴

• The Comparative Effectiveness Research in Genetics in Colorectal Cancer (CERGEN, 2010), an independent research team commissioned by the National Cancer Institute (NCI) [NCI RC 2-CA148471] to conduct comparative effectiveness research with funds secured from the American Recovery and Reinvestment Act of 2009, concluded:⁵,⁶

  o “To date, no prospective studies have been conducted to establish the clinical utility of the Oncotype DX colon cancer assay.”
  o “Although Genomic Health launched the Oncotype Dx colon cancer assay worldwide in January 2010, additional research is clearly needed before the value of this assay for clinical practice can be determined.”

• A prospective, multi-center study of the Oncotype Dx Colon Cancer Assay did demonstrate a change in physician decision-making behavior:⁸

  o Analyses on 141 stage IIA colon cancer patients enrolled from 17 centers showed that treatment recommendations changed for 63 patients (45%; 95% CI: 36%–53%), with treatment intensity decreasing for 47 (33%) and increasing for 16 (11%). Recommendations for chemotherapy decreased from 73 patients (52%) to 42 (30%), following review of recurrence score (RS) results. Increased
treatment intensity was more often observed at higher RS values, and decreased intensity was observed at lower values (p = .011).

- Improvement in patient outcomes based on this change in behavior were not included in the study.

- Analyses of test validation data from a subset of stage II colon cancer patients (n=1436) in the QUASAR study (randomized to chemotherapy versus observation) demonstrated:
  - The Recurrence Score (RS) was shown to be significantly associated with recurrence risk (p=0.004). T stage and mismatch repair status were the best pathology predictors of recurrence, but the RS predicted prognosis beyond those markers.
  - A “treatment score” was also calculated, but it did not predict chemotherapy benefit.

- Clinical validation in patients with stage II and stage III colon cancer (n=892) in the NSABP C-07 study.
  - The RS was shown to predict recurrence risk (p=.001). RS was not shown to predict stage (p=.90).
  - RS in combination with stage, may provide better insight into recurrence risk.

- The laboratory that developed this assay, Genomic Health, and collaborators originally published candidate gene study results that led to the selection of seven recurrence-risk genes and five reference genes for further validation.

- While not evidence, it is worthwhile noting that Palmetto GBA Medicare instituted coverage for this test stating:
  - “Palmetto GBA has completed the Oncotype DX Colon Cancer Assay assessment and determined that the test meets criteria for analytical and clinical validity and clinical utility as a reasonable and necessary Medicare benefit. Effective September 18, 2011, Palmetto GBA will reimburse Oncotype DX Colon Cancer Assay services for patients diagnosed with Stage II colon cancer.”

Criteria

- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical
management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.

- In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References


PALB2 Genetic Testing for Breast Cancer Risk

Procedure addressed

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What is PALB2 genetic testing

Definition

Breast cancer is the most frequently diagnosed malignancy and the leading cause of cancer mortality in women around the world. Hereditary breast cancer accounts for 5% to 10% of all breast cancer cases.

- Screening with breast magnetic resonance imaging (MRI) is recommended for women with a greater than 20% lifetime risk for disease based on estimates of risk models that are largely dependent on family history. A large body of evidence indicates that an increased lifetime risk of >20% can also be established through genetic testing. In particular, two cancer susceptibility genes, BRCA1 and BRCA2, are implicated in about 20% of all hereditary breast cancer cases. Other genes have also been identified in the literature as being associated with inherited breast cancer risk, including CHEK2, NBS1, ATM, BRIP1, and PALB2.¹²

- In particular, PALB2 is a gene that encodes a protein that may be involved in tumor suppression, and is considered a partner and localizer of BRCA2. Specifically, ~50 truncating mutations in PALB2 have been detected among breast cancer families worldwide. Kluska et al. (2017) estimates that a relative risk (RR) of 2.3 (95% CI, 1.4 to 3.9) is conferred by PALB2 mutations, indicating an approximate two-fold increased risk of developing hereditary breast cancer.³
The availability of multiple gene panel testing of variant genes implicated in hereditary breast cancer has led to increased interest in hereditary risk assessment in clinical practice. Clinical decisions based on risk assessment measures include screening with breast magnetic resonance imaging (MRI) and risk-reduction surgery, which have been shown to reduce the morbidity and mortality associated with breast cancer. However, results of peer-reviewed published clinical studies evaluating the clinical validity and clinical utility of multiple gene panels, particularly of unknown clinical significance, or of low-to-moderate penetrance, are still unclear. Broad application of such testing has yet to be fully adopted.

Genetic testing allows patients with an increased risk of cancer to receive appropriate medical management that may reduce risk for themselves and their family members. Early identification of at-risk women allows for increased clinical surveillance and may prompt more aggressive prevention strategies, such as prophylactic surgery or chemoprevention. The National Comprehensive Cancer Network (NCCN) guidelines have been expanded to incorporate genes known to be associated with an increased risk of breast cancer into medical management recommendations.

**Test information**

- **Full sequence analysis** of the PALB2 gene looks at all of the coding regions of the PALB2 gene.
- **Deletion/duplication analysis** looks for large rearrangements, duplications, and deletions in the PALB2 gene.
- **Known familial mutation testing** looks for a specific mutation in the PALB2 gene previously identified in a family member.

**Guidelines and evidence**

- The National Comprehensive Cancer Network (NCCN, 2017) includes breast cancer risk and management recommendations for individuals with PALB2 in a table located in their Genetic/Familial High-Risk Assessment: Breast and Ovarian guideline. However, it is noted that, “The inclusion of a gene on this table below does not imply endorsement either for or against multi-gene testing for moderate-penetrance genes.” Recommendations are as follows:
  - “Screening: Annual mammogram and consider breast MRI with contrast at 30y.”
  - “RRM: Consider based on family history.”

- The European Society for Medical Oncology (ESMO, 2016) states the following prevention and screening strategies for individuals with a PALB2 mutation:
  - “1) Clinical breast examination every 6-12 months staring from age 20-25”
  - “2) Annual breast MRI from age 20-29”
o “3) Annual breast MRI and/or mammogram at age 30-87.”

- ESMO (2016) also states the following regarding PALB2 testing, “The following genes might have moderate- to high-penetrance germline mutations for breast or ovarian cancer: p53, PTEN, CDH1, PALB2, CHEK2, ATM, RAD51C, STK11, RAD51D, BRIP1, MLH1, MSH, MSH6, and PMS2. Prevention and screening strategies for these mutations are summarized in Table 1 – due to limited research in individuals harboring these mutations, the level of evidence for these recommendations is mostly expert opinion, and a full discussion is beyond the scope of these guidelines.”

- The Second International Consensus Conference for Breast Cancer in Young Women (BCY2, 2016) led to publication of consensus recommendations. The following is stated regarding PALB2 genetic testing: 

  o “Other moderate-high-penetrance genes (e.g., CDH1, CHEK2, PALB2, RAD51C, BRIP1) should be tested for as deemed necessary by the geneticist.”

- A review of the available PALB2 literature revealed the following:

  o Direct evidence from a number of case control studies reporting relative risk and odds ratio values suggest that PALB2 testing accurately identifies PALB2 mutations, which are associated with an increased risk of developing breast cancer. Indirect evidence suggests that the clinical utility of PALB2 testing may alter clinical decision making enough to lead to improved patient health outcomes.

  o Direct and indirect evidence regarding clinical validity and clinical utility suggest that expanded panel testing may be used to identify more women who can benefit from appropriate breast cancer risk reduction strategies. However, gaps in knowledge persist regarding the precise cancer risk estimates for PALB2 (e.g., wide confidence intervals) and the predictive value in individuals and relatives who test negative for pathogenic variants yet have a strong family history of disease.

  o Before clinical utility of PALB2 testing can be adequately established, well-designed studies are crucial to directly evaluate the clinical utility of PALB2 to alter treatment and overall disease management strategies and improve morbidity and mortality outcomes in women who develop hereditary breast cancer.

Criteria

Known Familial Mutation Analysis

- Genetic Counseling:

  o Pre- and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Genetic Testing:
  o No previous full sequence testing or deletion/duplication analysis, and
  o Known family mutation in PALB2 identified in 1st, 2nd, or 3rd degree relative(s), AND
• Age 18 years or older, AND
• Rendering laboratory is a qualified provider of service per the Health Plan policy.

Full Sequence Analysis

• Member has not had previous PALB2 genetic testing, AND
• Member has met criteria for BRCA1/2 analysis,** AND
• Member has had BRCA1/2 analysis and no mutations were found

**Please see the guideline BRCA Analysis for criteria

References


Procedures addressed

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What are pancreatic cysts

Definition

Four of the most common types of pancreatic cysts are serous cystadenomas (SCA), solid-pseudopapillary neoplasm (SPN), mucinous cystic neoplasm (MCN), and intraductal papillary mucinous neoplasms (IPMN).¹

- Pancreatic cysts are reported as incidental findings in 3 to 13% of individuals undergoing imaging procedures. Given that pancreatic cancer is a rare, but lethal disease, proper assessment of pancreatic cysts is crucial for the definitive diagnosis and optimal treatment of individuals with malignant disease.
- Clinicians typically rely on imaging, cytology, and fluid chemistry to assess the malignancy risk of pancreatic cysts. Despite first-line assessments, individuals often undergo invasive surgery to treat suspicious pancreatic cysts.
- In cases where an individual’s diagnosis based on conventional pathologic and imaging approaches is inconclusive, PancraGEN has been proposed as an adjunctive risk stratification tool to provide additional clarifying information to inconclusive results of standard diagnostic tools, including imaging, carcinoembryonic antigen (CEA), cytology, and clinical risk factors.

Test information

- PancraGEN represents a form of topographic genotyping, a process that combines conventional imaging and pathologic analyses with molecular analyses.
- According to the test manufacturer, PancraGEN provides molecular results for DNA quantity and quality, oncogene point mutations (KRAS and GNAS), and tumor suppressor gene mutations to stratify patients according to their risk of progression to malignancy.²

1. Reference
2. Reference
• The test requires specimens of pancreatobiliary fluid, pancreatic masses, or pancreatic tissue usually obtained by endoscopic ultrasound (EUS) guided fine needle aspiration (FNA).\textsuperscript{2,3}

• The PancraGEN report categorizes patients into one of four groups: low risk category that supports surveillance (a. benign; b. statistically indolent) or high risk category that supports treatment intervention decisions (c. statistically higher risk; d. aggressive).

• This test is intended to determine a patient’s risk of cancer progression and assess the best course of treatment. Based on test results, low-risk patients with benign cysts may benefit from early disease surveillance and avoidance of invasive surgical resection, while higher risk patients with aggressive cysts can receive proper surgical treatment for malignant lesions.\textsuperscript{2}

Guidelines and evidence

• A small base of evidence comprised of a few clinical studies have evaluated the correlation between genetic testing using the PancraGen test and histology, cytology and pathology of surgical or biopsy specimens of pancreatic tissue.\textsuperscript{4-8} Two of the most relevant studies, both published by the manufacturer and evaluating the same patient population, reported results of a retrospective analysis of the National Pancreatic Cyst Registry study (n=492).
  
  o In the study by Al-Haddad et al. (2015), patients underwent testing with PathFinderTG (now PancraGEN) and were followed to evaluate disease progression to malignancy.\textsuperscript{4} Diagnostic performance of PathFinder TG testing were compared with a set of international consensus guidelines, published in 2012, used for disease management in clinical practice.\textsuperscript{5} After a median follow-up of 35 months, negative predictive values and sensitivity values for PathFinderTG and consensus guidelines were comparable, although positive predictive value and positive likelihood ratios were significantly improved for PathFinder TG. Study authors concluded that the PathFinder TG test may improve disease management by supporting a surveillance decision established by the Sendai guideline criteria.
In the same study population from the National Pancreatic Cyst Registry described in by Al-Haddad et al. (n=491), Loren et al. (2016) compared the association between diagnoses made with PancraGEN and those made with the consensus guidelines by Sendai and Fukouka (2012), and also reported on the subsequent clinical decisions made in the real world regarding choices made for either surveillance or surgical intervention. Study results suggest that testing with PancraGEN testing is significantly associated with real-world decisions, although it is not known if physician influence or patient preferences could have also impacted these decisions. Study results suggest that PancraGEN testing might properly reclassify some patients misclassified by consensus guidelines.

- Limitations of the evidence include retrospective study designs, limited follow-up times to adequately observe malignant progression, and a very small number of cases where results of PancraGEN and consensus guidelines do not agree.
- Given that the evidence base consists primarily of retrospective study designs, it is not clear if PancraGEN would perform well in a broad, general population of patients with pancreatic cysts. Small sample sizes may lead to imprecise estimates of test accuracy.

**Criteria**

- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  - In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

**References**


PCA3 Testing for Prostate Cancer

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<td>PCA3 Score</td>
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What is prostate cancer antigen 3 (PCA3)

Definition

Prostate cancer gene 3 (PCA3) is a non-protein-coding messenger RNA (mRNA) that is highly overexpressed in >95% prostate cancer tissue compared with normal prostate tissue or benign prostatic hyperplasia.\(^1\)

- The strong association between PCA3 mRNA levels and prostate cancer led to the development of a urinary assay to measure this analyte to aid in cancer detection.\(^1\)

Test information

- Following a digital rectal examination, the PCA3 gene can be quantified in urine specimens together with the prostate-specific antigen (PSA) to generate a PCA3 score.
- A high (>25) PCA3 Score indicates an increased likelihood of a positive biopsy. A low (<25) PCA3 Score is associated with a decreased likelihood of a positive biopsy.\(^2\)
- A multi-center study which included a total of 466 men found that at a score cutoff of 25 for men with at least one previous negative biopsy, PCA3 demonstrated 77.5% sensitivity, 57.1% specificity, and negative and positive predictive values of 90% and 33.6%, respectively. Men with a PCA3 score of <25 were 4.56 times more likely to have a negative repeat biopsy than men with a score of >25.\(^3\)

Guidelines and evidence

- Data from many peer-reviewed publications suggest that PCA3 gene testing, when used with other patient information, may help address some of the well-known challenges urologists face, such as identifying prostate cancers while reducing unnecessary repeat biopsies.\(^4-6\)
• The U.S Food and Drug Administration (2012) approved the Progensa PCA3 assay with the following intended use:⁷

  o “The PROGENSA® PCA3 Assay is indicated for use in conjunction with other patient information to aid in the decision for repeat biopsy in men 50 years of age or older who have had one or more previous negative prostate biopsies and for whom a repeat biopsy would be recommended by a urologist based on current standard of care, before consideration of PROGENSA PCA3 Assay results.”

  o “The Clinical Study only included men who were recommended by urologists for repeat biopsy. Therefore, the performance of the PROGENSA PCA3 Assay has not been established in men for whom a repeat biopsy was not already recommended.”

  o “Black Box Warning: The PROGENSA PCA3 Assay should not be used for men with atypical small acinar proliferation (ASAP) on their most recent biopsy. Men with ASAP on their most recent biopsy should be treated in accordance with current medical guidelines.”

• The National Comprehensive Cancer Network (NCCN, 2017) guidelines for prostate cancer early detection recognize the FDA-approved use of PCA3 testing and state:⁸

  o “It is well known that a negative prostate biopsy does not preclude a diagnosis of prostate cancer on subsequent biopsy. Those patients with negative prostate biopsies should be followed with DRE and PSA. Tests that improve specificity in the post-biopsy state—including 4Kscore, PHI, percent free PSA, PCA3, and ConfirmMDx—should be considered in patients thought to be higher risk despite a negative prostate biopsy.”

  o “Biomarkers that improve the specificity of detection are not, as yet, recommended as first-line screening tests. However, there may be some patients who meet PSA standards for consideration of prostate biopsy, but for whom the patient and/or the physician wish to further define the probability of high grade cancer. A percent free PSA <10%, PHI >35 or 4Kscore (which provides an estimate of the probability of high-grade prostate cancer) are potentially informative in patients who have never undergone a biopsy or after a negative biopsy; a PCA3 score >35 is potentially informative after a negative biopsy. The predictive value of the serum biomarkers discussed above has not been correlated with that of MRI. Therefore it is not known how such tests could be applied in optimal combination.”

• The American Urological Association (AUA 2013, confirmed 2015) guideline on the early detection of prostate cancer concluded:⁹

  o “At this point, the use of DRE, PSA derivatives (PSA density and age specific reference ranges) and PSA kinetics (velocity and doubling time), PSA molecular forms (percent free PSA and proPSA), novel urinary markers (PCA3), and
prostate imaging should be considered secondary tests (not primary screening tests) with potential utility for determining the need for a prostate biopsy, but with unproven benefit as primary screening tests.”

- “The Panel recognizes that these tests can be used as adjuncts for informing decisions about the need for a prostate biopsy—or repeat biopsy—after PSA screening, but emphasizes the lack of evidence that these tests will increase the ratio of benefit to harm.”

Criteria
Prostate cancer antigen testing (PCA3) may be indicated in males with ALL of the following:

- Age >50 years, and
- One or more previous negative prostate biopsies, and
- Continued clinical suspicion of prostate cancer based on digital rectal exam (DRE) or elevation of prostate specific antigen (PSA) of >3 ng/mL, and for whom a repeat biopsy would be recommended by a urologist based on current standard of care, and
- Atypical small acinar proliferation (ASAP) was NOT identified on the most recent biopsy.

References


Peutz-Jeghers Syndrome Testing

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What is Peutz-Jeghers syndrome

Definition

Peutz-Jeghers syndrome (PJS) is a genetic disorder characterized by the development of polyps (hamartomas) in the gastrointestinal (GI) tract, most commonly the small intestine. Polyps also occur in the stomach and colon and on occasion in the renal pelvis, urinary bladder, ureters, lungs, nares, and gallbladder. About a third of affected individuals present with polyps by age 10, and by age 20, about half have clinical signs and symptoms. Affected people also typically have mucocutaneous pigmented lesions—lip freckling is classic, but pigmentation may also develop in the mouth, gums, nose, perianal area, and on the fingers and toes. In addition to gastrointestinal polyps and cancer, people with PJS have an increased risk for other cancers, including those of the pancreas, lung, breast, uterus, cervix, ovaries, and testes. PJS is caused by mutations in the STK11 gene. STK11 is a tumor suppressor gene. Its normal role is to control growth and development of cells in the GI tract. Mutations in STK11 cause cells to grow and divide uncontrollably, leading to the development of polyps and an increased risk for cancer. PJS is inherited in an autosomal dominant pattern. Children of an affected person have a 1 in 2 (50%) chance to be affected. In large series, 60-78% of individuals with PJS had affected relatives and 17-40% of individuals represented isolated cases within their families. The proportion of a new (de novo) mutation is unclear due to variable expressivity and the frequency of subtle signs in parents is unknown.
Because of the potential early onset of polyp growth, surveillance is complex and involves monitoring at-risk individuals for related cancers, starting with baseline colonoscopy and upper GI endoscopy at age 8.\textsuperscript{1-4}

**Test information**

- Over 200 distinct STK11 gene mutations or deletions have been identified in people with PJS.

Molecular genetic testing is performed in parallel by two methods:\textsuperscript{1}

- **STK11 Sequence Analysis** is used to identify smaller mutations in STK11. Approximately 81\% of individuals with PJS will have a mutation detected by this method.

- **STK11 Deletion/Duplication Analysis** is used to identify larger deletions. Approximately 15\% of individuals with PJS will have a mutation detected by this method.

- Ninety-four to 96\% of individuals with PJS will have an STK11 pathogenic variant.\textsuperscript{5,6} The detection rate in familial versus sporadic cases is 87\% and 97.8\%, respectively.\textsuperscript{6}

- **STK11 Known Familial Mutation Analysis**: Once an STK11 mutation is identified in an affected person, predictive testing is available for at-risk family members, as is prenatal or preimplantation genetic diagnosis.\textsuperscript{1} Family members should be tested using the method that can accurately identify the familial mutation.

- A multi-gene panel can also be used to test individuals suspected of having PJS.

**Guidelines and evidence**

- Evidence-based guidelines for the diagnosis and management of PJS were published in 2010.\textsuperscript{2} These guidelines outline clinical diagnostic criteria for PJS and surveillance recommendations, but do not specifically address the utility of genetic testing.

- A clinical diagnosis of PJS may be made in an affected person when any ONE of the following is present (directly quoted):
  - Two or more histologically confirmed PJ polyps
  - Any number of PJ polyps detected in one individual who has a family history of PJS in close relative(s)
  - Characteristic mucocutaneous pigmentation in an individual who has a family history of PJS in close relative(s)
• Any number of PJ polyps in an individual who also has characteristic mucocutaneous pigmentation
  o “No clear genotype-phenotype correlation has been demonstrated in PJS, and no clear differences found between cases with STK11 mutation and in those in whom no mutation has been detected.”

• The National Comprehensive Cancer Network (2017) guidelines outline similar clinical diagnostic criteria and provide some guidance on surveillance, but do not address the use of genetic testing.
  o “A clinical diagnosis of PJS can be made when an individual has two or more of the following features:”
    - “Two or more Peutz-Jeghers-type hamartomatous polyps of the small intestine”
    - “Mucocutaneous hyperpigmentation of the mouth, lips, nose, eyes, genitalia, or fingers”
    - “Family history of PJS”
  o “The majority of cases occur due to mutations in the STK11 (LKB1) gene and clinical genetic testing is available.”
  o Screening procedures and intervals are outlined for breast, colon, stomach, pancreatic, small intestine, cervical, ovarian, uterine, and testicular cancers.

• Clinical diagnostic criteria have been validated by genetic testing in one series of 71 patients. Of 56 patients who met clinical criteria for PJS, 94% had an STK11 mutation found by a combination of sequencing and deletion/duplication analysis. Twelve patients had only a “presumptive diagnosis” of PJS based on the presence of hyperpigmentation or isolated PJS polyps, with no known family history. No STK11 mutations were found in those 12 patients.

• A 2016 expert-authored review states:1
  o “Testing of at-risk asymptomatic adults for Peutz-Jeghers syndrome is available after the disease-causing STK11 mutation has been identified in an affected family member.”
  o “Testing for the disease-causing mutation in the absence of definite symptoms of the disease is predictive testing. At-risk asymptomatic adult family members may seek molecular genetic testing in order to make personal decisions regarding medical surveillance, reproduction, financial matters, and career planning.”
“Because early detection of at-risk individuals who have an STK11 mutation affects medical management, particularly surveillance, testing of at-risk individuals during childhood is beneficial.”

The American Society of Clinical Oncologists (ASCO) position statement on genetic testing (originally published 1996; revised/affirmed in 2003, 2010, and 2015) outlines general recommendations for genetic testing for hereditary cancer syndromes and specifically addresses issues around genetic testing in at-risk children:

- Indications for Genetic Testing: ASCO recommends that genetic testing be offered when 1) the individual has personal or family history features suggestive of a genetic cancer susceptibility condition, 2) the test can be adequately interpreted, and 3) the results will aid in diagnosis or influence the medical or surgical management of the patient or family members at hereditary risk of cancer.

- Special Issues in Testing Children for Cancer Susceptibility: ASCO recommends that the decision to offer testing to potentially affected children should take into account the availability of evidence-based risk-reduction strategies and the probability of developing a malignancy during childhood. Where risk-reduction strategies are available or cancer predominantly develops in childhood, ASCO believes that the scope of parental authority encompasses the right to decide for or against testing.

- Tests for high-penetration mutations in appropriate populations have clinical utility, meaning that they inform clinical decision making and facilitate the prevention or amelioration of adverse health outcomes.

Criteria

STK11 (LKB1) gene testing may be considered for individuals with a suspected or known clinical diagnosis of Peutz-Jeghers syndrome, or a known family history of a STK11 (LKB1) mutation.

Testing is indicated for individuals whose medical and/or family history is consistent with ANY of these:

- A relative with a known deleterious STK11 (LKB1) gene mutation; OR
- A clinical diagnosis of PJS based on at least two of the following features:
  - At least two PJS-type hamartomatous polyps of the gastrointestinal tract
  - Mucocutaneous hyperpigmentation of the mouth, lips, nose, eyes, genitalia, or fingers
- A family history of PJS

**References**


Pharmacogenomic Testing Panels for Major Depressive Disorder

Procedures addressed

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What is major depressive disorder

Definition

Major depressive disorder (MDD) is a serious mental illness and one of the most common mental disorders in the United States, carrying the heaviest burden of disability among all mental and behavioral disorders. In 2015, roughly 16 million adults in the United States experienced at least one major depression episode in the previous year; this number represented 6.7% of all adults in the United States. A major depressive episode can include a number of symptoms, including depressed mood, insomnia or hypersomnia, change in appetite or weight, low energy, poor concentration, and recurrent thoughts of death or suicide, among other symptoms.

- Although mental health disorders are common in the United States, the burden of illness is concentrated among individuals with serious mental illness. In 2015, there were approximately 9.8 million adults in the United States with serious mental illness, representing 4% of all Americans. Serious mental illness can affect activities of daily living and may be accompanied by fatigue, insomnia, sudden weight loss, depressed mood, among other symptoms.

- Individuals with MDD experience high levels of recurrence; after recovery from one episode, the estimated risk of recurrence over a two year period is 40% and after two episodes, the risk is almost 75%.

- Treatment for MDD generally consists of a combination of psychotherapy (ie, cognitive behavioral therapy [CBT]) and pharmacotherapy (ie, antidepressants). The goal of treatment for MDD is primarily enabling remission of symptoms and restoring functioning.
• To find the optimal treatment approach, many clinicians try different antidepressants to maximize treatment response and reduce risk of remission. However, this “trial and error” approach is not always effective since the rates of remission are relatively low and vary considerably across individuals. Approximately 11 to 30% of patients recur after one year of antidepressant treatment.\(^5,6\) Consequences of treatment failure include the continuation of disabling symptoms that adversely affect work productivity, social functioning, and increase the risk of suicide.\(^5\)

• Pharmacogenomic testing has been developed to assist clinicians to predict those medications that could yield the most optimal treatment response and/or predict the lowest risk of side effects for an individual with mental health disorders, including MDD.

• Although this guideline will focus on the use of the GeneSight Psychotropic for management of major depressive disorder, it will apply broadly to pharmacogenomic testing for mental and behavioral health disorders. The focus of the guideline is guided by the preponderance of evidence (consisting of randomized or nonrandomized studies with control groups) in the peer-review literature available for the GeneSight test for the MDD disease indication.

Test information

• Researchers in the field of psychiatric pharmacogenomics have identified single nucleotide polymorphisms (SNPs) within genes that affect an individual's metabolism and response to anti-depressant medications.

• These SNPs have been combined into a medication decision support tool, GeneSight Psychotropic.\(^7\) Based on the composite phenotype measured for each patient, the GeneSight test has been proposed to assist clinicians in selecting psychotropic medication.\(^8\) Pharmacogenomic testing may be most useful in psychiatric patients who have treatment resistance, intolerable adverse effects, or the potential for experiencing adverse events or contraindications.\(^9\)

• GeneSight Psychotropic is a genetic panel that provides clinicians additional information about specific genetic variants to assist with decisions about drug selection. GeneSight tests for genetic variants in multiple pharmacokinetic and pharmacodynamic genes, which may impact drug tolerance and/or drug response. Specifically, the test analyzes 12 genes that may affect an individual's response to ~55 antidepressant and antipsychotic medications, four of which are pharmacokinetic genes from the cytochrome P450 family and 2 pharmacodynamic genes related to serotonin production.

• The GeneSight Psychotropic test is a proprietary combination of genotype, phenotype, and drug metabolism information for each subject with respect to CYP2D6, CYP2C19, CYP1A2, SLC6A4, and HTR2A. The GeneSight test represents majority of the SSRI and SNRI drug classes, as well as tricyclic antidepressants, and typical and atypical antipsychotics for the treatment of depression, anxiety, bipolar disorder, panic disorder, posttraumatic stress disorder
(PTSD), premenstrual dysphoric disorder, obsessive compulsive disorder (OCD), and schizophrenia.

- Results of the GeneSight Psychotropic are detailed in a report provided to the clinician, describing the most common medications for the patient’s diagnosed condition categorized by cautionary level.

- Additional pharmacogenomic panels or individual tests address treatment of mental health disorders, and are marketed by different labs or manufacturers. A few specific tests included in each panel are listed below:
  - Genecpt™ Assay (Genomind)
  - SureGene Test for Antipsychotic and Antidepressant Response (STA2R)
  - Proove Opioid Risk panel (Proove Biosciences)
  - Mental Health DNA Insight™ panel (Pathway Genomics)
  - IDgenetix-branded tests
  - Empowering Personalized Medicine (EPM) Panel
  - GeneSight ADHD, GeneSight Analgesic+
  - Informed PGx ADHD, Informed PGx Depression, Informed PGx Psychotropic
  - YouScript Personalized Prescribing System
  - ABCB1 Genotyping
  - CNSDose

Guidelines and evidence

- No specific evidence-based U.S. testing guidelines were identified.

- The published evidence is currently insufficient to support the use of pharmacogenomic testing to aid in the treatment of the psychiatric disorders, specifically MDD.10-15
• The manufacturer of GeneSight Psychotropic recently announced results from a randomized, double-blind, controlled trial (NCT02109939). The study is a 24 week RCT that examined 1,200 patients with moderate or severe MDD to assess the impact of the GeneSight test on psychiatric treatment response. Patients were randomized to the study group (those receiving the GeneSight test) or the usual treatment group. Assessments were completed at baseline and at 4, 8, 12, and 24 weeks, with patients and raters blinded at the 4 and 8 week weeks. Study results are slated to be presented at the American Psychiatric Association annual meeting in May, 2018. The study has not yet been published in the peer-reviewed scientific literature or indexed in PubMed.

Criteria
• These tests are considered investigational and/or experimental.
  o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  o In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

Billing and reimbursement considerations
• Due to these tests typically being performed as gene panels and reported out as associated risks using proprietary algorithms, individual CPT codes will also not be reimbursed under this guideline.
• If single gene testing is being requested and performed to determine an individual’s response to a specific medication (e.g. CYP2D6, CYP2C19, etc), please see either the pharmacogenomic testing clinical use guideline or a test-specific guideline to determine criteria for coverage.

References


Results-from-a-Large-Prospective-Trial-in-Patients-with-Major-Depressive-Disorder.html.
Polymerase Gamma (POLG) Related Disorders Genetic Testing

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

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What are POLG-related disorders

Definition

POLG-related disorders encompass a wide spectrum of disorders involving multiple organ systems, with variable severity and age of onset.¹²

- Onset of the POLG-related disorders ranges from infancy to late adulthood. Pediatric patients typically present with seizures and lactic acidosis, while myopathy, chronic progressive external ophthalmoplegia (CPEO), and sensory ataxia are the major presenting features later in life.³ Liver failure may also occur, particularly with exposure to the antiepileptic drug, valproic acid.¹
- POLG-related disorders are categorized into 6 recognizable phenotypes. Most affected individuals have some, but not all, of the features of a given phenotype.
  - Alpers-Huttenlocher syndrome (AHS)
    - AHS is typically characterized by refractory seizures, psychomotor regression, and hepatopathy. Other symptoms include migraine with visual auras, cortical blindness, hypotonia, ataxia, extrapyramidal movements, peripheral neuropathy, progressive spastic paraparesis, renal tubular acidosis, hearing loss, cyclic vomiting, and pancreatitis.⁴⁵ Development is often normal until disease onset, which is typically before 4 years of age. However, congenital static encephalopathy and later juvenile-onset has also been described.² AHS is inherited in an autosomal recessive pattern.⁶ When seizure etiology is unknown, valproic acid must be used with extreme caution, as it can precipitate liver dysfunction and/or failure in AHS.⁷⁸
  - Childhood myocerebrohepatopathy spectrum (MCHS)
Childhood myocerebrohepatopathy spectrum (MCHS) is a rapidly progressive disease with a fatal outcome that usually presents between the first few months of life and 3 years. It presents with failure to thrive, lactic acidosis, developmental delay, encephalopathy, dementia, myopathy, and hypotonia. Other features include liver failure, renal tubular acidosis, pancreatitis, cyclic vomiting, and hearing loss. MCHS has a similar presentation to AHS, however severe myopathy, specific liver pathology, and nonspecific brain MRI brain findings (diffuse atrophy) help differentiate MCHS from AHS. MCHS is inherited in an autosomal recessive pattern. In addition, seizures are less prominent and more easily controlled in MCHS compared to AHS.

- Myoclonic epilepsy myopathy sensory ataxia (MEMSA)
  - MEMSA is characterized by epilepsy, myopathy, and ataxia without ophthalmoplegia, and has also been known as spinocerebellar ataxia with epilepsy (SCAE). Disease onset typically occurs in adolescence and presents with cerebellar and sensory ataxia. Epilepsy usually follows, with refractory seizures leading to a progressive encephalopathy. MEMSA is inherited in an autosomal recessive pattern.

- Ataxia neuropathy spectrum (ANS)
  - The typical symptoms of ANS includes ataxia, neuropathy, and encephalopathy with seizures. The neuropathy may be sensory, motor, or mixed. Disease onset ranges between adolescence and adulthood. Migraine headaches may be the first presenting symptom and precede the other symptoms by many years. Psychiatric disturbance is a common feature. Other features include myoclonus, blindness, hearing loss, and a varying degree of liver failure. Clinical myopathy is very rare. The encephalopathy is often milder than AHS and more slowly progressive. ANS is inherited in an autosomal recessive pattern. ANS was previously referred to as mitochondrial recessive ataxia syndrome (MIRAS) and sensory ataxia neuropathy dysarthria and ophthalmoplegia (SANDO).

- Autosomal recessive progressive external ophthalmoplegia (arPEO)
  - arPEO is characterized by progressive weakness of the extraocular eye muscles resulting in ptosis and ophthalmoparesis without associated systemic involvement. Onset is typically in adulthood.

- Autosomal dominant progressive external ophthalmoplegia (adPEO)
  - adPEO is characterized by progressive weakness of the extraocular eye muscles resulting in ptosis and ophthalmoparesis. In addition, adPEO typically includes a generalized myopathy, sensorineural hearing loss, axonal neuropathy, ataxia, depression, Parkinsonism, hypogonadism, and cataracts. Previously, adPEO was called Chronic Progressive External Ophthalmoplegia plus (CPEO+).
• A 2014 expert-authored review states the following regarding management of POLG-related disorders:\(^1\)
  - “Clinical management is largely supportive and involves conventional approaches for associated complications including physiotherapy, speech therapy, nutritional interventions, respiratory support, and management of seizures and movement disorders.”
  - “Dose reductions of medications metabolized by hepatic enzymes to avoid toxicity.”
  - “Evaluations by a multidisciplinary team of healthcare providers based on clinical findings; monitoring of liver enzymes every two to four weeks after introduction of any new anticonvulsants.”
  - Valproic acid (Depakene\(^\text{®}\)) and sodium divalproate (divalproex) (Depakote\(^\text{®}\)) should be avoided because of the risk of precipitating and/or accelerating liver disease.

• Physical stressors such as infection, fever, dehydration, and anorexia can result in a sudden deterioration and should be avoided as much as possible.

• All 6 POLG-related disorders are caused by mutations in the POLG gene. However, inheritance varies.
  - AHS, MCHS, MEMSA, ANS, and arPEO are inherited in an autosomal recessive inheritance pattern. Males and female are equally likely to be affected. If both parents are carriers of one of these conditions, the risk for a pregnancy to be affected is 1 in 4 (25%).
  - adPEO is inherited in an autosomal dominant manner. When a parent has this condition, each of her/his offspring have a 50% risk of inheriting the mutation.

**Test information**

• Given that clinical diagnostic criteria do not exist, genetic testing of POLG is required in order to confirm the diagnosis of a POLG-related disorder.\(^1\)
  - For individuals with suspected adPEO, identification of one POLG mutation is required to confirm the diagnosis.
  - For individuals presenting with clinical features consistent with one of the five other phenotypes, identification of two (biallelic) mutations is required to confirm the diagnosis.

• **POLG Targeted Mutation Analysis** involves targeted testing of three common mutations in the POLG gene (p.Ala467Thr, p.Trp748Ser, and p.Gly848Ser).\(^1\) These three mutations account for about 70% of POLG mutations found in individuals with POLG related conditions.\(^1\)
• **POLG Full Gene Sequencing** can be performed to identify the remaining mutations in individuals with POLG-Related Disorders. Full sequencing is typically needed given that POLG-related disorders are mainly autosomal recessive conditions and the identification of two mutations in necessary for the diagnosis.

• **POLG Deletion/Duplication Analysis** can be performed if no mutations or only one mutation is found on targeted mutation analysis and/or full gene sequencing.

• **Multi-Gene Panels** - A number of large panels are available that sequence numerous nuclear-encoded mitochondrial genes for a broad approach to testing. Multi-gene panel tests, even for similar clinical scenarios, vary considerably laboratory by laboratory in the genes that are included and in technical specifications (e.g. depth of coverage, extent of intron/exon boundary analysis, methodology of large deletion/duplication analysis).

• While *biochemical analyses* of an affected tissue may be informative, they are not sensitive or specific enough to definitively diagnose a POLG-related disorder. Muscle biopsy can be completely normal in children and adults with a POLG-related disorder and in clinically unaffected tissue.13

**Guidelines and evidence**

• The Food and Drug Administration (FDA, 2017) states that Depakene (valproate) and Depakote ER (divalproex sodium) are contraindicated for patients known to have mitochondrial disorders caused by POLG mutations and children under two years of age who are clinically suspected of having a mitochondrial disorder.16

  o “Valproate-induced acute liver failure and liver-related deaths have been reported in patients with hereditary neurometabolic syndromes caused by mutations in the gene for mitochondrial DNA polymerase γ (POLG) (e.g., Alpers-Huttenlocher Syndrome) at a higher rate than those without these syndromes. Most of the reported cases of liver failure in patients with these syndromes have been identified in children and adolescents.”

  o “POLG-related disorders should be suspected in patients with a family history or suggestive symptoms of a POLG-related disorder, including but not limited to unexplained encephalopathy, refractory epilepsy (focal, myoclonic), status epilepticus at presentation, developmental delays, psychomotor regression, axonal sensorimotor neuropathy, myopathy, cerebellar ataxia, ophthalmoplegia, or complicated migraine with occipital aura. POLG mutation testing should be performed in accordance with current clinical practice for the diagnostic evaluation of such disorders. The A467T and W748S mutations are present in approximately 2/3 of patients with autosomal recessive POLG-related disorders.”

• Although not specific to genetic testing for POLG, the Mitochondrial Medicine Society (2015) 14 developed consensus recommendations for the diagnosis and management of mitochondrial disease. Testing strategies, including strategies for genetic testing, were discussed.
o Recommendations for DNA testing include the following:
  ▪ “When considering nuclear gene testing in patients with likely primary mitochondrial disease, NGS methodologies providing complete coverage of known mitochondrial disease gene is preferred. Single-gene testing should usually be avoided because mutations in different genes can produce the same phenotype. If no mutation is identified via known NGS panels, then whole exome sequencing should be considered.”

• The European Federation of Neurological Sciences/European Neurological Society (EFNS/ENS) 2014 consensus guidelines on the diagnosis and management of chronic ataxias in adulthood recommend POLG testing in the following evaluation of individuals with autosomal recessive cerebellar ataxia:15
  o “Step 1: mutation analysis of the FRDA gene for Friedreich’s ataxia (although one can refrain from this in the case of severe cerebellar atrophy), and biochemical testing that includes cholestanol, vitamin E, cholesterol, albumin, creatine kinase (CK) and a-fetoprotein. Also consider doing nerve conduction studies/EMG (presence versus absence of peripheral neuropathy, axonal versus demyelinating) and referral to an ophthalmologist (retinitis pigmentosa, cataract, cherry red spot etc.) (Table S2) (good practice point).”
  o “Step 2: mutation analysis of the SACS, POLG, Aprataxin (APTX) and SPG7 genes (taking into account specific phenotypes, as given in Table S2), and biochemical testing for white cell enzymes, phytanic acid and long chain fatty acids (good practice point).”
  o “Step 3: referral to a specialized centre, e.g. for skin or muscle biopsy targeted at diagnoses such as Niemann - Pick type C, recessive ataxia with coenzyme Q deficiency [aarF domain containing kinase 3 (ADCK3)/autosomal recessive spinocerebellar ataxia 9 (SCAR9)] and mitochondrial disorders, or for extended genetic screening using gene panel diagnostics (good practice point).”

• A 2014 expert-authored review suggests the following testing strategy for those with a known or suspected diagnosis of a POLG related disorder:1
  o “Standard clinical investigations can identify findings that, in the context of an appropriate family history, can suggest one of the POLG-related phenotypes.”
  o “Confirmation of the diagnosis of a POLG-related disorder requires identification of POLG pathogenic variants by molecular genetic testing.”
  o “One of the following two approaches can be used:”
    ▪ “Direct sequencing of POLG”
- “Two tiered analysis: targeted analysis for the common POLG pathogenic variants p.Ala467Thr, p.Trp748Ser, and p.Gly848Ser, followed by sequence analysis of the entire coding region if no pathogenic variants or only one pathogenic variant is found.”

  - “In persons meeting the diagnostic criteria of an autosomal recessive POLG-related disorder but in whom sequence analysis identifies only one disease-causing “POLG” allele, further testing may be considered to search for a second pathogenic variant in regulatory regions (e.g., the POLG promoter) or in related mitochondrial DNA replication genes such as C10orf2 (formerly PEO1; (encodes the twinkle helicase) and POLG2 to investigate the possibility of digenic inheritance.”

    - “Digenic inheritance has been reported in arPEO in a simplex case with pathogenic variants in POLG and C10orf2.”

    - “Oligonucleotide array should be strongly considered as microdeletions involving intragenic regions of POLG are reported and therefore relevant in a symptomatic individual with a single heterozygous pathogenic variant.”

  - “An alternative genetic testing strategy is use of a multi-gene panel that includes POLG and other genes of interest.”

**Criteria**

**Known POLG Family Mutation Testing**

- Genetic Counseling:
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Diagnostic Testing for Symptomatic Individuals
  - No previous genetic testing of POLG, and
  - If adPEO is suspected:
    - Clinical examination is consistent with a diagnosis of adPEO, and
    - POLG mutation identified in 1st degree biological relative, OR
  - If AHS, MCHS, MEMSA, ANS, or arPEO is suspected:
    - Clinical examination is consistent with a diagnosis of AHS, MCHS, MEMSA, ANS, or arPEO, and
    - Two POLG mutations identified in a sibling, or
■ One POLG mutation identified in both parents

POLG Full Gene Sequencing

• Genetic Counseling:
  ○ Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  ○ No previous genetic testing for POLG, and
  ○ No known POLG mutation in the family, AND

• Diagnostic Testing for Symptomatic Individuals:
  ○ If adPEO is suspected:
    ▪ Clinical examination is consistent with a diagnosis of adPEO, and
    ▪ Genetic testing is needed to confirm the diagnosis, OR
  ○ If AHS, MCHS, MEMSA, ANS, or arPEO is suspected:
    ▪ Clinical examination is consistent with a diagnosis of AHS, MCHS, MEMSA, ANS, or arPEO, and
    ▪ Genetic testing is needed to confirm the diagnosis, OR
  ○ If evaluating the risk for valproate-induced hepatic toxicity:
    ▪ The member has epilepsy, and
    ▪ There is suspicion for a POLG-related disorder based on the presence of at least one of the following:
      • unexplained encephalopathy, or
      • refractory epilepsy, or
      • status epilepticus at presentation, or
      • developmental delays, or
      • psychomotor regression, or
      • axonal sensorimotor neuropathy, or
      • myopathy and/or hypotonia, or
      • progressive spastic paraparesis, or
      • renal tubular acidosis, or
      • sensorineural hearing loss, or
• cyclic vomiting, or
• pancreatitis, or
• cerebellar ataxia, or
• ophthalmoplegia and/or ptosis, or
• complicated migraine with occipital aura, and

  ▪ The member is currently on Depakene (valproate) or Depakote ER (divalproex sodium) therapy, or the use of one of these medications is being proposed.

POLG Deletion/Duplication Analysis

• Genetic Counseling:
  o Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Criteria for POLG Full Gene Sequencing is met, AND
• If adPEO is suspected:
  o No mutations found on POLG Full Gene Sequencing, OR
• If AHS, MCHS, MEMSA, ANS, or arPEO is suspected:
  o No mutations or only one mutation found on POLG Full Gene Sequencing, OR
• If evaluating the risk for valproate-induced hepatic toxicity:
  o No mutations or only one mutation found on POLG Full Gene Sequencing

References


16. FDA label: Depakote ER. Available at: https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/021168s036lbl.pdf?page=45
Prader-Willi Syndrome Testing

Procedures addressed

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What is Prader-Willi syndrome

Definition

Features of Prader-Willi syndrome are caused when the Prader-Willi critical region (PWCR) on chromosome 15 is only inherited from the mother and there is no copy from the father. Prader-Willi syndrome can be caused by a chromosome deletion, uniparental disomy (two copies of the maternal chromosome), or imprinting defect. There are several genetic tests available that can help diagnose Prader-Willi syndrome.1,2

- Prader-Willi syndrome (PWS) is characterized by:1
  - Decreased muscle tone (hypotonia) and feeding difficulties in early infancy
  - Insatiable appetite in childhood that often results in obesity
  - Developmental delay
  - Short stature
  - Behavior problems
  - Small hands and feet
Underdeveloped genitalia and infertility

Test information

- **SNRPN Methylation Analysis**: This test is typically the first test in the evaluation of both Angelman syndrome and Prader-Willi syndrome. It will detect about 80% of patients with Angelman syndrome and >99% of patients with Prader-Willi syndrome. However, DNA methylation analysis does not identify the underlying cause, which is important for determining the risk to future siblings. This risk ranges from less than 1% to up to 50%, depending on the genetic mechanism. Follow-up testing for these causes may be appropriate.

- **FISH Analysis for 15q11-q13 Deletion**: If DNA methylation analysis for Angelman (AS) or Prader-Willi syndrome (PWS) is abnormal, deletion analysis is typically the next step. Approximately 70% of cases of both AS and PWS have a deletion in one copy of chromosome 15 involving the 15q11.2-q13 region. When looking specifically for this deletion, FISH (fluorescence in situ hybridization) analysis is most commonly performed. However, chromosome microarray can also detect such deletions (see that policy for guidance). If chromosomal microarray (CMA, array CGH) has already been done, FISH is not likely to be necessary.

- **Chromosome 15 Uniparental Disomy (UPD)**: If DNA methylation analysis is abnormal but deletion analysis is normal, UPD analysis next may be appropriate for evaluation of both Angelman (AS) and Prader-Willi syndrome (PWS). About 28% of PWS cases are due maternal UPD (both chromosome 15s are inherited from the mother). Both parents must be tested to diagnose UPD.

- **Imprinting Center Defect Analysis**: This test may be considered in the evaluation of Angelman syndrome (AS) and Prader-Willi syndrome (PWS) when methylation is abnormal, but FISH (or array CGH) and UPD studies are normal. Individuals with such results are presumed to have an imprinting defect. An abnormality in the imprinting process has been described in a minority of cases. However, imprinting center deletions may be familial, and if familial, the recurrence risk can be up to 50%.

- **Imprinting Center Known Familial Mutation Analysis**: If a mutation in the imprinting center has been identified in an affected family member, testing for just the known familial mutation in the imprinting center can be performed for at-risk relatives, including at-risk pregnancies.

Guidelines and evidence

- The American College of Medical Genetics and American Society of Human Genetics (2006) recommends two equally-accepted tiered approaches to testing for individuals exhibiting symptoms of Prader-Willi syndrome.²
  
  - **Approach one**:
    
    - Methylation analysis will detect >99% of individuals with PWS including those with deletion, uniparental disomy, or imprinting defect.
• If methylation testing is abnormal, it confirms the clinical diagnosis. However, to help determine whether there are risks of PWS in other family members it may be necessary to perform FISH, UPD and/or Imprinting Center testing to determine the exact cause of the abnormal methylation.

  ▪ FISH 15q11-q13 (deletion analysis)
    • If FISH testing is abnormal (70% of individuals with PWS will have a deletion) chromosome analysis may be considered to rule out a familial chromosome rearrangement (rare)
    • If FISH testing is normal, it is appropriate to consider UPD analysis.

  ▪ Uniparental Disomy (UPD) analysis of chromosome 15 determines if the patient inherited both copies of chromosome 15 from the mother.

  ▪ If methylation analysis is abnormal, but FISH and UPD analysis are normal, it is usually assumed there is an imprinting center mutation (which carries a higher recurrence risk than other causes). There is limited clinical testing available.\(^1\)

  o **Approach two:**

    ▪ FISH 15q11-q13 (deletion analysis) \(^2\)
      • If abnormal, a diagnosis of PWS is confirmed. Chromosome analysis may be considered to rule out a familial chromosome rearrangement (rare)
      • If normal then proceed to methylation analysis.

    ▪ Methylation analysis
      • If methylation analysis is abnormal, PWS diagnosis is confirmed, but UPD testing can occur to better understand recurrence risk

    ▪ Uniparental Disomy (UPD) analysis of chromosome 15
      • If methylation analysis is abnormal, but FISH and UPD analysis are normal, it is usually assumed there is an imprinting center mutation (which carries a higher recurrence risk than other causes). There is limited clinical testing available.\(^3\)

• Some of the same authors of the ACMG guidelines separately suggested the following:

  o “Clinical findings that should prompt diagnostic testing have been proposed based on analysis of diagnostic criteria met in individuals in whom the diagnosis of PWS has been molecularly confirmed. These differ by age group. The presence of all of the following findings at the age indicated is sufficient to justify DNA methylation analysis for PWS.”
• “A DNA methylation analysis consistent with PWS is sufficient for clinical diagnosis but not for genetic counseling, which requires identification of the underlying genetic mechanism.”¹

• “Prader-Willi syndrome (PWS) is a complex disorder whose diagnosis may be difficult to establish on clinical grounds and whose genetic basis is heterogeneous.”²

Criteria

SNRPN Methylation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous SNRPN methylation analysis, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay or intellectual disability, and
  o Some combination of the following:
    ▪ Neonatal hypotonia, or
    ▪ Feeding problems (i.e., poor suck) or poor growth in infancy, or
    ▪ Obesity and/or food-related behavior problems (i.e., hyperphagia; obsession with food), or
    ▪ Characteristic facial features, or
    ▪ Hypogonadism AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

FISH Analysis for 15q11-q13 Deletion

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous 15q11-q13 deletion analysis, and
  o No previous chromosomal microarray, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay or intellectual disability, and
o Some combination of the following:
  - Neonatal hypotonia, or
  - Feeding problems (i.e., poor suck) or poor growth in infancy, or
  - Obesity and/or food-related behavior problems (i.e., hyperphagia; obsession with food) or
  - Characteristic facial features, or
  - Hypogonadism, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Chromosome 15 Uniparental Disomy**

- Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Testing:
  o SNRPN methylation analysis results are abnormal, and
  o 15q11-q13 deletion analysis is negative, and
  o No previous chromosome 15 UPD studies, AND
- Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay or intellectual disability, and
  o Some combination of the following:
    - Neonatal hypotonia, or
    - Feeding problems (i.e., poor suck) or poor growth in infancy, or
    - Obesity and/or food-related behavior problems (i.e., hyperphagia; obsession with food), or
    - Characteristic facial features, or
    - Hypogonadism AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**Imprinting Center Defect Analysis**

- Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
• Previous Testing:
  o SNRPN methylation analysis results are abnormal, and
  o 15q11-q13 deletion analysis is negative, and
  o Previous chromosome 15 UPD studies negative, and
  o No previous imprinting center (IC) analysis, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Developmental delay or intellectual disability, and
  o Some combination of the following:
    ▪ Neonatal hypotonia, or
    ▪ Feeding problems (i.e., poor suck) or growth failure in infancy, or
    ▪ Obesity and/or food-related behavior problems (i.e., hyperphagia; obsession with food), or
    ▪ Characteristic facial features, or
    ▪ Hypogonadism AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy

Imprinting Center Known Familial Mutation Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous imprinting center defect analysis testing, AND

• Family History:
  o Familial imprinting center defect mutation known in blood relative, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

References


Prenatal Aneuploidy FISH Testing

Procedures addressed

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<table>
<thead>
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<th>Procedures addressed by this guideline</th>
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What is a chromosome abnormality

Definition

A chromosome abnormality is any difference in the structure, arrangement, or amount of genetic material packaged into the chromosomes.\(^1\) Aneuploidy refers to an abnormal number of chromosomes (i.e. extra or missing).\(^1\)

- Humans usually have 23 pairs of chromosomes. Each chromosome has a characteristic appearance that should be the same in each person.
- Chromosome abnormalities can lead to a variety of developmental and reproductive disorders. Common chromosome abnormalities that affect development include: Down syndrome (trisomy 21), trisomy 18, trisomy 13, Turner syndrome, and Klinefelter syndrome.
- About 1 in 200 newborns has some type of chromosome abnormality\(^2\) and a higher percentage of pregnancies are affected but lost during pregnancy. According to the American College of Obstetricians and Gynecologists (ACOG), "Fetuses affected with Down syndrome often do not survive pregnancy; between the first trimester and full term, an estimated 43% of pregnancies end in miscarriage or stillbirth."\(^3\)
• The risk of having a child with an extra chromosome, notably Down syndrome, increases as a woman gets older. Historically, invasive prenatal diagnosis was routinely offered to women 35 and over due to this risk. However, many babies with Down syndrome are born to women under 35. Prenatal screening for Down syndrome and certain other chromosome abnormalities is now routinely offered to all pregnant women. Prenatal diagnosis is also an option for any woman, although it is most commonly done for women with recognized risk factors.

Test information
• Fluorescence in situ hybridization (FISH) can be used to assess how many copies of a chromosome or smaller piece of DNA is in a cell.
  o FISH uses fluorescent probes that bind only to certain regions of a chromosome.
  o After binding, these fluorescent signals can be viewed by microscopy and counted in a sample of cells to determine if the appropriate number of copies is present.
  o Because chromosomes come in pairs, most normal cells will have two fluorescent signals for each probe.

• FISH analysis of prenatal samples (amniocentesis and CVS) is widely available for the chromosomes that are most commonly involved in prenatal chromosome abnormalities: 13, 18, 21, X, and Y.
  o FISH does not require dividing cells like conventional karyotyping. Therefore, results are generally available much more quickly (often within 2 days of the procedure) than for standard chromosome analysis (which usually takes at least 7 days).
  o While FISH results have been shown to be highly accurate, most experts recommend that no irreversible decisions be made unless the FISH results are either confirmed by karyotyping or the abnormal result fits with the remainder of the clinical findings (e.g., ultrasound anomalies are consistent with the particular chromosome abnormality).

Guidelines and evidence
• The American College of Obstetricians and Gynecologists (ACOG, 2016) issued prenatal diagnosis guidelines recommending the following:
  o “All pregnant women should be offered prenatal assessment for aneuploidy by screening or diagnostic testing regardless of maternal age or other risk factors.”
  o ACOG recommended the following in regards to FISH testing:
    • “When structural abnormalities are detected by prenatal ultrasound examination, chromosomal microarray will identify clinically significant chromosomal abnormalities in approximately 6% of the fetuses that have a
normal karyotype. For this reason, chromosomal microarray analysis should be recommended as the primary test (replacing conventional karyotype) for patients undergoing prenatal diagnosis for the indication of a fetal structural abnormality detected by ultrasound examination. If a structural abnormality is strongly suggestive of a particular aneuploidy in the fetus (e.g., duodenal atresia or an atrioventricular heart defect, which are characteristic of trisomy 21), karyotype with or without FISH may be offered before chromosomal microarray analysis."

- "An abnormal FISH result should not be considered diagnostic. Therefore, clinical decision making based on information from FISH should include at least one of the following additional results: confirmatory traditional metaphase chromosome analysis or chromosomal microarray, or consistent clinical information (such as abnormal ultrasonographic findings or a positive screening test result for Down syndrome or trisomy 18)."

- The American College of Medical Genetics (ACMG) and the American Society of Human Genetics (ASHG) issued a joint position statement on FISH in 2000. For prenatal FISH application, they state:  
  "For management of the fetus, it is reasonable to report positive FISH test results. Clinical decision-making should be based on information from two of three of the following: positive FISH results, confirmatory chromosome analysis, or consistent clinical information."

**Criteria**

Testing with aneuploidy FISH is allowed once per pregnancy AND only when a result is needed in less than one week in order to exercise some pregnancy management option AND at least one of the following indicate an increased risk for a chromosome abnormality:

- Screening result suggests aneuploidy
- Advanced maternal age
- One major or at least two minor fetal structural defects found on ultrasound
- Previous fetus or child with aneuploidy
- Parent of this pregnancy has a structural chromosome abnormality (e.g., translocation, inversion) involving chromosome 21, 13, 18, X, or Y
• Parent of this pregnancy has an extra chromosome (e.g., Down syndrome, XXX syndrome, Klinefelter syndrome)

References


### Prenatal Chromosome Analysis

#### Procedures addressed

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#### What is a chromosome abnormality

**Definition**

A chromosome abnormality is any difference in the structure, arrangement, or amount of genetic material packaged into the chromosomes.\(^1\)

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- Chromosome abnormalities can lead to a variety of developmental and reproductive disorders. Common chromosome abnormalities that affect development include Down syndrome (trisomy 21), trisomy 18, trisomy 13, Turner syndrome, and Klinefelter syndrome.
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• The risk of having a child with an extra chromosome, notably Down syndrome, increases as a woman gets older. Historically, invasive prenatal diagnosis was only offered to women over the age of 35. However, many babies with Down syndrome are born to women under 35. Prenatal screening for Down syndrome and certain other chromosome abnormalities is now routinely offered to all pregnant women. As a result, prenatal diagnosis is now an option for most pregnant women.

**Test information**

• Chromosome analysis — also called karyotyping — requires stimulating cells to divide, arresting cell division at metaphase when the chromosomes can be seen microscopically, and staining to visualize the banding patterns. Routine chromosome analysis allows visualization of about 400-550 bands per karyotype. High resolution chromosome analysis allows visualization of finer details and up to 1000 bands per karyotype.

• Chromosome analysis can be done on fetal cells from amniotic fluid (amniocentesis) or placenta (from CVS), as well as blood, tissue from a pregnancy loss, and other tissues when necessary. These tests are discussed separately.

• Once the chromosomes are prepared, chromosome analysis will identify any differences from normal that can be seen under the microscope. This includes entire missing or extra chromosomes, deletions or duplications within a chromosome that are large enough to be seen by microscope, and rearrangements including translocations and inversions.

• Chromosome analysis will not detect submicroscopic abnormalities, such as microdeletions. Specific probes or array CGH is required.

• Chromosome analysis also cannot detect any single gene disorders (such as cystic fibrosis, Tay-Sachs, etc.).

• Prenatal diagnosis usually takes about 2 weeks to complete.

**Guidelines and evidence**

• Prenatal diagnosis through amniocentesis and CVS is standard of care in obstetrics practice.

• Consensus guidelines from the American College of Obstetricians and Gynecologists (ACOG, 2016) recommend that:
  - “All pregnant women should be offered prenatal assessment for aneuploidy by screening or diagnostic testing regardless of maternal age or other risk factors.”
  - “Prenatal genetic testing cannot identify all abnormalities or problems in a fetus, and any testing should be focused on the individual patient’s risk, reproductive goals, and preferences.”
“Genetic testing should be discussed as early as possibly in pregnancy, ideally at the first obstetrics visit, so that first trimester options are available.”

- Practice Committee opinion from the Society for Assisted Reproductive Technology (SART) and American Society for Reproductive Medicine (ASRM, 2008) indicates that “Prenatal diagnostic testing to confirm the results of PGD is encouraged strongly because the methods used for PGD have technical limitations that include the possibility for a false negative result.”

- The Genetics Committee of the Society of Obstetricians and Gynaecologists of Canada (SOGC, 2006) indicate, “Couples considering IVF-ICSI for male-factor infertility should receive information, and if necessary formal genetic counseling, about the increased risk of de novo chromosomal abnormalities (mainly sex chromosomal anomalies) associated with their condition. Prenatal diagnosis by chorionic villus sampling (CVS) or amniocentesis should be offered to these couples if they conceive. (Evidence level II-2A).”

Criteria

Amniocentesis or CVS will be allowed once per pregnancy when at least one of the following criteria is met, indicating an increased risk of a chromosome abnormality in the pregnancy:

- Advancing maternal age; OR
- Abnormal first or second trimester nuchal translucency or maternal serum screening result; OR
- Abnormal cell-free fetal DNA screening result; OR
- Previous pregnancy with a chromosome abnormality; OR
- Parental chromosome abnormality; OR
- Abnormal fetal ultrasound; OR
- Family history of known or suspected chromosome problem; OR
- Pregnancy was conceived after preimplantation genetic diagnosis (PGD); OR
- Intracytoplasmic sperm injection (ICSI) due to male-factor infertility.

References


Prosigna Breast Cancer Prognostic Gene Signature Assay

MOL.TS.222.A
v2.0.2018

Procedures addressed

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<td>Oncology (Breast), mRNA analysis of 58 genes using hybrid capture, on formalin-fixed paraffin-embedded (FFPE) tissue, prognostic algorithm reported as a risk score</td>
<td>81520</td>
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What is Prosigna

Definition

Prosigna is a gene expression test designed to predict the chance of 10 year recurrence of breast cancer.

- Prosigna is indicated in post-menopausal women with hormone receptor positive, node negative (Stage I or II) or node positive (Stage II), early stage breast cancer.\(^1,2\)
- This assay is intended to assist patients and providers considering treatment with adjuvant chemotherapy.\(^1,2\)

Test information

- Prosigna is based on the 50 gene expression signature called PAM50. This assay uses RNA from formalin fixed paraffin embedded (FFPE) samples to calculate a risk score.\(^1,2\)
- The algorithm used for the Prosigna score uses the 50-gene expression profile in combination with clinical variables to classify breast cancer into one of the following four types: Luminal A, Luminal B, HER2-enriched, and Basal-like.\(^1,2\)
• A risk of recurrence (ROR) score is also calculated using gene expression and clinical variables (such as tumor size and degree of proliferation). This ROR score is reported as 0-100 and reflects the probability of disease recurrence at 10 years.\(^1,2\)
  
  o A ROR score of 1-10 corresponds to a 10 year distant recurrence of 0%. This risk increases to approximately 15% and then 33.3% when the ROR score reaches 61-70 and 91-100, respectively.\(^3\)

**Guidelines and evidence**

• Evidence-based clinical guidelines from the American Society of Clinical Oncology (ASCO) 2016 state:\(^4\)
  
  o “If a patient has ER/PgR-positive, HER2-negative (node-negative) breast cancer, the clinician may use the PAM50 risk of recurrence (ROR) score (Prosigna Breast Cancer Prognostic Gene Signature Assay; NanoString Technologies, Seattle, WA), in conjunction with other clinicopathologic variables, to guide decisions on adjuvant systemic therapy. Type: evidence based. Evidence quality: high. Strength of recommendation: strong.”
  
  o “If a patient has ER/PgR-positive, HER2-negative (node-positive) breast cancer, the clinician should not use the PAM50-ROR to guide decisions on adjuvant systemic therapy. Type: evidence based. Evidence quality: intermediate. Strength of recommendation: moderate.”
  
  o “If a patient has HER2-positive breast cancer, the clinician should not use the PAM50-ROR to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: insufficient. Strength of recommendation: strong.”
  
  o “If a patient has TN breast cancer, the clinician should not use the PAM50-ROR to guide decisions on adjuvant systemic therapy. Type: informal consensus. Evidence quality: insufficient. Strength of recommendation: strong.”

• The National Comprehensive Cancer Network (NCCN) 2016 Clinical Practice Guidelines for Breast Cancer state the following:\(^5\)
  
  o “The NCCN Panel members acknowledge that many assays have been clinically validated for prediction of prognosis. However, based on the currently available data, the panel believes that the 21-gene assay has been best validated for its use as a prognostic test as well as in predicting who is most likely to respond to systemic chemotherapy.”
  
  o “Patients with a high recurrence score obtained using the 21-gene assay clearly benefit from chemotherapy, whereas patients with a low score do not appear to benefit from the addition of chemotherapy regardless of the number of positive lymph nodes.”
“The 21-gene assay using reverse transcription polymerase chain reaction (RT-PCR) on RNA isolated from paraffin-embedded breast cancer tissue (Oncotype DX) is among the best-validated prognostic assays, and there are data showing that it can predict who is most likely to respond to systemic chemotherapy.”

- The European Society of Medical Oncology (ESMO) 2015 published new clinical practice guidelines and stated the following:
  
  "Gene expression profiles, such as MammaPrint (Agendia, Amsterdam, the Netherlands), Oncotype DX Recurrence Score (Genomic Health, Redwood City, CA), Prosigna (Nanostring Technologies, Seattle, WA) and EndoPredict (Myriad Genetics), may be used to gain additional prognostic and/or predictive information to complement pathology assessment and to predict the benefit of adjuvant chemotherapy. The three latter tests are designed for patients with ER-positive early breast cancer only.”

  "In cases of uncertainty regarding indications for adjuvant chemotherapy (after consideration of other tests), gene expression assays, such as MammaPrint, Oncotype DX, Prosigna and EndoPredict, may be used, where available.”

  "In cases when decisions might be challenging, such as luminal B HER2-negative and node-negative breast cancer, commercially available molecular signatures for ER-positive breast cancer, such Oncotype DX, EndoPredict, Prosigna, and for all types of breast cancer (pN0–1), such as MammaPrint and Genomic Grade Index, may be used in conjunction with all clinicopathological factors, to help in treatment decision making.”

- The St. Gallen International Expert Consensus (2015) stated the following:
  
  "The Panel considered the role of multiparameter molecular marker assays for prognosis separately in years 1-5 and beyond 5 years, and their value in selecting patients who require chemotherapy. Oncotype DX®, MammaPrint®, PAM-50 ROR® score, EndoPredict® and the Breast Cancer Index® were all considered usefully prognostic for years 1-5. Beyond 5 years, the Panel was divided almost equally on the prognostic value of Oncotype DX (despite the available data from NSABP Trial B-14 [32]); EndoPredict® (despite the report of Dubsky et al. [36]); and Breast Cancer Index (despite the report of Zhang et al. [37]). (All these reports show the respective tests to be prognostic beyond 5 years.) PAM50 ROR® score was agreed to be clearly prognostic beyond 5 years, and a clear majority rejected the prognostic value of MammaPrint® in this time period.”

- The Molecular Oncology Advisory Committee 2013 published a comparison of Oncotype DX with MammaPrint, PAM50, Adjuvant! Online, Ki-67, and IHC. Their recommendation is as follows:
  
  "In cases of breast carcinoma where Oncotype DX is indicated for clinical prognosis and treatment decisions, other assays should not currently be considered equivalent with respect to data generated or risk stratification.”
• The National Institute for Health and Care Excellence (NICE) 2013 stated the following:  
  o “MammaPrint, IHC4 and Mammostrat are only recommended for use in research in people with ER+, LN− and HER2− early breast cancer, to collect evidence about potentially important clinical outcomes and to determine the ability of the tests to predict the benefit of chemotherapy (see section 7 of the original guideline document). The tests are not recommended for general use in these people because of uncertainty about their overall clinical benefit and consequently their cost effectiveness.”
• The US Food and Drug Administration (FDA) cleared Prosigna for clinical use in 2013.

Criteria

• Previous Testing:
  o No repeat Prosigna testing on the same sample when a result was successfully obtained, and
  o No previous gene expression assay (e.g. OncotypeDx Breast) performed on the same sample when a result was successfully obtained, AND

• Testing Multiple Samples:
  o When more than one breast cancer primary is diagnosed:
    ▪ There should be reasonable evidence that the tumors are distinct (e.g., bilateral, different quadrants, different histopathologic features, etc.), and
    ▪ There should be no evidence from either tumor that chemotherapy is indicated with or without knowledge of the Prosigna test result (e.g., histopathologic features or previous Gene Expression Assay result of one tumor suggest chemotherapy is indicated), and
    ▪ If both tumors are to be tested, both tumors must independently meet the required clinical characteristics outlined below, AND

• Required Clinical Characteristics:
  o Invasive breast cancer meeting all of the following criteria:
    ▪ Tumor size ≥0.4cm (4mm) in greatest dimension (T1b-T3), and
    ▪ Hormone receptor positive (ER+/PR+), and
    ▪ HER2 negative, and
  o Patient has no regional lymph node metastasis, and
Chemotherapy is a treatment option for the patient; results from this Prosigna test will be used in making chemotherapy treatment decisions, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

References
2. Prosigna website. Available at: http://prosigna.com/
3. Prosigna Packet Insert US. Available at: http://prosigna.com/docs/Prosigna_Packet_Insert_US.pdf
8. Chang M, Ismaila N, Kamel-Reid S, Rutherford M, Hart J, Bedard P, Trudeau M, Eisen A, Molecular Oncology Advisory Committee. Comparison of Oncotype DX with multi-gene profiling assays (e.g., MammaPrint, PAM50) and other tests (e.g., Adjuvant! Online, Ki-67 and IHC4) in early-stage breast cancer. Toronto (ON): Cancer Care Ontario (CCO); 2013 Nov 20. 39 p. Available at: https://www.guideline.gov/summaries/summary/47790
PTEN Hamartoma Tumor Syndromes Testing

MOL.TS.223.A
v2.0.2018

Procedures addressed

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<td>PTEN Deletion/Duplication Analysis</td>
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What is PTEN hamartoma tumor syndrome

Definition

PTEN hamartoma tumor syndrome (PHTS) is used to describe the group of conditions caused by PTEN mutations that include hamartomatous growths: Cowden syndrome, Bannayan-Riley-Ruvalcaba syndrome, Proteus syndrome and Proteus-like syndrome, and autism spectrum disorder with macrocephaly.

- Historically, these conditions have been considered clinically distinct but share an underlying genetic etiology, and show some overlap in families.¹
  - **Cowden syndrome** (CS) is characterized by an increased risk for benign and malignant tumors of the breast, endometrium, and thyroid (non-medullary).¹,² Other common features include macrocephaly and growths on the skin or mucous membranes (mucocutaneous lesions). Prevalence is estimated to be 1 in 200,000 individuals, although CS is believed to be underdiagnosed.¹ Up to 85% of people with a clinical diagnosis of CS have a PTEN mutation.¹
  - **Lhermitte-Duclos disease** (LDD) is a rare, benign tumor of the cerebellum called dysplastic gangliocytoma that may present in childhood or adulthood.¹,² Most adult-onset LDD is caused by a PTEN mutation even when no other signs of CS are present.¹
  - **Bannayan-Riley-Ruvalcaba syndrome** (BRRS) is a genetic disorder characterized by macrocephaly, multiple benign intestinal polyps (hamartomatous type), lipomas, colored spots on the tip of the penis (pigmented macules of the glans penis), and hemangiomas. Some people with BRRS have intellectual disability and/or birth defects. There may be an increased risk for
several types of cancer, including breast, thyroid and endometrial.\textsuperscript{2} Up to 65\% of people with a clinical diagnosis of BRRS have a PTEN mutation.\textsuperscript{1}

- **Proteus and Proteus-like syndromes** are highly variable conditions characterized by overgrowth of several different tissues usually in a patchy asymmetric pattern (mosaic) that is often present from birth but gets worse over time.\textsuperscript{1}

  Clinical signs and symptoms include connective tissue and epidermal nevi (hamartomatous growths), ovarian cystadenomas, parotid monomorphic adenomas, lipomas, capillary/venous/lymphatic malformations, and a characteristic facial dysmorphology. Up to 50\% of people with Proteus-like syndrome and 20\% of people with Proteus syndrome have a PTEN mutation.\textsuperscript{1}

- **Autism spectrum disorder with macrocephaly**, defined as >2.5 SDs above the age mean or ≥97\textsuperscript{th} percentile, may be caused by a mutation in the PTEN gene.\textsuperscript{1} An estimated 3-20\% of all people with ASD/macrocephaly have a PTEN mutation.\textsuperscript{1,3} The likelihood may be greater if other family members have signs and symptoms in the PHTS spectrum.

- An online tool is available to estimate the likelihood of identifying a PTEN mutation based on clinical findings: http://www.lerner.ccf.org/gmi/ccscore/.

- People with CS need heightened cancer surveillance starting at age 18 (or earlier if warranted: “For those with a family history of a particular cancer type at an early age screening may be initiated five to ten years prior to the youngest diagnosis in the family”\textsuperscript{1,2}). The exception is children should have a yearly thyroid ultrasound from the time of diagnosis and skin check with physical examination.\textsuperscript{1} Because of the overlap in clinical phenotypes, people with other PTEN-related conditions are advised to follow the same heightened cancer surveillance guidelines as for CS.\textsuperscript{4,5}

  - The lifetime risk for breast cancer is 25-50\% with an average age at diagnosis of 38-46 years.\textsuperscript{1} However, a 2012 publication by Tan et al. reports that this lifetime risk may be as high as 85\%, particularly in individuals with PTEN promoter mutations.\textsuperscript{6} The lifetime risk for thyroid cancer can range from 10\% to as high as 35\%.\textsuperscript{1,6} Benign thyroid growths are also found in up to 75\% of people with CS.\textsuperscript{1} “However, the high frequency of thyroid disease in the general population means that when taken on their own, thyroid neoplasms have a low predictive value for identifying mutations carriers.”\textsuperscript{7}

  - Endometrial cancer has an estimated 5-10\% lifetime risk, although this is not well-defined.\textsuperscript{1} Tan et al. reports a lifetime risk of up to 28\%.\textsuperscript{6}

  - The gastrointestinal polyp risk (often colonic) in patients with CS may be 80\% or higher and the lifetime risk for colorectal cancer is estimated to be 9\%.\textsuperscript{6}

  - Early onset colorectal cancer has been reported in 13\% of patients with PTEN associated CS indicating earlier and more frequent colonoscopy is warranted in this population.\textsuperscript{6,8,9}
Additionally, an increased lifetime risk for kidney cancer (approximately 34%) and melanoma (about 5-6%) has been reported.\textsuperscript{1,2,6}

- PTEN mutations are inherited in an autosomal dominant manner, meaning that a person only needs a mutation in one copy of the gene to be affected. A child of an affected person has a 50% chance to inherit the mutation. Nearly all people with a PTEN mutation will develop symptoms (complete penetrance).\textsuperscript{1,2}

**Test information**

- **PTEN Sequencing:** Evaluates each DNA nucleotide to identify mutations throughout the gene. Such testing will detect a mutation in about 80% of people with a clinical diagnosis of CS and 60% of people with a clinical diagnosis of BRRS.\textsuperscript{1}

  - **Sequencing of the promoter region** will detect an additional 10% of PTEN mutations that cause CS.\textsuperscript{1} NCCN recommends comprehensive testing, which should include full sequencing, gene deletion/duplication analysis, and promoter analysis of the PTEN gene.\textsuperscript{2} As such, it is important to determine whether or not the selected laboratory includes PTEN promoter analysis in their testing.\textsuperscript{2}

- **PTEN Deletion/Duplication Analysis:** Used in cases where a mutation is not found by sequencing. The likelihood of identifying a deletion or duplication in people with clinically diagnosed CS is unknown, but expected to be relatively low.\textsuperscript{1} About 11% of people with BRRS have large PTEN gene deletions.\textsuperscript{1}

- **PTEN Known Familial Mutation Analysis:** Once the familial mutation is identified, testing for that one mutation can be offered to at-risk relatives. Such testing is much less expensive than complete gene testing and the results are highly reliable.

**Guidelines and evidence**

- Evidence-based guidelines (Category 2A) from the National Comprehensive Cancer Network (NCCN, 2017) support the use of PTEN genetic testing in those with clinical features or a family history. They recommend PTEN genetic testing in any of the following situations:\textsuperscript{2}

  - Family history of a known PTEN mutation [PTEN known familial mutation testing is appropriate]
  - A personal history of any of the following:
    - Bannayan-Riley-Ruvalcaba syndrome (BRRS)
    - Adult-onset Lhermitte Duclos disease (cerebellar dysplastic gangliocytoma)
    - Autism spectrum disorder and macrocephaly (greater than or equal to 97th percentile)
    - Two or more biopsy proven trichilemmomas
    - Macrocephaly and at least one other major** criteria
Three major** criteria without macrocephaly
- One major** and three or more minor*** criteria
- Four or more minor*** criteria

- At-risk relative of someone clinically diagnosed with Cowden syndrome or BRRS (who has not had genetic testing), when the at-risk relative has at least one major** or two minor*** criteria. Ideally, the at-risk person is a first-degree relative (parent, sibling, child) of someone clinically diagnosed, but testing more distant relatives is acceptable if closer relatives are not available or willing to have testing.

**Major:
- Breast cancer
- Endometrial cancer
- Follicular thyroid cancer
- Multiple GI hamartomas or ganglioneuromas
- Macrocephaly (at least 97th percentile: 58cm in adult women and 60cm in adult men)
- Macular pigmentation of glans penis
- Mucocutaneous lesions: one biopsy-proven trichilemmoma, multiple palmoplantar keratoses, multifocal or extensive oral mucosal papillomatosis, multiple cutaneous facial papules (often verrucous)

***Minor:
- Autism spectrum disorder
- Colon cancer
- 3 or more esophageal glycogenic acanthoses
- Lipomas
- Intellectual disability (IQ less than or equal to 75)
- Papillary or follicular variant of papillary thyroid cancer
- Thyroid structural lesions (e.g., adenoma, nodule(s), goiter)
- Renal cell carcinoma
- Single GI hamartoma or ganglioneuroma
- Testicular lipomatosis
- Vascular anomalies (including multiple intracranial developmental venous anomalies)

Note
Note that these NCCN defined major and minor criteria for genetic testing do not fully align with the major and minor criteria required for a clinical diagnosis.

- The American College of Medical Genetics and Genomics (ACMG, 2008) issued consensus practice guidelines on the genetics evaluation of autism. They propose an evaluation scheme with three tiers. The first tier includes routine studies such as chromosome analysis and fragile X genetic testing. PTEN gene testing is recommended as a second tier test when the head circumference is greater than 2.5 SDs above the mean (if no diagnosis is made via first tier testing).10
• An expert-authored review (2014) of the PTEN hamartoma syndromes states:\(^1\)
  
  o “The diagnosis of PHTS is made only when a PTEN mutation is identified.”
  o “The appropriate order of PTEN testing to optimize yield:”
    
    i. “Sequence all PTEN coding exons 1-9 and flanking intronic regions. If no pathogenic variant is identified, perform:”
    
    ii. “Deletion/duplication analysis. If no pathogenic variant is identified, consider:”
    
    iii. “Sequence analysis of the promoter region for variants that decrease gene expression”
  
  o “The most serious consequences of PHTS relate to the increased risk of cancers including breast, thyroid, endometrial, and to a lesser extent, renal. In this regard, the most important aspect of management of any individual with a PTEN pathogenic variant is increased cancer surveillance to detect any tumors at the earliest, most treatable stages.”

Criteria

PTEN gene testing may be considered in individuals with a suspected or known clinical diagnosis of Cowden syndrome, Bannayan-Riley-Ruvalcaba syndrome (BRRS), or another PTEN-related hamartoma syndrome; or who have a known family history of a PTEN mutation.

PTEN Known Familial Mutation Analysis

• Genetic Counseling:
  
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  
  o No previous genetic testing of PTEN, AND

• Diagnostic and Predisposition Testing:
  
  o Known deleterious family mutation in PTEN identified in 1\(^{st}\), 2\(^{nd}\), or 3\(^{rd}\) degree biologic relative(s), AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

PTEN Sequencing with promoter analysis

• Genetic Counseling:
PTEN Hamartoma Tumor Syndromes

Criteria for testing purposes are:\(^2\)
**Major:**

- Breast cancer
- Endometrial cancer
- Follicular thyroid cancer
- Multiple GI hamartomas or ganglioneuromas
- Macrocephaly (at least 97\textsuperscript{th} percentile: 58cm in adult women and 60cm in adult men)
- Macular pigmentation of glans penis
- Mucocutaneous lesions: one biopsy-proven trichilemmoma, multiple palmoplantar keratoses, multifocal or extensive oral mucosal papillomatosis, multiple cutaneous facial papules (often verrucous)

***Minor:***

- Autism spectrum disorder
- Colon cancer
- \( \geq 3 \) esophageal glycogenic acanthoses
- Lipomas
- Intellectual disability (IQ≤75)
- Papillary or follicular variant of papillary thyroid cancer
- Thyroid structural lesions (e.g., adenoma, nodule(s), goiter)
- Renal cell carcinoma
- Single GI hamartoma or ganglioneuroma
- Testicular lipomatosis
- Vascular anomalies (including multiple intracranial developmental venous anomalies)

### References


Rett Syndrome Testing

Introduction

Rett syndrome testing is addressed by this guideline.

Procedures addressed

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What is Rett syndrome

Definition

Rett syndrome, or classic Rett syndrome, is an X-linked brain development disorder that typically affects females. Atypical Rett syndrome may be more mild or severe than classic Rett syndrome.

Prevalence

Rett syndrome affects about 1 in 10,000 females. Males are rarely affected with less than 100 overall affected patients reported.¹

Symptoms

Girls with Rett syndrome may not show signs at birth or during infancy, but by the age of 6 to 18 months they begin to lose their motor and language skills, which eventually stabilizes.¹

Signs and symptoms of Rett syndrome usually include ¹ ²

- intellectual disability or developmental delay
- specific hand movements, like hand “wringing” and clapping for no reason
- loss of speech
- problems with sleep
• seizures
• growth failure
• autistic behaviors, and
• gait abnormalities, either impaired or complete absence of ability.

Cause

Rett syndrome is caused by genetic changes (mutations) in the MECP2 gene, located on the X chromosome. Females have two X chromosomes and males have one X chromosome and one Y chromosome.¹

Inheritance

Rett syndrome is an X-linked condition. A female who is found to be a MECP2 mutation carrier has a 50% chance to pass the mutation to her children.

Approximately 99% of cases of Rett syndrome are the result of a new genetic change (de novo mutation) in the affected person and are not inherited from a carrier parent.¹⁻³ Cases of minimally affected or unaffected female carriers of MECP2 mutations have been reported.¹⁻⁴

Cases of MECP2 mutations in only the germline (egg or sperm) of parents of affected people have been reported.¹⁻³ In one study, prenatal diagnosis was offered to nine couples who had a previous child with Rett syndrome due to a known de novo MECP2 mutation.³ One of the nine pregnancies was found to have the same MECP2 mutation as in the affected sibling.² Since germline mosaicism cannot be predicted or ruled out in families who have a child with Rett syndrome, prenatal diagnosis may be offered.

If a mutation of unclear significance is found in an affected person, testing both the mother and the father may be appropriate to help to determine whether the mutation is actually causing the disease.¹

Diagnosis

Classic Rett syndrome is generally diagnosed by established clinical diagnostic criteria.¹⁻² Diagnostic criteria have also been suggested for atypical Rett syndrome, but diagnostic criteria are imperfect for reliably diagnosing Rett syndrome.¹⁻²

Genetic testing may be useful to confirm a diagnosis (particularly when unclear based on clinical criteria) and to identify the mutation for genetic counseling purposes.

MECP2 mutation

The presence of a mutation in the MECP2 gene alone does not diagnose Rett syndrome. MECP2 mutations may cause conditions other than Rett syndrome.¹ Conversely, some people who meet the clinical diagnostic criteria for Rett syndrome do not have an identifiable MECP2 mutation.¹
When a male has a MECP2 mutation, he has no second normal copy of the gene to help lessen the effect of the mutation. This mutation can cause a severe disease called neonatal encephalopathy and these boys usually die before 2 years of age.\textsuperscript{1} Surviving males generally have an abnormal gait or truncal movements and severe speech delay and intellectual disability.\textsuperscript{2}

**Treatment**

Treatment for Rett syndrome is based on the symptoms and usually involves therapies to help with movement and communication.\textsuperscript{1} Medications can control difficult behavior and seizures, when present.\textsuperscript{1}

People with Rett syndrome are at risk for an irregular heart rhythm (arrhythmia). They may need heart monitoring and should avoid certain drugs that are known to affect the heart rhythm.\textsuperscript{1}

**Test information**

**Introduction**

Testing for Rett syndrome may include MECP2 sequencing, deletion/duplication analysis, or known familial mutation analysis.

**Sequence analysis**

MECP2 sequencing identifies an MECP2 gene mutation in about 80% of people with classic Rett syndrome and 40% of people with atypical Rett syndrome.\textsuperscript{1}

**Deletion/duplication analysis**

When MECP2 gene sequencing is normal, deletion and duplication analysis can be performed to look for other types of gene mutations. About 8% of people with classic Rett syndrome and 3% of people with atypical Rett syndrome have an MECP2 gene deletion.\textsuperscript{1}

**Known familial mutation analysis**

If a MECP2 mutation is found in an affected person, then other family members may be offered testing.\textsuperscript{1} Prenatal testing is available when the MECP2 mutation in the family is known.\textsuperscript{1-8}

**Guidelines and evidence**

**Introduction**

This section includes relevant guidelines and evidence pertaining to Rett syndrome.
testing.

**National Institute for Health and Clinical Excellence**

The National Institute for Health and Clinical Excellence (NICE) released evidence-based guidelines entitled *Autism spectrum disorder in under 19s: recognition, referral and diagnosis* in 2011 (updated in 2017). These guidelines state that Rett syndrome should be considered as a type of developmental regression. Genetic testing for such conditions should be considered on an individual basis.⁴

**American Academy of Pediatrics**

The consensus guideline from the American Academy of Pediatrics (2014)⁵ on the clinical genetic evaluation of a child with intellectual disability (ID) or global developmental delays (DD) and the American College of Medical Genetics (ACMG)⁶ 2013 Practice Guidelines for identifying the etiology of autism spectrum disorders state that:

“If the diagnosis is unknown and no clinical diagnosis is strongly suspected, begin with a stepwise evaluation including: chromosome microarray, specific metabolic testing, and Fragile X syndrome testing. If no diagnosis is established and the patient is female, then MECP2 sequencing, deletion, and duplication testing is appropriate.”⁶

**Criteria**

**Introduction**

Requests for Rett syndrome testing are reviewed using these criteria.

**MECP2 Known Familial Mutation Analysis**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Testing:
  - No previous genetic testing of MECP2, and
  - MECP2 mutation identified in 1st degree biologic relative, OR
- Prenatal Testing for At-Risk Pregnancies:
  - MECP2 mutation identified in a previous child of either parent.

**MECP2 Sequencing**

- Genetic Counseling:
Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  - No previous MECP2 sequencing, and
  - No known MECP2 mutation in family, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Meets clinical diagnostic criteria for classic Rett syndrome, atypical Rett syndrome or probable or possible Rett syndrome, and
  - Genetic testing is necessary because there is uncertainty in clinical diagnosis.

MECP2 Deletion/Duplication Analysis

- Previous testing:
  - No previous deletion/duplication analysis of MECP2, and
  - No mutations detected in full sequencing of MECP2.

References

Introduction

These references are cited in this guideline.


Procedures addressed

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What is SelectMDx

Definition

SelectMDx is a proprietary test that is designed to identify an individual’s risk of prostate cancer without the need for a biopsy.

- Prostate cancer is the most common cancer among men, with over 200,000 new cases identified each year in the United States.\(^1\)\(^2\) The median age at diagnosis is 66 years.\(^3\) Older men are more likely to be affected than younger men, and African American men have higher rates compared to men of other ethnic backgrounds.\(^3\)
- Screening programs for prostate cancer allow for its early detection. Screening is typically performed by prostate-specific antigen (PSA) test and digital rectal examination (DRE).\(^2\)
- Diagnosis is confirmed by prostate biopsy.\(^4\)\(^-\)\(^6\) Biopsy is typically performed by collection of approximately 12 needle biopsy cores.\(^6\)
- Initial biopsies only detect 65-77% of prostate cancers, and repeat biopsies are frequently performed.\(^7\)\(^,\)\(^8\) The false negative rate of biopsy may be as high as 25%.\(^9\)

Test information

- SelectMDx is a urine based assay that measures mRNA levels of DLX1 and HOXC6 to determine an individual’s risk of prostate cancer. KLK3 expression is used as an internal reference.\(^10\)
  - Higher levels of DLX1 and HOXC6 are associated with an increased risk of prostate cancer.
- This test is performed on first-void urine samples in patient’s post-digital rectal exam.
- Individuals with a high risk score on SelectMDx may need a biopsy.\(^10\)
• Individuals with a low risk score on this test may be able to avoid a biopsy.\textsuperscript{10}

**Guidelines and evidence**

• No specific evidence-based U.S. testing guidelines were identified.

• 2 clinical studies were identified for SelectMDx, which detail the results of a total of 3 studies describing the development and initial clinical validation of SelectMDx. One of the studies detailed the process with which the genetic markers utilized in the test were discovered, and the second study described the development of a risk score that incorporates the genetic markers with traditional risk factors, and the subsequent clinical validation of the risk score.\textsuperscript{11,12}

• Though the initial results are encouraging, there is an overall paucity of sufficient evidence currently available in the peer-reviewed literature to evaluate the clinical utility of this test. Only two studies have been published to date regarding the performance of SelectMDx. Of the three cohorts studied among the two publications, the first two were utilized to establish analytical validity, and the third was utilized for clinical validity. Furthermore, studies were conducted in the Netherlands; thus the results are not generalizable to men living in other countries, including the United States.\textsuperscript{1,2}

• Given that most of the results were focused on test validation, SelectMDx needs to be tested in larger and heterogeneous populations in diverse clinical settings to further assess the clinical validity and clinical utility of this test.\textsuperscript{1,2}

**Criteria**

• This test is considered investigational and/or experimental.

  o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.

  o In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

**References**

3. National Cancer Institute. SEER Stat Fact Sheet: Prostate Cancer. Available at: 

4. Centers for Disease Control and Prevention. Prostate Cancer. Available at: 


10. SelectMDx for Prostate Cancer. SelectMdx website. Available at: 
http://mdxhealth.com/selectmdx-prostate-cancer


SensiGene Fetal RHD Genotyping

Procedures addressed

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What is RhD

Definition

Women who are Rhesus D negative (RhD-) exposed to fetal RhD+ erythrocytes can develop anti-Rh antibodies (Rh-isoimmunization), which have the potential to permeate the placenta and cause hemolytic disease. If proper screening and treatment procedures are not carried out, Rh-isoimmunization has the potential to result in fetal/neonatal morbidity and mortality. Early identification of pregnancies with high risk for isoimmunization can better prepare both mothers and physicians for proper course of treatment, which may include fetal transfusion (during early pregnancy) or scheduled labor induction (for later pregnancies). The prevalence of RhD- blood type varies by race, where 15% of Caucasians, 5%-8% of African Americans, and 1%-2% of Asians and Native Americans are RhD-.¹

- Amniocentesis is the chief modality employed to assess fetal blood type; however, invasive procedures such as amniocentesis and chorionic villus sampling (CVS) have the potential to increase risk of fetal-maternal hemorrhage, which can lead to severe complications. An estimated 50% of previously sensitized women are considered low-risk for fetal RhD incompatibility; therefore, an alternative, non-invasive assay may reduce the number of unnecessary invasive procedures, as well as parental stress in cases involving sensitized women.² In cases where RhD hemolytic antibodies are not produced, identification of the fetus as RhD- is necessary to manage the use of anti-D prophylaxis immune globulin, and it may reduce need for intensive prenatal monitoring to predict and treat fetal anemia.³

Test information

- SensiGene® RHD Assay is a non-invasive, prenatal blood-test developed to evaluate circulating cell-free fetal DNA (ccffDNA) in maternal plasma to detect the incompatibility between the mother’s blood-type and the fetus’ blood-type, determining the presence of the fetal RhD genotype in RhD- mothers, as early as
10 weeks’ gestation. This assay may reduce the utilization of invasive procedures that may increase maternal sensitization.\textsuperscript{4}

- **SensiGene\textsuperscript{®} Fetal RHD Genotyping assay**, produced by Sequenom, isolates exons 4, 5, and 7 of the RhD gene (locus chromosome 1), psi-pseudogene in exon 4, and 3 targets on the Y chromosome (SRY, TTTY, DBY), all of which are known to be directly related to the genetic basis for RhD- phenotypes.

- Reasons to determine fetal RhD status utilizing maternal blood screening may include
  - avoiding the cost of paternity test/paternal genotyping
  - identifying fetal RhD status in the absence of maternal anti-Rh antigens
  - determining fetal RhD- status for parents opposed to immunization/vaccination
  - reducing the need for CVS or amniocentesis in RhD-sensitized patients

- This assay analyzes ccffDNA, which comprises approximately 3% to 6% of maternal plasma. Identification of specific exons of the RhD gene, not generally present in RhD- patients, has been found to predict an RhD+ fetus.\textsuperscript{4}

### Guidelines and evidence

- No U.S guidelines have been identified that address the use of SensiGene.

- The available evidence suggests that the clinical validity of the fetal RHD genotyping using the SensiGene\textsuperscript{®} assay is relatively high since the test demonstrates good accuracy in correctly predicting fetal RHD status.\textsuperscript{1,2,5-8} However, there is a risk of false positive results with use of the test.

- There is limited evidence regarding the clinical utility of the SensiGene RHD Assay to change clinician treatment decisions and ultimately improve patient-health outcomes compared with conventional modes of testing. Additional research is required to determine the performance of RhD genotyping with maternal plasma in improving patient health outcomes compared with amniocentesis and CVS.

- Furthermore, no professional society has published recommendations regarding the use of fetal RhD genotyping with ccffDNA in maternal plasma.

### Criteria

- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes...
(clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.

- In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

**References**


Sept9 Methylation Analysis for Colorectal Cancer

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What is Sept9 methylation analysis for colorectal cancer

Definition

Colorectal cancer (CRC) is one of the most common types of cancers, with over 136,000 new cases identified each year in the United States.\(^1\) It typically affects adults over 55 years old, with a median age at diagnosis of 68 years.\(^1\)

- Screening programs for CRC allow for its early detection. The earlier CRC is caught, the better chance a person has of surviving. Five year survival rates are 89.8% for localized cancer, 70.5% for cancer that has spread regionally, and 12.9% for CRC with distant metastasis.\(^1\)

- Standard recommended screening for CRC includes guaiac-based fecal occult blood test (gFOBT), fecal immunochemical test (FIT), multitargeted stool DNA test (FIT-DNA), colonoscopy, CT colonography, and flexible sigmoidoscopy. Screening begins at age 50 years and continues until at least age 75 for people at average risk for CRC.\(^2\)

- Although several screening tests have been endorsed and found to be cost-effective, compliance with CRC screening recommendations is limited. According to 2010 data from the Centers for Disease Control and Prevention (CDC), the percentage of adults over 50 years who reported their CRC screening was up to date ranged from 54.1% to 75.2%, depending on the state. The CDC estimates that 28 million Americans are not up-to-date on CRC screening.\(^3\)

- Two tests designed to detect colorectal cancer by analyzing Sept9 methylation will be addressed in this guideline: Epi proColon and ColoVantage.

- The Epi proColon Test (Epigenomics) is a Septin 9 assay that measures the presence of methylated Septin 9 DNA in a blood sample. It is intended to identify early stage colorectal cancer.\(^4\) It offers an alternative to current screening options.
The ColoVantage Test (Quest Diagnostics) is a Septin 9 assay that measures the presence of methylated Septin 9 DNA in a blood sample. This test “aids in the detection of colorectal cancer in patients non-adherent to current testing approaches.”

Test information
- Both Epi proColon and ColoVantage are performed on a blood sample. No bowel preparation or dietary or medication restrictions are required to complete either test.
- Both tests measure methylation of Septin 9 DNA, which is increased in colorectal cancer. Tumors often have increased methylation of Septin 9. When tumor DNA is shed into the bloodstream, this increase in methylation of Septin 9 can be found in the blood.
- Epi proColon provides a qualitative result: positive or negative. People who receive positive results should be referred for a diagnostic colonoscopy. Those with negative results can continue with standard CRC screening recommendations.

Guidelines and evidence
- There are currently no US guidelines that specifically address the use of either Epi proColon or ColoVantage testing.
- Current CRC cancer screening guidelines from the U.S. Preventative Services Task Force (USPSTF, 2016) recommend the use of gFOBT, FIT, FIT-DNA, colonoscopy, CT colonography, and flexible sigmoidoscopy for individuals ages 50 years to 75 years at average risk of colorectal cancer. These guidelines specifically state the following regarding Septin DNA testing:
  - “Although a serology test to detect methylated SEPT9 DNA was included in the systematic evidence review, this screening method currently has limited evidence evaluating its use (a single published test characteristic study met inclusion criteria, which found it had a sensitivity to detect colorectal cancer of <50%). It is therefore not included in this table.”
- For other age groups, the USPSTF guidelines recommend the following:
  - “For older adults aged 76 to 85 years, the benefits of screening for colorectal cancer decline, and the risk of experiencing serious associated harms increases. The most important consideration for clinicians and patients in this age group is whether the patient has previously been screened. Patients in this age group who have never been screened for colorectal cancer are more likely to benefit than those who have been previously screened.”
  - “Screening [in adults aged 76 to 85 years] would be most appropriate among adults who 1) are healthy enough to undergo treatment if colorectal cancer is detected and 2) do not have comorbid conditions that would significantly limit their life expectancy.”
o “The USPSTF does not recommend routine screening for colorectal cancer in adults 86 years and older. In this age group, competing causes of mortality preclude a mortality benefit that would outweigh the harms.”

• The U.S. Food and Drug Administration approved Epi proColon in 2016 as an in vitro diagnostic.

  o “The Epi proColon test is indicated to screen adults of either sex, 50 years or older, defined as average risk for CRC, who have been offered and have a history of not completing CRC screening.”

  o “The Epi proColon test is not intended to replace colorectal cancer screening tests that are recommended by appropriate guidelines (e.g., 2008 USPSTF guidelines) such as colonoscopy, sigmoidoscopy and high sensitivity fecal occult blood testing.”

  o “The Epi proColon test is not intended for patients who are willing and able to undergo routine colorectal cancer screening tests that are recommended by appropriate guidelines.”

  o “Tests that are available and recommended in the USPSTF 2008 CRC screening guidelines should be offered and declined prior to offering the Epi proColon test.”

• **Epi proColon**

  o “The performance of Epi proColon has been established in cross-sectional (i.e., single point in time) studies. Programmatic performance of Epi proColon (i.e., benefits and risks with repeated testing over an established period of time) has not been studied. Performance has not been evaluated for patients who have been previously tested with Epi proColon. Non-inferiority of Epi proColon programmatic sensitivity as compared to other recommended screening methods for CRC has not been established.”

  o “Screening with Epi proColon in subsequent years following a negative test result should be offered only to patients who after counseling by their healthcare provider, again decline CRC screening methods according to appropriate guidelines. The screening interval for this follow-up has not been established.”

  o The frequency interval that follow up Epi proColon testing should be performed has yet to be established.

  o A large, prospective multicenter trial (PRESEPT) evaluated men and women between the ages of 50 to 85 years who were at average risk for colorectal cancer.

    ▪ Clinical performance of the Epi proColon test in terms of sensitivity and specificity was based on 1544 samples from subjects whose colorectal cancer status was determined by colonoscopy.
Sensitivity was determined to be 68.2% with a specificity of 78.8%. Positive predictive value (PPV) was 2.4% with a negative predictive value (NPV) of 99.7%.

In 6 clinical validation studies, values of sensitivity and specificity of the Epi proColon test were reported. Sensitivity ranged from 72% to 79%, and specificity ranged from 82% to 96%. One study showed increasing sensitivity for higher CRC stages (~89% at Stage IV). In a comparative clinical validation trial, Epi proColon showed better sensitivity but worse specificity, when compared with gFOBT or FIT. Another study showed that the performance of the test is negatively impacted by risk factors frequently observed in CRC screening populations, such as early-stage disease, age > 65 years, diabetes, arthritis, and arteriosclerosis. Specifically, increased age was associated with increased rates of false positive and false negative results.

Results of a recent meta-analysis/systematic review indicate that the area under the receiver operating curve (AUC) for the pooled diagnostic accuracy results for Epi proColon test was 0.8709. In head-to-head comparisons, the AUC of the combined results of 1) Epi proColon and mSEPT 9 tests and 2) FOBT for CRC diagnosis were 0.7857 and 0.6571, respectively.

**ColoVantage**

The analytical validity, clinical validity, and clinical utility of the ColoVantage test for detecting CRC has not been established.

ColoVantage Plasma is currently undergoing clinical trials in Australia.

**Criteria**

**Epi proColon and ColoVantage testing** are considered investigational and/or experimental.

Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

**References**


Spinal Muscular Atrophy Testing

Procedures addressed

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What is spinal muscular atrophy

Definition

Spinal muscular atrophy (SMA) is a severe, autosomal recessive neuromuscular disease that affects 1 in 8000 to 1 in 10,000 people.\(^1,2\)

- SMA is caused by loss of lower motor neurons (anterior horn cells) in the spinal cord, resulting in progressive symmetrical muscle weakness and atrophy.\(^1-3\)
- SMA has historically been divided into three to five clinical subtypes based on age of onset and clinical course. While genetic testing has shown these clinical subtypes are not completely distinct, they are still widely used, and include:\(^1-3\)
  - Prenatal onset form (“Type 0” proposed) is characterized by polyhydramnios, decreased fetal movements, breech presentation, arthrogryposis multiplex congenita, respiratory failure at birth, and life span less than 6 months.
  - Type I (infantile or Werdnig-Hoffmann type) is the most common form (60-70% of cases). It presents before 6 months of age with death often before age 2 due to respiratory failure. Affected children have severe, generalized weakness and do not ever sit without support.
  - Type II (intermediate type) causes muscle weakness with onset after 6 months, although children often are able to sit alone and often survive early childhood. Intelligence is normal.
Type III (juvenile, Kugelberg-Welander type) is milder. Onset ranges from infancy to youth, but affected people usually walk unassisted albeit with frequent falls or trouble with stairs. Survival is prolonged and intelligence is normal.

Type IV (adult type) has much later onset with muscle weakness generally presenting at 20-30 years of age. People may or may not become wheelchair dependent, have normal lifespan and normal intelligence.

- SMA is caused by mutations in the SMN1 gene.
  - Large gene deletions (exon 7 +/- exon 8) cause SMA in the vast majority (95-98%) of affected individuals.\(^3\)
  - The remaining 2-5% of individuals with SMA have a deletion in one SMN1 gene and a different mutation in the other.\(^3\)

- SMN2 is another gene that is almost identical to SMN1 and located on the same chromosome. SMN2 gene mutations do not cause SMA. In fact, about 15% of unaffected people have no copies of the SMN2 gene. However, SMN2 has been shown to modify the disease severity in people with SMA. More copies (usually 3 or more) of SMN2 are associated with milder disease course. Individuals may have between 0-5 copies of SMN2.\(^3\)

- SMA is inherited in an autosomal recessive manner.
  - An affected person has two SMN1 gene mutations.\(^2,3\) Most do not have a known family history of the condition.
  - People with only one mutation in the SMN1 gene are called carriers. Carriers do not show symptoms of SMA, but have a 50% chance of passing on their mutation to their children.
  - SMA is present in all ethnic groups. About 1 in 40 to 1 in 60 people are carriers.\(^2\)
  - Two carriers of SMA have a 25% chance of having a child with the disorder.
  - About 2% of SMA patients have a de novo (new) mutation in one of their two SMN1 genes. In this case, only one parent is a carrier of SMA.\(^3\)

**Test information**

- **SMN1 Deletion Analysis**: Diagnostic testing in an affected individual begins with deletion or copy number analysis, which will identify a deletion of exon 7 in the SMN1 gene. For most affected individuals, both SMN1 genes will be missing exon 7. If both SMN1 genes do not have an exon 7 deletion, SMN1 gene sequencing can be considered.

- **SMN1 Sequencing Analysis** is typically performed in reflex, when one or no deletions are identified by deletion analysis. About 2-5% of affected individuals fall into this group. Sequencing detects the other mutation in virtually all cases.\(^2,3\)
• **Carrier testing** is usually performed by quantitative analysis that determines the dosage, or copy number, of exon 7-containing SMN1 genes.\(^3\-^5\)
  - Gene dosage ranges from one to three copies in most people. Asymptomatic carriers typically have one intact copy of the SMN1 gene and one SMN1 gene with the common deletion.
  - However, some unaffected carriers have two intact copies of the SMN1 gene. These may be on the same chromosome with no intact SMN1 gene on the other chromosome. Rare mutations and those carrying two SMN1 genes on the same chromosome will not be detected by gene dosage analysis. Therefore, a negative gene dosage analysis reduces the carrier risk but cannot completely rule out that a person is an SMA carrier.\(^3\-^6\)
  - The detection rate of carrier screening varies based on ethnicity, ranging from 71% in African Americans to 95% in Caucasians.\(^2\)

• **SMN2 Gene Copy Number Analysis** is performed by quantitative PCR to determine the number of copies of the SMN2 gene.
  - Most people have 0-3 copies of SMN2, although copy numbers as high as 5 have been reported.\(^3\)
  - The clinical severity of SMA can be influenced by the number of copies a person has of the SMN2 gene.\(^3\) Although a higher copy number of SMN2 is generally associated with a milder phenotype, SMA is still a highly variable disease. It is difficult to use SMN2 copy number to reliably predict the clinical manifestations of SMA in an affected person because sequence variation in SMN2 may also influence disease course regardless of copy number.\(^4\)

• **Known Familial Mutation Testing**: Once mutations have been identified in carriers or affected individuals, family members can be tested for the known familial mutation(s). Preimplantation diagnosis and prenatal testing can be considered when both parents are known SMA carriers.

**Guidelines and evidence**

**Diagnostic Testing**

• The International Standard of Care Committee for Spinal Muscular Atrophy issued a consensus statement in 2007 that stated the following:\(^5\)
  - “The first diagnostic test for a patient suspected to have spinal muscular atrophy should be the SMN gene deletion test.”\(^5\)
The current literature suggests SMN2 copy numbers correlate with spinal muscular atrophy clinical phenotypes. However, although a higher copy number of SMN2 is correlated with milder phenotype, phenotypes can vary substantially given SMN2 copy number. Therefore, predicting clinical phenotype using SMN2 copy number can be risky and is not currently recommended.

The European Federation of Neurological Societies (EFNS, 2011) published guidelines on the molecular diagnosis of various neuromuscular disorders. Regarding SMA testing they state:

- Screening for SMN1 deletions is indicated in SMA I-III to confirm the diagnosis and provide genetic counseling (Level B).
- In adult-onset SMA, genetic testing for SBMA should be considered in males with bulbar manifestations, gynecomastia and X-linked inheritance (Level B).
- As nearly all of these studies have a retrospective design and look for a specific mutation in a previously ascertained and clinically diagnosed cohort of patients, the highest achievable recommendation level will be B.

Carrier Testing

- The American College of Obstetricians and Gynecologists (ACOG, 2017) stated the following in regards to carrier testing for SMA in an updated Committee Opinion:
  - Screening for spinal muscular atrophy should be offered to all women who are considering pregnancy or are currently pregnant.

- The American College of Medical Genetics (ACMG, 2008; reaffirmed 2013) state the following regarding carrier testing for SMA:
  - Because SMA is present in all populations, carrier testing should be offered to all couples regardless of race or ethnicity. Ideally, the testing should be offered before conception or early in pregnancy. The primary goal is to allow carriers to make informed reproductive choices.

- In 2011 the Association of Molecular Pathology issued their statement on SMA carrier screening stating that it is “a technology on the threshold of feasibility.” They outlined 6 concerns, 2 of which related to population carrier frequencies, another regarding the need for pilot programs, need for genotype/phenotype research, and another about technical issues with SMN1/SMN2 as outlined above.

Spinraza

- In 2016, the FDA approved the use of Spinraza in individuals with SMA. While the FDA label does not require SMN2 copy number analysis, the study of 121 patients on which FDA approval was based used the following inclusion criteria:
  - 5q SMN1 homozygous gene deletion or mutation or compound heterozygous mutation
o 2 copies of the SMN2 gene (98% of enrolled patients had 2 copies of SMN2)
o Onset of SMA symptoms at or before 6 months of age
o No hypoxemia at baseline screening at age 7 months or younger

Criteria

SMN1 Exon 7 Deletion

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o No previous genetic testing of the SMN1 gene, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Child with hypotonia and weakness (generally symmetrical, proximal more than distal), or
  o Young adult (through twenties) onset of weakness more severely affecting the legs than arms (may be associated with frequent falls, difficulty with stairs), and
  o No obvious signs of different neurological disorder, OR

• Carrier Screening:
  o SMN1 exon 7 deletion testing is not suitable for carrier screening. SMN1/SMN2 dosage analysis is necessary, OR

• Prenatal Testing:
  o Both parents are carriers of an SMA mutation (at least one of which is an exon 7 deletion mutation), AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

SMN1/SMN2 Dosage Analysis

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Testing:
  o No previous genetic testing of the SMN1 gene in the carrier testing setting, or
- Non-diagnostic results from SMN1 exon 7 deletion testing (not homozygous SMN1 deletion) in the diagnostic setting, AND

- Diagnostic Testing for Symptomatic Individuals:
  - Index of suspicion for SMA remains high despite non-diagnostic SMN1 exon 7 deletion testing based on:
    - Proximal greater than distal weakness, and
    - Normal creatine kinase (CK), and
    - Neurogenic EMG, OR

- Carrier Screening:
  - Be of reproductive age, and
  - Have potential and intention to reproduce, OR

- Prenatal Testing:
  - SMN1/SMN2 Dosage Analysis is not suitable for preimplantation/prenatal diagnosis. Other forms of SMA testing may be indicated based on the mutation status of parents. See those sections for guidance, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**SMN1 Known Familial Mutation Analysis**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous genetic testing for known SMN1 family mutation(s), AND

- Diagnostic Testing for Symptomatic Individuals:
  - Known family SMN1 point mutation(s) in biological relative, OR

- Carrier Screening
  - Known family SMN1 point mutation(s) in biological relative, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**SMN1 Sequencing**

- Genetic Counseling:
• Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o SMN1 exon 7 deletion testing did not reveal a homozygous SMN1 deletion and SMN1/SMN2 gene dosage analysis identified a single copy of SMN1 exon 7 in the diagnostic setting, or
  o SMN1/SMN2 gene dosage analysis did not confirm carrier status of an exon 7 deletion in the carrier testing setting, AND

• Diagnostic Testing for Symptomatic Individuals:
  o Individual suspected to have compound heterozygous SMA based previous test results and:
    o Proximal greater than distal weakness, and
    o Normal creatine kinase (CK), and
    o Neurogenic EMG, OR

• Carrier Screening:
  o Have one of the following increased risk indication with a noninformative SMN1/SMN2 gene dosage analysis result:
    ▪ Have a reproductive partner who is a carrier of SMA, or
    ▪ Have a reproductive partner with SMA, OR

• Prenatal Testing:
  o SMN1 full gene sequencing is not generally necessary for preimplantation/prenatal diagnosis as parental mutation status should have already been determined with SMN1 exon 7 deletion testing +/- SMN1 known familial variant analysis, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

**SMN2 Gene Copy Analysis**

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Member meets the following criteria:
  o Member has a genetically confirmed diagnosis of SMA, and
  o Member has a diagnosis of either SMA Type 1 or SMA Type 2, and
  o Member has not had previous SMN2 copy number analysis performed, and
Treatment with Spinraza is being considered, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

Exclusions

Genetic testing is not approved for SMN2 gene copy analysis for the purposes of predicting SMA prognosis because it is currently considered experimental, investigational or is unproven.

References


Tay-Sachs Disease Testing

Procedures addressed

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What is Tay-Sachs disease

Definition

Tay-Sachs disease is a neurodegenerative genetic disorder. Affected individuals typically present in infancy with progressive weakness, loss of motor skills, decreased attentiveness, and increased startle response between 3-6 months of age. Eventually they develop seizures and blindness, with death in early childhood. There is no cure for Tay-Sachs disease and treatment is supportive.

- Rare, less severe, Tay-Sachs variants exist that are associated with later onset, and less progressive symptoms, and cause more variable neurological problems. These variants include juvenile, chronic, and adult-onset forms.
- Tay-Sachs disease is caused by mutations in the HEX A gene. HEX A gene mutations lead to reduced activity of the β-hexosaminidase A enzyme, allowing toxic substances to build up in the cells of the brain and spinal cord. Eventually, neurons are destroyed, causing the signs and symptoms of Tay-Sachs disease.
- Before widespread carrier screening, Tay-Sachs disease affected about 1 in 3,600 Ashkenazi Jewish births.
- Tay-Sachs disease is an autosomal recessive disorder. An affected individual must inherit a HEX A gene mutation from both parents.
  - Individuals who inherit only one mutation are called carriers. Carriers do not show symptoms of Tay-Sachs disease, but have a 50% chance of passing on the mutation to their children.
  - About 1 in 30 Ashkenazi Jewish individuals are carriers for Tay-Sachs disease.
Two carriers of Tay-Sachs disease have a 25% chance of having a child with the disorder.

- Individuals at increased risk to have a child with Tay-Sachs should routinely be offered carrier screening. This includes those with:
  - Ashkenazi Jewish, French Canadian, or Cajun ancestry
  - A family history of Tay-Sachs disease (regardless of ethnicity)
  - A partner who is a known carrier of Tay-Sachs (or affected with a late-onset variant)
- Carrier screening for Tay-Sachs disease is widely available as part of an “Ashkenazi Jewish Panel” that includes several other genetic diseases that are more common in this population (See the Ashkenazi Jewish Carrier Screening Test Summary).

Test information

- **Hexosaminidase A (HEXA) enzyme analysis** measures the activity of HEXA in the serum or white blood cells. This test is used both for diagnostic testing of symptomatic individuals, and carrier screening.
  - Individuals with classic Tay-Sachs have little to no HEX A enzyme activity in the presence of normal or elevated activity of the beta-hexosaminidase B (HEX B) isoenzyme. HEX A enzyme activity levels correctly diagnose the vast majority of people with all forms of Tay-Sachs disease.
  - Carriers have about 50% of the normal level of HEX A activity. HEX A enzyme analysis detects 97%-98% of carriers, regardless of ethnicity. Enzyme analysis is recommended as the first step for all people being screened.
  - A small percentage of individuals will get a false positive result by enzyme analysis. This means that they have enzyme activity that appears to be in the carrier range, but they are not actually carriers of a disease-causing mutation. These individuals carry a “pseudodeficiency allele.” Inconclusive enzyme analysis results are also possible where enzyme activity is in the overlap range between carrier and normal levels. If HEXA enzyme analysis is abnormal or inconclusive, HEXA mutation analysis may be considered.
  - Prenatal diagnosis in an at-risk pregnancy can be performed by HEXA enzyme activity measurement in a fetal sample obtained from chorionic villus sampling (CVS) or amniocentesis.

- **HEXA mutation panel.** This genetic test looks for the most common HEXA gene mutations (such as +TATC1278, +1 IVS 12, +1 IVS 9, G269, R247W, and R249W), which account for up to 98% of all Ashkenazi Jewish Tay-Sachs mutations. The detection rate of standard HEXA mutation panels is much lower in other ethnicities. Some panels include mutations more common in other at-risk ethnic groups (e.g., a
7.6kb deletion more common in French Canadians). \(^1\) If using mutation panels in non-Ashkenazi Jewish, providers should confirm those mutation panels include any ethnicity-specific mutations.

- **HEXA sequencing** analyzes the entire coding region of the HEXA gene and finds the vast majority of HEXA mutations that cause Tay-Sachs disease. Sequencing is most useful for individuals diagnosed by enzyme analysis, but for whom mutation panels found only one or no disease-causing mutations. \(^1\)

- **HEXA known familial mutation analysis:** Once the disease-causing mutations have been identified in an affected family member or known carriers, other at-risk relatives can be tested for just those mutations. Prenatal diagnosis can be performed by mutation analysis if both parental mutations are known. This method may also be used in reflex if HEX A enzyme activity testing is performed first and is inconclusive. \(^1\)

### Guidelines and evidence

- Professional guidelines support population-based Tay-Sachs carrier screening for those at increased risk. They do not generally recommend a specific testing strategy (enzyme and/or mutation analysis) for Ashkenazi Jewish individuals, but do recommend enzyme analysis as a first-line test for non-Jewish individuals. \(^2,3\)

- Consensus guidelines from the American College of Obstetricians and Gynecologists (ACOG, 2005) recommend: \(^3\)
  
  - “Screening for TSD should be offered before pregnancy if both members of a couple are of Ashkenazi Jewish, French–Canadian, or Cajun descent. Those with a family history consistent with TSD also should be offered screening.”
  
  - “When one member of a couple is at high risk (i.e., of Ashkenazi Jewish, French–Canadian, or Cajun descent or has a family history consistent with TSD) but the other partner is not, the high-risk partner should be offered screening….If the high-risk partner is determined to be a carrier, the other partner also should be offered screening. If the woman is already pregnant, it may be necessary to offer screening to both partners simultaneously to ensure that results are obtained promptly and that all options are available to the couple.”
  
  - “Biochemical analysis should be used for individuals in low-risk populations.”

- Consensus guidelines from the American College of Medical Genetics (ACMG, 2008) recommend carrier screening for a group of disorders that includes Tay-Sachs disease when at least one member of the couple is Ashkenazi Jewish and that couple is pregnant or planning pregnancy. \(^2\)

- No evidence-based U.S. testing guidelines that address Tay-Sachs diagnostic testing have been identified.

- A 2006 comprehensive literature review states that: “The diagnosis of hexosaminidase A deficiency relies upon the demonstration of absent to near-
absent beta-hexosaminidase A (HEX A) enzymatic activity.”¹ HEXA mutation analysis can be used in follow-up to resolve inconclusive results or to identify the familial mutations for reproductive purposes.¹

- Professional guidelines generally recommend prenatal testing for Tay-Sachs disease in any of the following situations:¹⁻⁴
  - HEXA enzyme activity testing revealed both parents to be carriers of Tay-Sachs disease and pseudodeficiency alleles have been ruled out.
  - Disease-causing mutations in HEXA have been identified in both parents.
  - One parent is a known carrier and HEXA enzyme activity testing in the other parent was inconclusive.
  - The mother is a known carrier and the father is unknown or unavailable for testing.

- Guidelines do not generally recommend a specific testing strategy (HEX A enzyme activity and/or mutation analysis). However, the clinical circumstances may deem one strategy more accurate than the other. For instance, mutation analysis is most accurate if both of the parental mutations are known.

- The American College of Obstetricians and Gynecologists (ACOG, 2005) guidelines for Tay-Sachs disease state: “If both partners are determined to be carriers of Tay-Sachs disease, genetic counseling and prenatal diagnosis should be offered.”³

- The American College of Obstetricians and Gynecologists (ACOG, 2009) guidelines for Ashkenazi Jewish carrier screening state: “Carrier screening for TSD, Canavan disease, cystic fibrosis, and familial dysautonomia should be offered to Ashkenazi Jewish individuals before conception or during early pregnancy so that a couple has an opportunity to consider prenatal diagnostic testing options. If the woman is already pregnant, it may be necessary to screen both partners simultaneously so that the results are obtained in a timely fashion to ensure that prenatal diagnostic testing is an option... Carrier couples should be informed of the disease manifestations, range of severity, and available treatment options. Prenatal diagnosis by DNA-based testing can be performed on cells obtained by chorionic villus sampling and amniocentesis.”⁶

Criteria

HEXA Known Familial Mutation Analysis

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - No previous molecular genetic testing of HEXA, AND
• Carrier Screening:
  o Known family mutation in HEXA identified in 1st, 2nd, or 3rd degree biologic relative(s), OR

• Prenatal Testing for At-Risk Pregnancies:
  o HEXA mutation identified in both biologic parents, and
  o Pseudodeficiency allele mutation has been ruled out, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

HEXA Targeted Mutation Analysis for Common Mutations and Pseudodeficiency Alleles

• Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  o This same test has not been performed previously, and
  o No known HEXA mutation in family, AND

• Diagnostic Testing:  
  o Abnormal or indeterminate HEXA enzymatic activity in serum, white blood cells, or other tissues, and clinical symptoms of TSD, but diagnosis remains uncertain, or
  o Asymptomatic individual with abnormal HEXA enzymatic activity in order to test for a pseudodeficiency allele, or
  o Children under the age of 6 months with
    ▪ Progressive weakness and loss of motor skills, or
    ▪ Decreased attentiveness, or
    ▪ Increased startle response, or
    ▪ Macular cherry red spot, or
    ▪ Seizures, or
    ▪ Blindness, or
  o Young children with
    ▪ Ataxia and incoordination, or
    ▪ Speech, life skills and cognition decline, or
Spasticity and seizures, or
Loss of vision, sometimes with:
  • Cherry red spot, or
  • Optic atrophy, or
  • Retinitis pigmentosa, or

  Adolescents/adults (and SMA type Kugelberg-Welander disease or early onset ALS have been ruled out) with
  • Progressive dystonia, or
  • Spinocerebellar degeneration, or
  • Motor neuron disease, or
  • Cognitive dysfunction, dementia, recurrent psychotic depression or bipolar symptoms, or

  French Canadian, Cajun, or Old Order Amish descent regardless of symptoms, OR

• Preconception/Prenatal Carrier testing
  • Ashkenazi Jewish descent, and
  • Intention to reproduce, AND

• Carrier testing for Individuals with Family History or Partners of Carriers:
  • 1st, 2nd, or 3rd degree biologic relative with Tay-Sachs clinical diagnosis, family mutation unknown, and affected relative unavailable for testing, or
  • Partner is monoallelic or biallelic for HEXA mutation, and
  • Have the potential and intention to reproduce, AND

• Rendering laboratory is a qualified provider of service per the Health Plan policy.

HEXA Sequencing

• Genetic Counseling:
  • Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

• Previous Genetic Testing:
  • No mutations found on targeted mutation analysis, and
  • No previous full sequencing of HEXA, AND

• Diagnostic Testing:

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- Abnormal or indeterminate HEXA enzymatic activity in serum, white blood cells, or other tissues, and clinical symptoms of TSD, but diagnosis remains uncertain, OR
- Children under the age of 6 months with one or more of the following:
  - Progressive weakness and loss of motor skills,
  - Decreased attentiveness
  - Increased startle response
  - Macular cherry red spot
  - Seizures
  - Blindness, or
- Young children, with one or more of the following:
  - Ataxia and incoordination
  - Speech, life skills and cognition decline
  - Spasticity and seizures
  - Loss of vision, sometimes with:
    - Cherry red spot
    - Optic atrophy
    - Retinitis pigmentosa, or
- Adolescence/adult (and SMA type Kugelberg-Welander disease or early onset ALS has been ruled out), with one or more of the following:
  - Progressive dystonia
  - Spinocerebellar degeneration
  - Motor neuron disease
  - Cognitive dysfunction, dementia, recurrent psychotic depression or bipolar symptoms, and
- Carrier testing for Individuals with Family History or Partners of Carriers:
  - 1st, 2nd, or 3rd degree biologic relative with Tay-Sachs clinical diagnosis, and familial mutation unknown, and affected relative unavailable for testing, or
  - Partner is monoallelic or biallelic for a HEXA mutation, and
- Have the potential and intention to reproduce, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy.

**References**


Thoracic Aortic Aneurysms and Dissections (TAAD) Panel Testing

Procedures addressed

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What are thoracic aortic aneurysms and dissections (TAAD)

Definition

The major cardiac problems seen in individuals with Thoracic Aortic Aneurysms and Dissections (TAAD) include “dilatation of the ascending thoracic aorta at the level of the sinuses of Valsalva or ascending aorta or both”¹ and “dissections of the thoracic aorta involving either the ascending (Stanford type A dissections) or descending aorta (Stanford type B)”¹ In some cases, vascular manifestations may be the only manifestation.¹

- TAAD can be diagnosed by various imaging studies, including echocardiography, computed tomography (CT), MRI, and angiography.¹
- The age of aortic dissection and the severity of the disease can range.¹ Treatment for TAAD may involve medications. Surgical repair of the aorta may be necessary in some cases to help prevent aortic dissection.¹
- Cardiac problems seen with TAAD are associated with approximately 15,000 deaths every year.¹
- Genetic testing can be helpful to determine if there is an underlying genetic condition causing the TAAD. There are many genes which can predispose someone to TAAD. Some of these genes are associated with specific genetic conditions which may require additional management or surveillance. Medical management, including timing of surgery, may differ based on the underlying genetic etiology.²,³ In many cases, a careful clinical examination by a specialist familiar with clinical features of these conditions can help to point toward one
condition. In these cases, testing for gene(s) associated with a single condition would be most appropriate.

- Specific genetic conditions that have TAAD as a clinical manifestation:
  
  - **Marfan syndrome (MFS)** – MFS is an autosomal dominant disorder that affects connective tissue in many parts of the body. MFS is caused by mutations in the FBN1 gene. Approximately 70-93% of people meeting diagnostic criteria for Marfan will have a mutation in this gene. Diagnostic criteria, called the Ghent criteria, exists for Marfan syndrome. Major manifestations of the disease include aortic enlargement and ectopia lentis. Other features include, but are not limited to, bone overgrowth and joint laxity, long arms and legs, scoliosis, sternum deformity (pectus excavatum or carinatum), long thin fingers and toes, dural ectasia (stretching of the dural sac), hernias, stretch marks on the skin, and lung bullae. Symptoms can present in males or females at any age. Symptoms typically worsen over time. Infants who present with symptoms typically have the most severe disease course.

  - **Loeys-Dietz syndrome (LDS)** - LDS is an autosomal dominant disorder that affects many parts of the body. LDS is mostly caused by mutations in either the TGFBR1 gene (20%) or TGFBR2 gene (70%). However, a small percentage of people with LDS may have mutations in the SMAD3 gene (5%) or TGFB2 gene (1%). Major manifestations of this condition include “vascular findings (dilatation or dissection of the aorta, other arterial aneurysms or tortuosity), skeletal findings (pectus excavatum or pectus carinatum, scoliosis, joint laxity or contracture, long thin fingers and toes, cervical spine malformation and/or instability), craniofacial findings (widely spaced eyes, bifid uvula/cleft palate, craniosynostosis), and cutaneous findings (translucent skin, easy bruising, dystrophic scars).” Given that there is no clinical diagnostic criteria established for LDS, genetic testing can help with the diagnosis.

  - **Ehlers-Danlos syndrome, Type IV (EDS type IV)** – EDS type IV is an autosomal dominant condition. It is caused by mutations in the COL3A1 gene. Major manifestations of this condition include “thin, translucent skin; easy bruising; characteristic facial appearance (in some individuals); and arterial, intestinal, and/or uterine fragility.” Many adults present with the following symptoms: vascular dissection or rupture, gastrointestinal perforation, or organ rupture. Infants and children may present with congenital dislocation of the hips, clubfoot, inguinal hernia, pneumothorax, and/or recurrent joint subluxation or dislocation.
- **Familial TAAD (TGFBR2, TGFBR1, MYH11, ACTA2, MYLK, and SMAD3)** – Familial TAAD is diagnosed based on the following: "dilatation and/or dissection of the thoracic aorta, absence of clinical features of MFS, LDS or EDS Type IV, and a positive family history of TAAD." Only 20% of people with a clinical diagnosis of Familial TAAD will have a mutation found in one of the above genes.

**Test information**

- Many laboratories offer testing for at least 9 genes that have been associated with TAAD in their panels, including the genes that cause MFS, LDS, EDS type IV and Familial TAAD. Detection rates of expanded panels vary by laboratory and depend on the genes included and the methods used for testing. In most cases, a careful and comprehensive clinical evaluation along with imaging studies will point to a specific diagnosis. Testing for conditions that are clinically indicated is the most appropriate place to start. Testing multiple genes, without supporting clinical features, has the potential to obtain results which may be hard to interpret. The chance that a variant of uncertain significance will be found increases as more genes are tested. However, given that many of the symptoms of conditions associated with TAAD overlap, if a person presents with overlapping features of more than one condition, a panel approach may be considered.

- Without symptoms of a specific genetic condition associated with TAAD, mutations in the ACTA2 gene are the most common. Mutations in this gene account for approximately 10-14% of Familial TAAD.

- Once a mutation is identified in a family member, the known familial mutation can be specifically identified in asymptomatic or symptomatic family members.

**Guidelines and evidence**

- The European Society of Cardiology (ESC, 2014) stated the following:
  
  - "Once a familial form of TAAD is highly suspected, it is recommended to refer the patient to a geneticist for family investigation and molecular testing." (Class I, Level C)

- The Canadian Cardiovascular Society (2014) stated the following:
  
  - "We recommend genetic screening for TAD-associated genes in non-BAV aortopathy index cases to clarify the origin of disease and improve clinical and genetic counselling (Strong recommendation, moderate quality evidence)."
  
  - "We recommend complete aortic imaging at initial diagnosis and at 6 months for patients with LDS or a confirmed genetic aortopathy (e.g., TGFBR1/2, TGFB, SMAD3, ACTA2, or MYH11) to establish if enlargement is occurring (Strong Recommendation, Moderate-Quality Evidence)."
o “We recommend that genetic counselling and testing be offered to first-degree relatives of patients in whom a causal mutation of a TAD-associated gene is identified. We recommend that aortic imaging be offered only to mutation carriers (Strong Recommendation, Low-Quality Evidence).”


o Predictive genetic testing for at-risk relatives is addressed in the following guidelines statement:
  ▪ “If the mutant gene (FBN1, TGFBR1, TGFBR2, COL3A1, ACTA2, MYH11) associated with aortic aneurysm and/or dissection is identified in a patient, first-degree relatives should undergo counseling and testing. Then, only the relatives with the genetic mutation should undergo aortic imaging.” [Evidence level I: “Evidence from only expert opinion, case studies, or standard of care.” Recommendation classification C: “Recommendation that procedure or treatment is useful/effective.”]⁹

o ACTA2 sequencing is addressed in the following guidelines statement:
  ▪ “Sequencing of the ACTA2 gene is reasonable in patients with a family history of thoracic aortic aneurysms and/or dissections to determine if ACTA2 mutations are responsible for the inherited predisposition (Pannu et al., 2005; Guo et al., 2007; Zhu et al., 2006; Loeys et al., 2006; Stheneur et al., 2008; Guo et al., 2009).” [Evidence level IIa: “Only diverging expert opinion, case studies, or standard of care.” Recommendation classification B: “Recommendation in favor of treatment or procedure being useful/effective.”]⁹

o Additional genetic testing is addressed in the following guidelines statement:
  ▪ “Sequencing of other genes known to cause familial thoracic aortic aneurysms and/or dissection (TGFBR1, TGFBR2, MYH11) may be considered in patients with a family history and clinical features associated with mutations in these genes (Pannu et al., 2005; Guo et al., 2007; Zhu et al., 2006; Loeys et al., 2006; Stheneur et al., 2008; Guo et al., 2009).” [Evidence level IIb: “Greater conflicting evidence from single randomized trial or nonrandomized studies.” Recommendation classification B: “Recommendation's usefulness/efficacy less well established.”]⁹
“Patients with Loeys-Dietz syndrome or a confirmed genetic mutation known to predispose to aortic aneurysms and aortic dissections (TGFBR1, TGFBR2, FBN1, ACTA2, or MYH11) should undergo complete aortic imaging at initial diagnosis and 6 months thereafter to establish if enlargement is occurring. (Level of Evidence: C)."

Criteria
Known Familial Mutation(s) for TAAD

- Genetic Counseling
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous genetic testing for TAAD, AND
- Diagnostic or Predisposition Testing for Symptomatic or Presymptomatic Individuals:
  - TAAD family mutation in 1st degree biological relative, AND
- Rendering laboratory is a qualified provider for service per the Health Plan policy

**NOTE:** Since symptoms may occur in childhood, testing of children who are at-risk for a pathogenic mutation may be considered.

TAAD Genetic Testing Sequencing Panel

Gene panels that are specific to TAAD that include the following genes will be eligible for coverage according to the criteria outlined in this policy: FBN1, TGFBR1, TGFBR2, COL3A1, MYH11, ACTA2, SLC2A10, SMAD3, and MYLK. This sequencing panel will only be considered for coverage when billed under the appropriate panel CPT code: 81410. For criteria specific to Marfan syndrome, please see the policy *Marfan Syndrome Genetic Testing*.

- Genetic Counseling
  - Pre and post-test counseling by an appropriate provider (as deemed by the Health Plan policy), AND
- Previous Genetic Testing:
  - No previous panel testing for TAAD, AND
- Diagnostic Testing for Symptomatic Individuals:
  - Cardiology examination consistent with a diagnosis of TAAD, and
  - Clinical features are not sufficiently specific to suggest a single condition, and
The results of the test will directly impact the diagnostic and treatment options that are recommended for the patient, AND

- Rendering laboratory is a qualified provider for service per Health Plan policy

**TAAD Genetic Testing Duplication/Deletion Panel**

This duplication/deletion panel will only be considered for coverage when billed under the appropriate panel CPT code: 81411.

- Criteria for TAAD Genetic Testing Sequencing panel met, AND
- No mutations found in TAAD Sequencing panel, AND
- No previous deletion/duplication analysis for TAAD

**Billing and reimbursement considerations**

- This guideline addresses testing specifically for TAAD. Additional indications are addressed in the *Hereditary Connective Tissue Disorder Testing* guideline.
- When multiple CPT codes are billed for components of a panel and there is a more appropriate CPT code representing the panel, eviCore will redirect to the panel code(s).
- If the laboratory will not accept redirection to a panel code, the medical necessity of each billed component procedure will be assessed independently.
  - In general, only a limited number of panel components that are most likely to explain the member's presentation will be reimbursable. The remaining panel components will not be reimbursable.
  - When a TAAD multi-gene panel is billed with multiple stacked codes, only the following genes may be considered for reimbursement:
    - TGFBR2
    - TGFBR1
    - MYH11
    - ACTA2
    - MYLK
    - SMAD3

**References**


ThyGenX and ThyraMIR miRNA Gene Expression Classifier

Procedures addressed

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<tr>
<td>ThyraMIR miRNA Gene Expression Classifier</td>
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What are ThyGenX and ThyraMIR gene expression classifier

Definition

ThyGenx and ThyraMIR miRNA Gene Expression Classifier are designed to aid in the diagnosis of thyroid nodules as either malignant or benign.

- Thyroid nodules are a common occurrence, especially in an aging population. Fine-needle aspiration (FNA) with accompanying cytology examination is the standard method for distinguishing between benign and malignant nodules and subsequent removal of tumors. Approximately 25% of thyroid nodules examined using FNA and traditional cytology examination are considered indeterminate. Clinicians are then faced with the decision to either remove the nodule unnecessarily or leave a potentially malignant nodule in place.¹

- Additional diagnostic procedures have been developed to help further classify indeterminate nodules as either benign or malignant. These procedures usually involve assessment of known genetic point mutations or through the expression activity of microRNA.¹

Test information

- Thyroid nodules are traditionally assessed through inspection of cell cytology; however, some aspirate samples may be indeterminate. ThyraMIR uses an algorithm of 10 microRNAs previously validated using nodules with known malignancy to assist in determining if indeterminate cytology is malignant. It is used in conjunction with ThyGenX, an oncogene panel that analyzes 8 genes that are most commonly altered in thyroid cancer (papillary and follicular carcinoma).²
When a thyroid fine needle aspirate sample is found to be indeterminate, the ThyGenX test is run on the sample. If the ThyGenX test result is negative for malignancy, the ThyraMIR miRNA classifier test is then used to increase the overall sensitivity and specificity of the test combination. The overall test result is either positive or negative for malignancy.

**Guidelines and evidence**

- No specific evidence-based U.S. testing guidelines were identified.

- The National Comprehensive Cancer Network (NCCN, 2016) states the following regarding molecular testing for thyroid cancer:
  
  - "The diagnosis of follicular carcinoma requires evidence of either vascular or capsular invasion, which cannot be determined by FNA. Molecular diagnostics (category 2B) may be useful to allow reclassification of follicular lesions (follicular neoplasm or follicular lesions of undetermined significance (FLUS)) as either more or less likely to be benign or malignant based on genetic profile. If molecular testing (category 2B) in conjunction with clinical and ultrasound features suggests papillary thyroid carcinoma, especially in the case of BRAF V600E, see (PAP-1). Use molecular markers with caution and caveat)."\(^3\)
  
  - "Molecular diagnostic testing may include multigene assays (e.g. the gene expression classifier) or individual mutational analysis. The gene expression classifier measures the expression of at least 140 genes."\(^3\)
  
  - "BRAF V600E mutation analysis was recommended by some panelists for the evaluation of thyroid nodules (not restricted to the follicular lesions)."\(^3\)

- The current evidence base consists of two analytical validity studies and two clinical validity studies.\(^4-7\)
  
  - Clinical validity studies reported area under the receiver operator curve (AUC) values ranging from 0.89 to 0.94; the test correctly classified 92% of benign lesions as low risk or negative and correctly classified 92% of malignant lesions as high risk or positive.\(^7\)
Another study reported that ThyraMIR correctly identified 64% of malignant lesions and 96% of benign lesions. The sensitivity and specificity of the test was reported as 89% (95% confidence interval [CI], 73-97%) and 85% (95% CI, 75-92%), respectively. With a 32% prevalence rate, 61% of the results were considered benign, generating a negative predictive value (NPV) of 94% (95% CI, 85-98%).

- Preliminary clinical validity studies indicate reasonable diagnostic accuracy; however, additional studies are needed to confirm these early findings. Furthermore, there are currently no available clinical utility studies that evaluated the effects of on patient-relevant outcomes (survival, quality of life). Additional research is necessary to assess how testing will be used in the disease management of patients with thyroid cancer.

Criteria
- This test is considered investigational and/or experimental.
  - Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  - In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References
1. UPMC University of Pittsburgh Medical Center. ThyroSeq® - Thyroid Cancer Next-Generation Sequencing Panel. http://mgp.upmc.com/Applications/mgp/Home/Test/ThyroSeq_Details

Procedures addressed

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What are thyroid nodules

Definition

Thyroid nodules are a common occurrence, especially in an aging population. Fine-needle aspiration (FNA) with accompanying cytology examination is the standard method for distinguishing between benign and malignant nodules and subsequent removal of tumors. Approximately 25% of thyroid nodules examined using FNA and traditional cytology examination are considered indeterminate. Clinicians are then faced with the decision to either remove the nodule unnecessarily or leave a potentially malignant nodule in place.¹

- Additional diagnostic procedures have been developed to help further classify indeterminate nodules as either benign or malignant. These procedures usually involve assessment of known genetic point mutations or through the expression activity of microRNA.¹

Test information

- ThyroSeq is designed to aid in the diagnosis of thyroid nodules as either malignant or benign.
- ThyroSeq is a gene sequencing panel used on thyroid cells obtained using fine needle aspiration to detect genetic mutations known to be associated with thyroid cancer. The panel sequences gene fusions and point mutations in over 60 genes related to thyroid cancer. The test is used when cytological examination of FNA cells are indeterminate, helping to identify malignant nodules and guide therapy, as well as avoiding surgery for those with benign nodules.
- The ThyroSeq v.2 test has an overall negative predictive value of 96% and a positive predictive value of 83%.² Results from the sequencing test give a prediction as either a positive or negative for malignancy.
• Depending on the Bethesda thyroid cancer grade the following risk of metastasis percentages are observed with negative and positive ThyroSeq results:\(^2\)
  
  o Bethesda III Atypia of Undetermined Significance/Follicular Lesion of Undetermined Significance (AUS/FLUS) with 14% risk of metastasis:
    ▪ ThyroSeq Negative: 3% risk of metastasis
    ▪ ThyroSeq Positive: 82% risk of metastasis
  
  o Bethesda IV Follicular neoplasm/suspicion for a follicular neoplasm (FN/SFN) with 27% risk of metastasis:
    ▪ ThyroSeq Negative: 4% risk of metastasis
    ▪ ThyroSeq Positive: 83% risk of metastasis
  
  o Bethesda V squamous metaplasia (SM) with 54% risk of metastasis
    ▪ ThyroSeq Negative: 20% risk of metastasis
    ▪ ThyroSeq Positive: 95% risk of metastasis

Guidelines and evidence

• No evidence-based U.S. testing guidelines were identified that specifically address the use of ThyroSeq.

• The National Comprehensive Cancer Network (NCCN, 2017) states the following regarding molecular testing for thyroid cancer:
  
  o “The diagnosis of follicular carcinoma requires evidence of either vascular or capsular invasion, which cannot be determined by FNA. Molecular diagnostics (category 2B) may be useful to allow reclassification of follicular lesions (follicular neoplasm or follicular lesions of undetermined significance (FLUS)) as either more or less likely to be benign or malignant based on genetic profile. If molecular testing (category 2B) in conjunction with clinical and ultrasound features suggests papillary thyroid carcinoma, especially in the case of BRAF V600E, see (PAP-1). Use molecular markers with caution and caveat).”\(^3\)
  
  o “The NCCN Panel recommends molecular diagnostic testing for evaluating FNA results that are suspicious for follicular cell neoplasms or AUS/FLUS.”\(^3\)
  
  o “Molecular diagnostic testing may include multigene assays (e.g. the gene expression classifier) or individual mutational analysis. The gene expression classifier measures the expression of at least 140 genes.”\(^3\)
o “BRAF V600E mutation analysis was recommended by some panelists for the evaluation of thyroid nodules (not restricted to the follicular lesions).”³

• The current evidence base of the ThyroSeq test consists of three clinical validity studies that evaluate diagnostic performance of the ThyroSeq test to detect thyroid cancer.
  
o In one study, the performance of the ThyroSeq test was reported as 70% (range, 46-88) sensitivity; 77% (range, 66-85), specificity 91% negative predictive value (NPV), and 42% positive predictive value (PPV). Diagnostic performance was showed to be significantly better in the Bethesda category III nodules compared to other categories (area under the curve [AUC] 0.84 vs 0.57; P=0.03).⁴
  
o In a retrospective clinical validity study, sensitivity was reported as 85%, specificity as 65%, PPV as 50%, and NPV as 91%.⁵
  
o A third retrospective, single-institution study reported diagnostic values as follows: sensitivity and specificity (Bethesda IV specimens) were 63 and 86%, resulting in an NPV and a PPV of 75% and 77%, respectively.⁶

• There are currently no available clinical utility studies that evaluated the effects on patient-relevant outcomes (survival, quality of life). Additional research is necessary to assess how testing will be used in the disease management of patients with thyroid cancer.

Criteria

• This test is considered investigational and/or experimental.
  
o Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
  
o In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

References

1. UPMC University of Pittsburgh Medical Center. ThyroSeq® - Thyroid Cancer Next-Generation Sequencing Panel.  
   http://mgp.upmc.com/Applications/mgp/Home/Test/ThyroSeq_Details


Tissue of Origin Testing for Cancer of Unknown Primary

Procedures addressed

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What is cancer of unknown primary testing

Definition

In order to determine the most effective treatment regimen for a patient with cancer it is important to identify the cancer cell type.¹

- When a cancer is found in one or more metastatic sites but the primary site is not known, it is called a cancer of unknown primary (CUP) or an occult primary cancer.² This happens in a small portion of cancers.
- The most commonly used techniques to identify tissue of origin (TOO) for CUP include light microscopy, immunohistochemistry (IHC) staining and computed tomography (CT) or positron emission tomography (PET) imaging.¹
- With advances in technology, some laboratory tests utilize gene expression profiling or other molecular techniques in cancer cells. Ramaswamy et al. found that a cancer-intrinsic gene expression pattern distinguished primary from metastatic adenocarcinomas.³ By comparing the pattern of gene expression in the CUP sample to the patterns seen with other known types of cancer, a CUP may be identified as belonging to a particular cancer type.

Test information

- A number of different companies and approaches are being utilized to diagnose metastatic neoplasms for patients with CUP. These include but are not limited to:
- Tissue of Origin Test from Cancer Genetics Incorporated - uses microarray analysis to measure the expression of over two thousand genes.\(^4\)
- CancerType ID from Biotheranostics analyzes the expression of 92 genes and requires only 300 cells.\(^5\)
- Cancer Origin Test from Rosetta Genomics - uses a RT-PCR platform to analyze the expression levels of 64 microRNAs (miRNAs).\(^6\)

**Guidelines and evidence**

- Under 2016 NCCN guidelines for CUP (occult primary), gene signature profiling for tissue of origin is not recommended for standard management at this time. The panel states that “there may be diagnostic benefit, though not necessarily clinical benefit” and characterizes the use of gene signature profiling for CUP as a category 3 recommendation. The panel also states that “until more robust outcomes and comparative effectiveness data are available, pathologists and oncologists must collaborate on the judicious use of these modalities on a case by case basis.”\(^7\)
- In a systematic review of cancer of unknown primary site in Lancet, gene-profiling diagnosis was noted to have high sensitivity, but additional prospective studies were deemed necessary to establish whether patients’ outcomes are improved by its clinical use.\(^8\)

**Criteria**

This test is considered investigational and/or experimental.

- Investigational and experimental (I&E) molecular and genomic (MolGen) tests refer to assays involving chromosomes, DNA, RNA, or gene products that have insufficient data to determine the net health impact, which typically means there is insufficient data to support that a test accurately assesses the outcome of interest (analytical and clinical validity), significantly improves health outcomes (clinical utility), and/or performs better than an existing standard of care medical management option. Such tests are also not generally accepted as standard of care in the evaluation or management of a particular condition.
- In the case of MolGen testing, FDA clearance is not a reliable standard given the number of laboratory developed tests that currently fall outside of FDA oversight and FDA clearance often does not assess clinical utility.

**References**

1. Sreeletha Meleth, N.W., Tammeka Swinson Evans, Linda Lux. Technology Assessment on Genetic Testing or Molecular Pathology Testing of Cancers with Unknown Primary Site to Determine Origin. Technology Assessment Report. February 20, 2013. Available at:


5. CancerTYPE ID. Biotheranostics Website. Available at: https://www.cancertypeid.com/hcp-what-is-ctid


TPMT Testing for Thiopurine Drug Response

Procedures addressed

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What is thiopurine drug toxicity

Definition

These drugs have a relatively narrow therapeutic window and adverse drug reactions are frequent, with estimates ranging from 5% to 40%.

1-2 Drug toxicity can result in myelosuppression or hepatotoxicity, and can be life-threatening.3 People taking thiopurine should have regular complete blood cell count (CBC) monitoring.4

- The thiopurine drugs – azathioprine (AZA), 6-mercaptopurine (6-MP), and 6-thioguanine (6-TG) – are commonly used to treat hematological malignancies, autoimmune conditions, inflammatory bowel disease, and solid organ transplant rejections.1

- These drugs are metabolized by the enzyme TPMT (thiopurine methyltransferase). Genetic variants in the TPMT gene are associated with lower enzyme activity, leading to an increased risk for drug toxicity.3

- TPMT enzyme activity is largely influenced by polymorphisms (changes) in the TPMT gene. About 29 TPMT variants have been identified. TPMT*2, TPMT*3A, TPMT*3C and account for 85-90% of intermediate or low TPMT enzyme activity.5

- About 1 in 300 (0.3%) people have deficient or undetectable TPMT activity, 11% have low (intermediate) activity and 89% have normal activity. Evidence of a fourth
group of ultra-high TPMT activity has recently been found in about 2% of the population.\textsuperscript{4-6}

- The overall distribution of low, intermediate and normal TPMT activity does not appear to vary among Caucasians, Asians or African-Americans. However, the TPMT variants are not equally distributed among ethnic populations. The frequency of the variant alleles for which commercial genetic testing is currently available is highest in Caucasians and African-Americans. These variants are less common in Southeast (Indonesian, Thai, Filipino, Taiwanese) and Southwest (Indian, Pakistani) Asians.\textsuperscript{4,7}

- TPMT activity can account for up to 75\% of the cases of neutropenia associated with thiopurines. People with absent TPMT activity treated with normal doses of thiopurines are at approximately 100\% risk of developing severe or fatal myelosuppression.\textsuperscript{7} People with low TPMT activity have a 30-40\% risk of developing adverse reactions to thiopurines when treated with standard doses.\textsuperscript{7}

Test information

- **Phenotyping** quantifies TPMT enzyme activity. Testing laboratories generally interpret results as normal, intermediate, or low. Some also report a high enzyme activity level. Phenotyping will detect any lowered enzyme activity, regardless of the specific underlying genetic variation. However, phenotyping results may not be accurate for:
  - People who have received recent blood transfusions (within the last four months).\textsuperscript{4}
  - People currently treated with thiopurine drugs.\textsuperscript{4}
  - People currently taking drugs that inhibit TPMT, including: naproxen, ibuprofen, ketoprofen, furosemide, sulfasalazine, mesalamine, olsalazine, mefenamic acid, thiazide diuretics, and benzoic acid inhibitors. Patients should abstain from these drugs for at least 48 hours prior to blood collection.\textsuperscript{2}

- **Genotyping** for TPMT sensitivity is done by targeted analysis for the most common variant alleles. TPMT*1 is the normal (wild-type) allele; the TPMT*2, *3A, *3B, and *3C alleles are variants common in the general population. Genetic test results are not affected by medication use or blood transfusion.

- Although FDA labeling for thiopurine drugs does not specify a testing method, phenotyping (for enzyme activity) is more common and preferred over genotyping (identifying specific variants), in the absence of a contraindication.\textsuperscript{6}

Guidelines and evidence

- The US Food and Drug Administration (2004) revised the labeling for azathioprine, 6-mecaptopurine and 6-thioguanine:
Azathioprine: “It is recommended that consideration be given to either genotype or phenotype patients for TPMT. Phenotyping and genotyping methods are commercially available. The most common non-functional alleles associated with reduced levels of TPMT activity are TPMT*2, TPMT*3A and TPMT*3C. Patients with two non-functional alleles (homozygous) have low or absent TPMT activity and those with one non-functional allele (heterozygous) have intermediate activity. Accurate phenotyping (red blood cell TPMT activity) results are not possible in patients who have received recent blood transfusions. TPMT testing may also be considered in patients with abnormal CBC results that do not respond to dose reduction. Early drug discontinuation in these patients is advisable.”

6-mecaptopurine (6-MP): “Homozygous-deficient patients (two non-functional alleles), if given usual doses of Mercaptopurine, accumulate excessive cellular concentrations of active thioguanine nucleotides predisposing them to Mercaptopurine toxicity. Heterozygous patients with low or intermediate TPMT activity accumulate higher concentrations of active thioguanine nucleotides than people with normal TPMT activity and are more likely to experience Mercaptopurine toxicity. TPMT genotyping or phenotyping (red blood cell TPMT activity) can identify patients who are homozygous deficient or have low or intermediate TPMT activity.”

6 thioguanine (6-TG): “There are individuals with an inherited deficiency of the enzyme thiopurine methyltransferase (TPMT) who may be unusually sensitive to the myelosuppressive effects of Thioguanine and prone to developing rapid bone marrow suppression following the initiation of treatment. Substantial dosage reductions may be required to avoid the development of life-threatening bone marrow suppression in these patients. Prescribers should be aware that some laboratories offer testing for TPMT deficiency.”


Ideally, TPMT activity testing should occur prior to initiating treatment with thiopurines, so that alternative treatment strategies can be considered in those at higher risk for toxicity.

Thiopurine use in patients with deficient TPMT activity is contraindicated.

Patients with intermediate TPMT activity should be treated with a reduced dose. Some guidelines have suggested a reduction of 50-67%.

TPMT testing may also be considered in patients with abnormal blood cell counts or when clinical evidence of severe toxicity does not respond to dose reduction.
The TARGET trial (TPMT: Azathioprine Response to Genotyping and Enzyme Testing) was a randomized controlled trial evaluating TPMT genotyping prior to treatment with azathioprine. Results from this trial indicated that individuals with homozygous TPMT variants were at risk for severe neutropenia whereas heterozygotes were not at increased risk when taking standard doses of azathioprine.  

Criteria
TPMT testing by phenotyping or genotyping is indicated in individuals considering treatment with any thiopurine drug:

- azathioprine (AZA, Imuran®, Azasan®)
- 6-mercaptopurine (6-MP, Mercaptopurinum®, Purinethol®)
- thioguanine (6-TG, Tabloid®, Thioguanine®)

References


Tumor Marker Testing-Solid Tumors

Procedures addressed

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What are molecular tumor markers

Definition

Molecular tumor markers are broadly defined here as any change in DNA, RNA, or chromosomes found in tumor tissue that is used to make cancer management decisions.

- Tumor markers refer to substances in the body that are altered in the presence of cancer. The alterations may be used to diagnose and subtype cancer, predict prognosis, make therapeutic decisions, and monitor disease progression. Tumor markers may be detected as abnormal protein levels, gene expression (RNA) patterns, chromosome abnormalities, or DNA changes.¹
Molecular tumor markers are increasingly useful for therapy selection. Many cancer therapies are targeted at particular gene functions (therapeutic targets) and some require information about tumor genetics to use the therapies effectively (companion diagnostics). In these cases, NCCN as well as the FDA have outlined tumor testing that is recommended for specific cancers and the associated treatment implications.\textsuperscript{2-5}

**Test information**

The specific methodology used to identify molecular tumor markers is dependent upon the type of tumor marker being investigated.

- DNA mutations are generally detected through direct analysis of hotspots or sequencing either parts of the gene or the whole gene.
- Chromosome abnormalities, such as translocations or deletions, may be detected through direct visualization of the chromosomes (karyotyping), in situ hybridization of probes (e.g., FISH) to detect deletions or duplications that are too small to see directly, or DNA-based methods that identify deletions or translocation breakpoints.
- Gene expression profiling simultaneously measures the amount of RNA or protein being made by many genes. Expression patterns may be used to predict the type of cancer present, tumor aggressiveness, and therapy needs.
- Whole exome sequencing is used to identify multiple molecular tumor markers, at the same time, which may be present in a tumor.

The efficiency of next generation sequencing (NGS) has led to an increasing number of large, multi-gene molecular tumor marker panels. Given that tumors can have multiple and unexpected genetic changes, these panels may provide physicians with information about therapeutic targets that would not otherwise be considered. The following are examples of molecular tumor marker panels (not intended to be a complete list):

- Foundation Medicine: FoundationOne tumor marker panel
- Caris Life Sciences: Caris Tumor profiling
- Paradigm: Paradigm Cancer Diagnostic (PCDx) Panel
- GenPath Diagnostics: OnkoMatch
- University of Washington: UW-OncoPlex-Cancer Gene Panel

**Guidelines and evidence**

The National Comprehensive Cancer Network (NCCN) provides the following types of guidance.

- NCCN Guidelines for Treatment of Cancer by Site provide detailed guidelines on the use of individual tumor markers for each cancer type addressed.\textsuperscript{5}
NCCN also makes the following recommendations specifically for using multi-gene panels in the evaluation of non-small cell lung cancer (NSCLC): “The NCCN NSCLC Guidelines Panel strongly endorses broader molecular profiling with the goal of identifying rare driver mutations for which effective drugs may already be available, or to appropriately counsel patients regarding the availability of clinical trials. Broad molecular profiling is a key component of the improvement of care of patients with NSCLC.”

NCCN has not recommended a panel approach for any other cancer types as of mid-2017.

NCCN also maintains a biomarker compendium stating “the goal of the NCCN Biomarkers Compendium is to provide essential details for those tests which have been approved by NCCN Guideline Panels and are recommended by the NCCN Guidelines.” Biomarkers for specific cancer types that are listed in the NCCN Biomarker Compendium have a level of evidence associated with their clinical utility.

• The National Academy of Clinical Biochemistry (NACB, 2009) issued general tumor marker quality practice guidelines “to encourage more appropriate use of tumor marker tests.” They provide the following guidelines to determine if a tumor marker is useful:
  o “The marker results are appropriate precisely for the required application (i.e., risk assessment, screening, diagnosis, prognosis, prediction, or post-treatment monitoring).”
  o “The marker results separate patients into two or more populations whose outcomes differ so strikingly that they and their caregiver would treat one group differently than another.”
  o “The estimate of the separation in outcomes for marker positive and negative is reliable.”

• Some FDA labels require results from tumor marker tests to effectively or safely use the therapy for a specific cancer type. A list of all Pharmacogenomic Biomarkers included in FDA labeling and associated implications can be found here.

Criteria

This guideline applies to all molecular tumor marker testing intended for use in solid tumors. This guideline does not apply to tumor marker testing for hematologic malignancies. This guideline also does not apply to tumor markers found by liquid biopsy. Please see Liquid Biopsy Testing – Solid Tumors. This guideline also does not apply when testing for germline (inherited) mutations in genes related to hereditary cancer syndromes (e.g., Hereditary Breast and Ovarian Cancer, Lynch syndrome, etc.). Although some of the same genes may be tested for inherited or acquired mutations, this guideline addresses only testing for acquired mutations from tumor tissue.
Medical necessity criteria differ based on the type of testing being performed (i.e., individual tumor markers separately chosen based on the cancer type versus pre-defined panels of tumor markers) and how that testing will be billed (one or more individual tumor marker-specific procedure codes, specific panel procedure codes, or unlisted procedure codes).

When separate procedure codes will be billed for individual tumor markers (e.g., Tier 1 MoPath codes 81200-81355 or Tier 2 MoPath codes 81400-81408), each individually billed tumor marker test will be evaluated separately. The following criteria will be applied:

- The member has a tumor type that will benefit from information provided by the requested tumor marker test based on at least one of the following:
  - All criteria are met from a test-specific guideline if one is available (See Common cancer types and associated tumor markers table below for tumor marker tests that have separate test-specific guidelines.), or
  - An oncology therapy FDA label requires results from the tumor marker test to effectively or safely use the therapy for the member’s cancer type (See Common cancer types and associated tumor markers table below for examples of currently recognized companion diagnostics), or
  - NCCN guidelines include the tumor marker test in the management algorithm for that particular cancer type and all other requirements are met (specific pathology findings, staging, etc.); however, the tumor marker must be explicitly included in the guidelines and not simply included in a footnote as an intervention that may be considered, or
  - The NCCN Biomarker Compendium has a level of evidence of at least 2A for the tumor marker’s application to the member’s specific cancer type

Note If five or more individually billed tumor marker tests are under review together (a “panel”) and the member either has non-small cell lung cancer OR meets criteria for 5 or more individual tumor markers, the panel will be approved. However, the laboratory will be redirected to use a panel CPT code for billing purposes (e.g. 81445 or 81455).

When a multi-gene panel is being requested and will be billed with a single panel CPT code (e.g. 81445 or 81455), the panel will be considered medically necessary when the following criteria are met:

- The member has a diagnosis of non-small cell lung cancer, OR
- At least 5 tumor markers included in the panel individually meet criteria for the member’s tumor type based on one of the following:
  - All criteria are met from a test-specific guideline if one is available (See Common cancer types and associated tumor markers table below for tumor marker tests that have separate test-specific guidelines.), or
An oncology therapy FDA label requires results from the tumor marker test to effectively or safely use the therapy for the member’s cancer type (See Common cancer types and associated tumor markers table below for examples of currently recognized companion diagnostics for available therapies.), or

NCCN guidelines include the tumor marker test in the management algorithm for that particular cancer type and all other requirements are met (specific pathology findings, staging, etc.); however, the tumor marker must be explicitly included in the guidelines and not simply included in a footnote as an intervention that “may be considered”, or

The NCCN Biomarker Compendium has a level of evidence of at least 2A for the tumor marker’s application to the member’s specific cancer type.

**Note** If the member meets criteria for less than 5 of the individual tumor markers in the panel, the panel will not be reimbursed. The laboratory will be redirected to billing for individual tests for which the member meets criteria.

### Common cancer types and associated tumor markers

This list not all inclusive.

<table>
<thead>
<tr>
<th>Cancer Type</th>
<th>Tumor Marker</th>
<th>CPT</th>
<th>Claim Code</th>
<th>Associated Treatments*</th>
<th>Applicable Guideline</th>
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</thead>
<tbody>
<tr>
<td>Colorectal7^ (Metastatic, stage IV. Prognostic purposes only.)</td>
<td>BRAF</td>
<td>81210</td>
<td>N/A</td>
<td>N/A</td>
<td>MOL.TS.133</td>
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<td>81275</td>
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<td>81210</td>
<td>N/A</td>
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<tr>
<td>Non-small</td>
<td>EGFR</td>
<td>81235</td>
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<td>erlotinib^2,</td>
<td>MOL.TS.163</td>
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<tr>
<td>Cancer Type</td>
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<td>CPT</td>
<td>Claim Code</td>
<td>Associated Treatments*</td>
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<td></td>
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<td>EML4ALK</td>
<td>crizotinib², ceritinib², alectinib</td>
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<td>ALK</td>
<td>crizotinib², ceritinib², alectinib</td>
<td>MOL.TS.230</td>
</tr>
</tbody>
</table>

**Note**: In general, when there is an associated treatment, results from the referenced tumor marker are necessary for the safe or effective use of that therapy (companion diagnostics). The therapies and tumor markers are only included for cancer types approved for treatment according to FDA labeling.

**References**


3. US Food and Drug Administration. Table of Pharmacogenomic Biomarkers in Drug Labeling. Available at: [http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm](http://www.fda.gov/Drugs/ScienceResearch/ResearchAreas/Pharmacogenetics/ucm083378.htm)


UGT1A1 Mutation Analysis for Irinotecan Response

Procedures addressed

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<table>
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<tr>
<th>Procedure addressed by this guideline</th>
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</tr>
</thead>
<tbody>
<tr>
<td>UGT1A1 Targeted Mutation Analysis</td>
<td>81350</td>
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</table>

What are UGT1A1 and irinotecan

Definition

Irinotecan is a chemotherapy drug often prescribed together with other standard agents for treating patients with metastatic and recurrent colorectal cancer. Irinotecan is metabolized by a gene called UGT1A1 in the liver.1

• A common change, or variant, in the UGT1A1 gene called UGT1A1*28 can lead to reduced enzyme activity. This can cause a buildup of drug metabolites, resulting in toxicity.1,2

• Several studies have confirmed an increased risk of having reduced white blood cell count, or neutropenia, in people with UGT1A1 genetic variants. Some studies, but not all, have shown an increased risk of severe diarrhea.3

• About 10% of North Americans have two copies of the UGT1A1*28 mutation (homozygous, also referred to as UGT1A1 7/7) and 40% have just one copy (heterozygous).2

• Not all people with UGT1A1*28 mutations will experience increased toxicity.3 People homozygous for the *28 mutation are 3.5 times more likely to develop severe neutropenia than those with the wild genotype.1

Test information

• Targeted mutation analysis of the UGT1A1 gene sequence by polymerase chain reaction (PCR) identifies any mutation in the region. The results are reported as negative, heterozygous or homozygous.1,2,4

  o **Negative** = UGT1A1 6/6 (*1/*1) genotype; Wild-type genotype; No UGT1A1*28 mutation is identified. Low risk of severe toxicity from standard initial dosages of irinotecan.
o **Heterozygous** = UGT1A1 6/7 (*1/*28) genotype; One wild-type allele and one UGT1A1*28 mutation allele identified. Increased risk for irinotecan toxicity, but initial standard doses may be still be tolerated.

o **Homozygous** = UGT1A1 7/7 (*28/*28) genotype. Increased risk for severe toxicity from standard initial doses of irinotecan, thus irinotecan product labeling recommends considering a reduced initial dose.

**Guidelines and evidence**

- In May 2010, the FDA announced a safety change to the prescribing information for Camptosar® (irinotecan) Injection:
  - “When administered in combination with other agents, or as a single-agent, a reduction in the starting dose by at least one level of Camptosar® should be considered for patients known to be homozygous for the UGT1A1*28 allele. However, the precise dose reduction in this patient population is not known and subsequent dose modifications should be considered based on individual patient tolerance to treatment.”
  - “A laboratory test is available to determine the UGT1A1 status of patients. Testing can detect UGT1A1 6/6, 6/7, 7/7 genotypes.”

- UGT1A1 *28 testing for irinotecan is recognized by the FDA as a valid genomic biomarker.  
  
- Guidelines for genetic testing have not been established by organizations such as the National Comprehensive Cancer Network (NCCN) and the Evaluation of Genomic Applications in Practice and Prevention (EGAPP) Working Group. However, both organizations recognize the availability and utility of testing UGT1A1 *28 prior to treatment with irinotecan. 

- The NCCN states the following:
  - “Also, a warning was added to the label for irinotecan indicating that a reduced starting dose of the drug should be used in patients known to be homozygous for UGT1A1*28.”
  - “A practical approach to the use of UGT1A1*28 allele testing with respect to patients receiving irinotecan has been presented, although guidelines for the use of this test in clinical practice have not been established.”
o “UGT1A1 testing on patients who experience irinotecan toxicity is not recommended, because they will require a dose reduction regardless of the UGT1A1 test result.”

Criteria
UGT1A1 testing is indicated in individuals with metastatic and/or recurrent colorectal cancer prior to the initiation of irinotecan therapy.

References
UroVysion FISH for Bladder Cancer

Procedures addressed

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<table>
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<tr>
<td>FISH Analysis for Bladder Cancer (UroVysion), Manual</td>
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<tr>
<td>FISH Analysis for Bladder Cancer (UroVysion), Computer-Assisted</td>
<td>88121</td>
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</table>

What is UroVysion FISH testing for bladder cancer

Definition

UroVysion™ was developed to be used with current standard diagnostic tools to aid in initial diagnosis of bladder cancer and monitoring for tumor recurrence in previously diagnosed patients.4

- Bladder cancer is one of the most common types of cancer in the U.S., especially among men. Approximately 76,960 new cases of bladder cancer are projected for 2016 (58,950 in men and 18,010 in women).1 Older individuals (median age, 65 years) are most often affected.
- Bladder cancer is categorized as non-muscle invasive disease (NMID) or muscle invasive disease (MID).2 The majority (≈80%) of bladder cancers are NMID.3
- Urothelial carcinoma (UC) accounts for most cases of bladder cancer.2,4
  - Most cases of UC are low-grade and easily treated.4
  - However, UC has a high risk of recurrence (70%), and patients must be monitored for several years after treatment.4
- Diagnostic monitoring usually consists of regular testing of cells in the urine (cytology).2,5 UroVysion FISH (fluorescence in situ hybridization) testing is an alternative to cytology.4,5

Test information

- UroVysion testing detects extra or missing chromosomes 3, 7 or 17 and gene changes to a piece of chromosome 9 often found in UC patients.4
• Cytology is the standard procedure for diagnosing and monitoring of UC. UroVysion testing can be performed if the cytology returns negative or atypical results.\textsuperscript{2,4,5}

• One study showed UroVysion testing to have 85% sensitivity for low-grade UC, and nearly 100% sensitivity for the more rare but serious high-grade UC.\textsuperscript{4}

**Guidelines and evidence**

• UroVysion testing is FDA approved,\textsuperscript{6} but reviews and guidelines call for additional study before it becomes standard procedure.\textsuperscript{5}

• The National Cancer Center Network (NCCN, 2016)\textsuperscript{2} has published clinical guidelines on bladder cancer, and state that urinary urothelial tumor markers are classified as category 2B, and an option for follow-up assessment in patients with low to high grade disease who have undergone adjuvant intravesical treatment. They report that urothelial tumor markers have better sensitivity, but lower specificity, for detecting bladder cancer compared with urinary cytology. It is not known whether urinary marker tests add useful clinical information for the detection and management of non-muscle invasive bladder tumors.

• The American Urological Association and the Society of Urologic Oncology (AUA, 2016)\textsuperscript{7} recently published clinical practice guidelines regarding microscopic hematuria and the management of non-muscle invasive bladder cancer (NMIB). For urinary markers utilized after diagnosis of bladder cancer, they state the following:
  - “In surveillance of NMIC, a clinician should not use urinary biomarkers in place of cystoscopic evaluation (Strong Recommendation; Evidence Strength: Grade B)”
  - “In a patient with a history of low-risk cancer and a normal cystoscopy, a clinician should not routinely use a urinary biomarker or cytology during surveillance. (Expert Opinion)”
  - “In a patient with NMIBC, a clinician may use biomarkers to assess response to intravesical BCG (UroVysion FISH) and adjudicate equivocal cytology (UroVysion FISH and ImmunoCyt). (Expert Opinion)”

• The National Institute for Health and Care Excellence (NICE, 2015)\textsuperscript{8} published a guideline regarding the diagnosis and management of bladder cancer. They stated that urinary biomarker tests (such as UroVysion using FISH, ImmunoCyt or a nuclear matrix protein 22 (NMP22) test may be used for the diagnosis of individuals with suspected bladder cancer.

• The American Urological Association (AUA, 2012)\textsuperscript{9} stated the following regarding the management of asymptomatic microhematuria:
  - “The use of urine cytology and urine markers (NMP22, BTA-stat, and UroVysion FISH) is NOT recommended as a part of routine evaluation of the asymptomatic microhematuria patient. (Recommendation: Evidence Strength C).”
• American Urological Association (2007) guidelines for diagnosis and management of bladder cancer consider techniques like UroVysion to “hold promise” in future assessment of risk, prognosis, and targeted treatment.⁵

• A systematic review of UroVysion was conducted by the Agency for Healthcare Research and Quality (AHRQ).¹⁰ Based on 11 studies that were reviewed, the following were noted by authors:
  
  o Diagnostic testing:
    - The sensitivity of Urovysion to detect bladder cancer among undiagnosed patients with clinical signs and symptoms was 63% (95% CI, 50% to 75%) and specificity was 87% (95% CI, 79% to 93%).
    - The positive likelihood ratio was 5.02 (95% CI 2.93 to 8.60) (moderate increase in the likelihood of disease). The negative likelihood ratio was 0.42 (95% CI 0.30 to 0.59) (small decrease in likelihood of disease).

  o Surveillance testing:
    - For individuals being monitored for cancer recurrence the sensitivity was 55% (95% CI, 36% to 72%; 7 studies) and specificity was 80% (95% CI, 66% to 89%; 6 studies).
    - For evaluation of symptoms, sensitivity was 73% (95% CI, 50% to 88%), based on two studies.

  o The sensitivity of the test increased with higher tumor stage and grade.

• A prior systematic review and meta-analysis provided a comparison of Urovysion with cystoscopy for diagnosing UC.¹¹
  
  o Pooled sensitivity of UroVysion was 72% (95% CI, 69% to 75%) compared with 42% (95% CI, 38% to 45%) for cytology.
  
  o Specificity of UroVysion was 83% (95% CI, 82% to 85%) and 96% (95% CI, 95% to 97%) for cytology.
  
  o Sensitivity increased if earlier stage tumors (Ta) were excluded: Urovysion, 86% (95% CI, 82% to 89%) and cytology, 61% (95% CI, 56% to 66%).
  
  o The overall diagnostic performance was higher for UroVysion (area under the curve [AUC], 0.867, standard error [SE], 0.021) versus cytology (AUC, 0.626 [SE 0.091]). However, these differences in diagnostic accuracy disappeared when Ta cases were excluded, indicating that the tests may be more comparable for accurately detecting later stage disease.

• A prospective, blinded study compared results of ImmunoCyt and UroVysion with urine cytology to detect recurrent UC.¹² A total of 100 patients with a history of bladder cancer submitted a single voided urine sample, immediately followed by cystoscopy and biopsy.
• The overall results of sensitivity for low- and high-grade UC for the various tests were as follows: Cytology, 15% and 27%; Immunocyt, 62% and 91%; and UroVysion, 8% and 18%.

• Specificity of the tests were: Cytology, 97%; Immunocyt, 63%; UroVysion, 90%.

• Positive predictive value (PPV) and negative predictive value (NPV) of each test demonstrated: Cytology 71% and 78%; Immunocyt 43% and 88%; UroVysion 33% and 72%.

• Immunocyt had significantly better sensitivity at detecting low-grade tumors compare with cytology or UroVysion (76% vs 21% and 13%, respectively; P<0.001 for both). However, specificity for Immunocyt was significantly lower than for cytology and UroVysion (63% vs 97% and 90%; P<0.001 for both).

• The authors concluded that UroVysion and cytology had comparably high rates of specificity for patients with negative cystoscopy, and as a result, UroVysion may have value to confirm cytology or Immunocyt.

• A retrospective study evaluating the ability of UroVysion to predict recurrence and progression in NMIBC with suspicious cytology and negative cystoscopy results was undertaken due to the high rate of recurrence and frequent cystoscopic monitoring required in this disease. Authors evaluated predictors for recurrence, progression and findings on subsequent cystoscopy.\(^{13}\)

  o A positive UroVysion result was a significant predictor for recurrence (hazard ratio [HR], 2.35; 95% CI, 1.42 to 3.90; \(P=0.001\)) and for progression (HR, 3.01; 95% CI, 1.10 to 8.21; \(P=0.03\)).

  o A positive UroVysion result, however, was not significantly associated with evidence of tumor on subsequent surveillance cystoscopy compared with negative UroVysion results (odds ratio [OR], 0.80; 95% CI, 0.26 to 2.74; NS).

  o The authors concluded that an “anticipatory positive” UroVysion result can predict recurrence and progression in patients with NMIBC when cystoscopy results are negative and cytology results are suspicious.

  o The lack of association between UroVysion results and tumor recurrence on cystoscopy during the immediate follow-up period, however, suggests that UroVysion has limited value to modify surveillance regimens in these patients.

**Criteria**

• Previous Testing:
  o No repeat UroVysion\(^\circ\) testing on the same sample when a result was successfully obtained, AND

• Diagnosis
- UroVysion is not indicated for the routine evaluation of hematuria or microhematuria and will not be reimbursed when billed with an ICD10 code in the R31 Hematuria range. Exceptions may be made for uncertain or equivocal results on standard diagnostic assessments, such as cytology, OR

- Surveillance
  - UroVysion is indicated when the individual has a personal history of bladder cancer defined as an ICD10 code in the C67 Malignant neoplasm of the bladder range, AND
  - The member is being monitored for cancer recurrence, AND
    - Member had been diagnosed with low grade bladder cancer and the results of cytology are equivocal, or
    - Member had been diagnosed with high grade bladder cancer and the results of cytology are negative or equivocal, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

References


Procedures addressed

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<thead>
<tr>
<th>Procedures addressed by this guideline</th>
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<td>VHL Deletion/Duplication Analysis</td>
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What is Von Hippel-Lindau (VHL) syndrome

Definition

Von Hippel-Lindau (VHL) syndrome is a hereditary cancer syndrome.

- Von Hippel-Lindau (VHL) syndrome is a hereditary cancer syndrome whose main clinical features include hemangioblastomas of the central nervous system (CNS) and retina, renal cysts and renal cell carcinoma, pheochromocytoma, and endolymphatic sac tumors.\(^1\)
  - The cardinal feature of VHL syndrome is hemangioblastoma. CNS hemangioblastomas present in 60%-80% of individuals, and retinal hemangioblastomas present in about 70-80% of individuals.\(^1,2\)
  - The risk to develop clear cell renal carcinoma by age 60 is as high as 70%, and is the leading cause of death for individuals with VHL syndrome.\(^1,2\)
  - Pheochromocytomas and endolymphatic sac tumors are less commonly seen in VHL syndrome than other manifestations.
  - Epididymal tumors have also been reported in VHL. Males with bilateral epididymal tumors may have infertility.\(^1\)
  - Clinical findings of VHL may include vision loss, hearing loss, gait disturbance, pain and sensory motor loss depending on the location of the tumor.\(^1\)
- The incidence of VHL is 1 in 36,000 people.\(^1\)
- VHL syndrome is caused by mutations in the VHL gene. More than 1500 germline and sporadic VHL gene mutations have been identified. The VHL gene is a tumor suppressor whose normal role is to control cell growth and proliferation.\(^1\)
mutations lead to a loss of function of the gene and an increased risk for uncontrolled growth of tumors and cysts.\textsuperscript{1}

- Most (80\%) of VHL mutations are inherited (germline), and about 20\% are new (de novo) mutations.\textsuperscript{1} VHL syndrome is an autosomal dominant condition with children of affected individuals having a 50\% chance of inheriting the disease-causing mutation.\textsuperscript{1}

- Almost 100\% of individuals with a VHL gene mutation show symptoms of the disease by age 65.\textsuperscript{1} Age of onset, disease severity, and tumor types vary between and within affected families.

- Surveillance recommendations for individuals diagnosed with or at-risk for inheriting VHL syndrome include annual ophthalmologic exams, MRI of the brain and total spine every two years starting at age 16 years, annual abdominal ultrasound, MRI of the abdomen every two years starting at 16 years, annual blood pressure monitoring, annual blood or urinary fractionated metanephrines starting at 5 years, and audiologic evaluation. Some of the screenings should begin at one year of age in at-risk/affected individuals.\textsuperscript{3,4} Early detection of VHL tumors may lead to improved outcome.\textsuperscript{1} However, at-risk individuals can forego screening if genetic testing for a known familial mutation is performed and they have a normal (negative) result.\textsuperscript{1,2}

### Test information

- **VHL full gene sequence analysis** checks all three exons and will detect about 72\% of mutations.\textsuperscript{1,3} Some laboratories perform only sequencing, while others do sequencing with reflex to deletion/duplication analysis or perform sequencing and deletion/duplication analysis concurrently.

- **VHL deletion/duplication analysis** detects partial or complete gene deletions which account for about 28\% of VHL mutations.\textsuperscript{1,3}

- **VHL known familial mutation analysis:** Once a VHL mutation is identified in an affected person, predictive testing is available for at-risk family members, as is prenatal or preimplantation genetic diagnosis.\textsuperscript{1} Family members should be tested using the method that can accurately identify the familial mutation. This testing is typically less expensive than a full gene evaluation and provides clear results about whether the family member is predisposed to developing VHL syndrome.\textsuperscript{1}

### Guidelines and evidence

- Consensus-based clinical diagnostic guidelines state that the diagnosis of VHL can be made in the following circumstances:\textsuperscript{5}
  
  - “Patients with a family history, and a CNS haemangioblastoma (including retinal haemangioblastomas), phaeochromocytoma, or clear cell renal carcinoma are diagnosed with the disease.”
“Those with no relevant family history must have two or more CNS haemangioblastomas, or one CNS haemangioblastoma and a visceral tumour (with the exception of epididymal and renal cysts, which are frequent in the general population) to meet the diagnostic criteria.”

- A 2012 expert-authored review recommends the following testing strategy to confirm/establish the diagnosis in an affected individual:¹
  - “Genetic testing is indicated in all individuals known to have or suspected of having VHL syndrome.”
  - “For individuals with manifestations of VHL syndrome who do not meet strict diagnostic criteria and who do not have a detectable VHL germline mutation, somatic mosaicism for a de novo VHL disease-causing mutation should be considered. In some instances, genetic testing of the offspring of such individuals reveals a VHL mutation.”
  - The high sensitivity of the molecular test for VHL makes confirming a diagnosis relatively straightforward in individuals who may have features of VHL but may not meet diagnostic criteria.¹

- A 2012 expert-authored review states: “Use of molecular genetic testing for early identification of at-risk family members improves diagnostic certainty and reduces the need for screening procedures in those at-risk family members who have not inherited the disease-causing mutation.”¹

- The American Society of Clinical Oncologists (ASCO) position statement on genetic testing (originally published 1996⁶; revised/affirmed in 2003⁷, 2010⁸, and 2015⁹) considers VHL syndrome a Group 1 disorder: “Tests for families with well defined hereditary syndromes for either a positive or negative result will change medical or prenatal management, and for whom genetic testing may be utilized as part of the routine medical care.”

  - The 2003 update specifically addresses issues around genetic testing in affected and at-risk children:
    - “ASCO recommends that the decision to offer testing to potentially affected children should take into account the availability of evidence-based risk-reduction strategies and the probability of developing a malignancy during childhood. Where risk-reduction strategies are available or cancer predominantly develops in childhood, ASCO believes that the scope of parental authority encompasses the right to decide for or against testing.”

Criteria

VHL Known Familial Mutation Analysis

- Genetic Counseling:
o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  o No previous VHL gene testing that would have detected the family mutation, AND

- Diagnostic and Predisposition Testing:**
  o Known family mutation in VHL identified in 1st degree relative(s). (Note: 2nd or 3rd degree relatives may be considered when 1st degree relatives are unavailable or unwilling to be tested), AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

** Includes prenatal testing for at-risk pregnancies.

**VHL Sequencing**

- Genetic Counseling:
  o Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Testing:
  o No previous VHL gene sequencing, and
  o No known familial mutation, AND

- Diagnostic Testing for Symptomatic Individuals:
  o A positive family history of VHL, and
    - Spinal or cerebellar hemangioblastoma, or
    - Retinal hemangioblastoma, or
    - Renal cell carcinoma, or
    - Pheochromocytoma, or
    - Multiple renal and/or pancreatic cysts, OR
  
  o No known family history of VHL-related findings, and
    - Two or more hemangioblastomas involving the retina, spine, and/or brain, or
    - A single hemangioblastoma and a characteristic visceral mass (such as renal cell carcinoma, pheochromocytoma, endolymphatic sac tumors, papillary cystadenomas of the epididymis or broad ligament, or neuroendocrine tumors of the pancreas), OR

- Predisposition Testing for Presymptomatic/Asymptomatic Individuals:
A first-degree relative of someone with a clinical diagnosis of VHL who has had no previous genetic testing (Note that testing in the setting of a more distant affected relative will only be considered if the first-degree relative is unavailable or unwilling to be tested); AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

**VHL Deletion/Duplication Analysis**

- Genetic Counseling:
  - Pre and post-test genetic counseling by an appropriate provider (as deemed by the Health Plan policy), AND

- Previous Genetic Testing:
  - There is no known familial mutation, and
  - No previous deletion/duplication analysis of the VHL gene has been performed, and
  - Above criteria for VHL full gene sequence analysis are met, and
  - VHL sequencing was previously performed and no mutations were found, AND

- Rendering laboratory is a qualified provider of service per the Health Plan policy

**References**


Whole Exome Sequencing

Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Exome (e.g., unexplained constitutional or heritable disorder or syndrome); sequence analysis</td>
<td>81415</td>
</tr>
<tr>
<td>Sequence analysis, each comparator exome (e.g., parent(s), sibling(s))</td>
<td>81416</td>
</tr>
<tr>
<td>Re-evaluation of previously obtained exome sequence (e.g., updated knowledge or unrelated condition/syndrome)</td>
<td>81417</td>
</tr>
</tbody>
</table>

What is whole exome sequencing

Definition

Whole exome sequencing (WES) utilizes DNA-enrichment methods and massively parallel nucleotide sequencing to identify disease-associated variants throughout the human genome.

- WES has been proposed for diagnostic use in individuals who present with complex genetic phenotypes suspected of having a rare genetic condition, who cannot be diagnosed by standard clinical workup, or when features suggest a broad differential diagnosis that would require evaluation by multiple genetic tests.

- The standard approach to the diagnostic evaluation of an individual suspected of having a rare genetic condition may include combinations of radiographic, biochemical, electrophysiologic, and targeted genetic testing such as a chromosomal microarray, single-gene analysis, and/or a targeted gene panel.¹

- WES is typically not an appropriate first-tier test, but can be appropriate if initial testing is unrevealing, or if there is no single-gene or panel test available for the particular condition.²
Identifying a molecularly confirmed diagnosis in a timely manner for an individual with a rare genetic condition can have a variety of health outcomes, including:

- guiding prognosis and improving clinical decision-making, which can improve clinical outcome by
  - application of specific treatments as well as withholding of contraindicated treatments for certain rare genetic conditions
  - surveillance for later-onset comorbidities
  - initiation of palliative care
  - withdrawal of care
- reducing the financial & psychological impact of diagnostic uncertainty and the diagnostic odyssey (e.g., eliminating lower-yield testing and additional screening testing that may later be proven unnecessary once a diagnosis is achieved)
- informing genetic counseling related to recurrence risk and prenatal diagnosis options
- allowing for more rapid molecular diagnosis than a sequential genetic testing approach

Test information

- WES is limited to the DNA sequence of coding regions (exons) and flanking intronic regions of the genome, which is estimated to contain 85% of heritable disease-causing variants.
- Pathogenic variants that can be identified by WES include missense, nonsense, splice-site, and small deletions or insertions.
- At the present time, WES typically fails to detect certain classes of disease-causing variants, such as structural variants (e.g., translocations, inversions), abnormal chromosome imprinting or methylation, copy-number variants, some mid-size insertions and deletions (ca. 10-500 bp), trinucleotide repeat expansion mutations, deeper intronic mutations, and low-level mosaicism.
- WES has the advantage of decreased turnaround time and increased efficiency relative to Sanger sequencing of multiple genes.
• WES is associated with technical and analytical variability, including uneven sequencing coverage, gaps in exon capture before sequencing, as well as variability in variant classification based on proprietary filtering algorithms and potential lack of critical clinical history or family samples.\(^{10}\)

**Guidelines and evidence**

• The American College of Medical Genetics (ACMG, 2012) states the following regarding the clinical application of whole exome and whole genome testing:\(^{11}\)
  
  o “WGS/WES should be considered in the clinical diagnostic assessment of a phenotypically affected individual when:”
    
    ▪ “The phenotype or family history data strongly implicate a genetic etiology, but the phenotype does not correspond with a specific disorder for which a genetic test targeting a specific gene is available on a clinical basis.”
    
    ▪ “A patient presents with a defined genetic disorder that demonstrates a high degree of genetic heterogeneity, making WES or WGS analysis of multiple genes simultaneously a more practical approach.”
    
    ▪ “A patient presents with a likely genetic disorder, but specific genetic tests available for that phenotype have failed to arrive at a diagnosis.”
    
    ▪ “A fetus with a likely genetic disorder in which specific genetic tests, including targeted sequencing tests, available for that phenotype have failed to arrive at a diagnosis.”
    
    ▪ “Prenatal diagnosis by genomic (i.e., next-generation whole-exome or whole-genome) sequencing has significant limitations. The current technology does not support short turnaround times, which are often expected in the prenatal setting. There are high rates of false positives, false negatives, and variants of unknown clinical significance. These can be expected to be significantly higher than seen when array CGH is used in prenatal diagnosis.”

  o The following are recommended pretest considerations:
    
    ▪ “Pretest counseling should be done by a medical geneticist or an affiliated genetic counselor and should include a formal consent process.”
    
    ▪ “Before initiating WGS/WES, participants should be counseled regarding the expected outcomes of testing, the likelihood and type of incidental results that could be generated, and what results will or will not be disclosed.”
    
    ▪ “As part of the pretest counseling, a clear distinction should be made between clinical and research-based testing. In many cases, findings will include variants of unknown significance that might be the subject for research; in such instances a protocol approved by an institutional review board must be in place and appropriate prior informed consent obtained from the participant.”

• The American College of Medical Genetics (ACMG, 2012) states the following regarding informed consent for whole exome and whole genome testing:\(^{12}\)
“Before initiating GS/ES, counseling should be performed by a medical geneticist or an affiliated genetic counselor and should include written documentation of consent from the patient.”

“Incidental/secondary findings revealed in either children or adults may have high clinical significance for which interventions exist to prevent or ameliorate disease severity. Patients should be informed of this possibility as a part of the informed consent process.”

“Pretest counseling should include a discussion of the expected outcomes of testing, the likelihood and type of incidental results that may be generated, and the types of results that will or will not be returned. Patients should know if and what type of incidental findings may be returned to their referring physician by the laboratory performing the test.”

“GS/ES is not recommended before the legal age of majority except for:

- Phenotype-driven clinical diagnostic uses;
- Circumstances in which early monitoring or interventions are available and effective; or
- Institutional review board–approved research.”

“As part of the pretest counseling, a clear distinction should be made between clinical and research-based testing.”

“Patients should be informed as to whether individually identifiable results may be provided to databases, and they should be permitted to opt out of such disclosure.”

“Patients should be informed of policies regarding re-contact of referring physicians as new knowledge is gained about the significance of particular results.”

The American College of Medical Genetics (ACMG, Updated 2016) published guidelines for the reporting of incidental findings in clinical exome and genome sequencing. They state the following:

- “We continue to support the reporting of known or expected pathogenic variants, but we do not recommend reporting variants of uncertain significant as secondary findings (SFs).”
- This guideline includes a table of “ACMG SF v2.0 genes and associated phenotypes recommended for return of secondary findings in clinical sequencing.”

Evidence for the clinical utility of WES in individuals with multiple congenital anomalies and/or a neurodevelopmental phenotype includes numerous large case series. Relevant outcomes include improved clinical decision-making (e.g., application of specific treatments, withholding of contraindicated treatments, changes to surveillance), changes in reproductive decision making, and resource
utilization. WES serves as a powerful diagnostic tool for individuals with rare genetic conditions in which the specific genetic etiology is unclear or unidentified by standard clinical workup.\textsuperscript{7,15,16} 

- The average diagnostic yield of WES is 20-40\% depending on the individual’s age, phenotype, previous workup, and number of comparator samples analyzed.\textsuperscript{5,15,17} Among individuals with a pathogenic or likely pathogenic findings by WES, 5-7\% received a dual molecular diagnosis (i.e., two significant findings associated with non-overlapping clinical presentations).\textsuperscript{15,17} 

- The use of family trio WES reduces the rate of uncertain findings, adds to the clinical sensitivity with regard to the interpretation of clinically novel genes, and increases the diagnostic utility of WES. For example, in three publications the positive rate ranges from 31-37\% in patients undergoing trio analysis compared to 20-23\% positive rate among proband-only WES.\textsuperscript{15,18,19} 

- Re-evaluation of previously obtained exome sequence has the potential for additional diagnostic yield because of constant expansions of existing variant databases, as well as periodic novel gene discovery.\textsuperscript{20}

**Criteria**

- Whole exome sequencing (WES) is considered medically necessary when ALL of the following criteria are met:
  - The patient and family history have been evaluated by a Board-Certified or Board-Eligible Medical Geneticist, AND
    - A clinical letter detailing the evaluation by a Geneticist is provided which includes ALL of the following information:
      - Differential diagnoses, and
      - Testing algorithm, and
      - Previous tests performed and results, and
      - A genetic etiology is the most likely explanation, and
      - Recommendation that whole exome sequencing is the most appropriate test, and
      - Predicted impact on member’s plan of care, AND
  - Patient is <21 years of age, AND
  - A genetic etiology is considered the most likely explanation for the phenotype, based on EITHER of the following:
    - Multiple congenital abnormalities defined by ONE of the following:
      - Two or more major anomalies affecting different organ systems**, or
• One major and two or more minor anomalies affecting different organ systems*, OR

  ▪ TWO of the following criteria are met:
    ▪ major abnormality affecting at minimum a single organ system, and/or
    ▪ formal diagnosis of autism, significant developmental delay, or intellectual disability (e.g., characterized by significant limitations in both intellectual functioning and in adaptive behavior), and/or
    ▪ symptoms of a complex neurodevelopmental disorder (e.g., self-injurious behavior, reverse sleep-wake cycles, dystonia, ataxia, alternating hemiplegia, neuromuscular disorder), and/or
    ▪ severe neuropsychiatric condition (e.g., schizophrenia, bipolar disorder, Tourette syndrome), and/or
    ▪ period of unexplained developmental regression, and/or
    ▪ biochemical findings suggestive of an inborn error of metabolism, AND

  o Alternate etiologies have been considered and ruled out when possible (e.g., environmental exposure, injury, infection), AND
  o Clinical presentation does not fit a well-described syndrome for which first tier testing (e.g., single gene testing, comparative genomic hybridization [CGH]/chromosomal microarray analysis [CMA]) is available, AND
  o Multiple targeted panels are appropriate based on the member’s clinical presentation, AND
  o There is a predicted impact on health outcomes including:
    ▪ Application of specific treatments, or
    ▪ Withholding of contraindicated treatments, or
    ▪ Surveillance for later-onset comorbidities, or
    ▪ Initiation of palliative care, or
    ▪ Withdrawal of care, AND
  o A diagnosis cannot be made by standard clinical work-up, excluding invasive procedures such as muscle biopsy

** Major structural abnormalities are generally serious enough as to require medical treatment on their own (such as surgery) and are not minor developmental variations that may or may not suggest an underlying disorder.

Exclusions and other considerations

• WES is considered experimental/investigational for screening for genetic disorders in asymptomatic or pre-symptomatic individuals.
Whole Genome Sequencing (WGS) is considered investigational/experimental for all indications.

Billing and reimbursement

WES will be considered for reimbursement when it is deemed more efficient and economical than the separate single-gene tests or panels that would be recommended based on the differential diagnosis (e.g., genetic conditions that demonstrate a high degree of genetic heterogeneity).

WES will be considered for reimbursement only when billed with an appropriate CPT code:

- 81415 should be billed for the proband. 81415 should only be billed when analyzing the entire whole exome sequence, rather than a targeted set of genes. At a minimum, genes associated with the clinical presentation and those constitutional mutations in genes listed on the ACMG minimum list entitled “Conditions, genes, and variants recommended for return of incidental findings in clinical sequencing” (Green et al)\(^{13}\) should be reported by the laboratory to the ordering clinician, regardless of the indication for which the exome sequence was ordered.

- 81416 should be billed when a comparator exome is performed. A trio of the proband and both parents is generally preferred, although other family members may be more informative based on the clinical presentation. A maximum of two units of 81416 will be considered for reimbursement.

81415 is not reimbursable for a targeted exome analysis (e.g., XomeDxSlice). The appropriate GSP panel code, unlisted code (e.g. 81479), or Tier 1 or Tier 2 code(s) must be billed.

81415 will be reimbursable once per lifetime.

When a single exome platform is used for more than one test (e.g., XomeDxSlice reflex to full exome analysis), all tests reported from the same exome analysis may be:

- Billed together under one unit of 81415, or
- Billed separately, but 81415 cannot be used. When billed separately, studies may be billed using Tier 1 codes, Tier 2 codes, or 81479 at an amount that does not exceed the cost of full exome analysis.

81417 is not an appropriate code for reflex from targeted to full exome.
• Re-evaluation of a previously obtained exome due to updated knowledge or for the purpose of evaluating a patient for an unrelated condition/syndrome on a different date of service will be considered for reimbursement only when billed using 81417.

References


Administrative Guidelines
# Molecular Pathology Tier 2 Molecular CPT Codes

**MOL.AD.102.A**  
v2.0.2018

## Introduction

Tier 2 Molecular Pathology CPT codes 81400-81408 are addressed by this guideline.

## Procedures addressed

The inclusion of any procedure code in this table does not imply that the code is under management or requires prior authorization. Refer to the specific Health Plan's procedure code list for management requirements.

<table>
<thead>
<tr>
<th>Procedures addressed by this policy</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Molecular pathology procedure, Level 1, (e.g., identification of single germline variant [e.g. SNP] by techniques such as restriction enzyme digestion or melt curve analysis)</td>
<td>81400</td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 2, (e.g., 2-10 SNPs, 1 methylated variant, or 1 somatic variant [typically using nonsequencing target variant analysis], or detection of a dynamic mutation disorder/triplet repeat)</td>
<td>81401</td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 3, (e.g., greater than 10 SNPs, 2-10 methylated variants, or 2-10 somatic variants [typically using non-sequencing target variant analysis], immunoglobulin and T-cell receptor gene rearrangements, duplication/deletion variants of 1 exon, loss of heterozygosity [LOH], uniparental disomy [UPD])</td>
<td>81402</td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 4, (e.g., analysis of single exon by DNA sequence analysis, analysis of &gt;10 amplicons using multiplex PCR in 2 or more independent reactions, mutation scanning or duplication/deletion variants of 2-5 exons)</td>
<td>81403</td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 5,</td>
<td>81404</td>
</tr>
</tbody>
</table>
### Procedures addressed by this policy

<table>
<thead>
<tr>
<th>Procedures addressed by this policy</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>(e.g., analysis of 2-5 exons by DNA sequence analysis, mutation scanning or duplication/deletion variants of 6-10 exons, or characterization of a dynamic mutation disorder/triplet repeat by Southern blot analysis)</td>
<td></td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 6, (e.g., analysis of 6-10 exons by DNA sequence analysis, mutation scanning or duplication/deletion variants of 11-25 exons)</td>
<td>81405</td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 7, (e.g., analysis of 11-25 exons by DNA sequence analysis, mutation scanning or duplication/deletion variants of 26-50 exons, cytogenomic array analysis for neoplasia)</td>
<td>81406</td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 8, (e.g., analysis of 26-50 exons by DNA sequence analysis, mutation scanning or duplication/deletion variants of &gt;50 exons, sequence analysis of multiple genes on one platform)</td>
<td>81407</td>
</tr>
<tr>
<td>Molecular pathology procedure, Level 9, (e.g., analysis of &gt;50 exons in a single gene by DNA sequence analysis)</td>
<td>81408</td>
</tr>
</tbody>
</table>

### What are molecular pathology tier 2 molecular CPT codes

**Definition**

Tier 2 codes are intended to report a wide range of molecular pathology procedures for which Tier 1 or other test-specific CPT codes have not been assigned.

**Tier 2 code organization**

Tier 2 codes are organized and assigned based on level of technical and interpretive effort required.

**Note** This administrative guideline provides general guidance for billing any test using CPT codes 81400-81408. Clinical criteria may also apply. Review the full list of guidelines to determine if a test-specific guideline is available.
How specific tests become assigned to a Tier 2 code

Requests to have a specific test assigned to a Tier 2 CPT code are reviewed and implemented by the AMA.¹

The expectation is that labs will not self-assign Tier 2 codes based upon their own interpretation of required effort.

Test not assigned to a Tier 2 code

If the test has not been assigned to the appropriate Tier 2 CPT code, use unlisted CPT code 81479.

Criteria

Authorization Requirements

• CPT codes 81400 through 81408 require medical necessity review. The following information must be submitted for review:
  o Details about the test being performed (test name, description, and/or unique identifier), and
  o Laboratory that will be performing the test, and
  o All CPT codes and units that will be billed related to the entire test, and
  o Clinical information, which may include:
    ▪ All information required by test-specific policy, or
    ▪ Test indication, including any applicable signs and symptoms or other reasons for testing, and
    ▪ Any applicable test results (laboratory, imaging, pathology, etc.), and
    ▪ Any applicable family history, and
    ▪ How test results will impact patient care if available

Claims Review and Payment Rules for 81400-81408

• A tier 2 code should only be used when the AMA has specifically assigned the performed test to a tier 2 code (i.e., laboratory self-assigned tier 2 codes will not be accepted).

• Claims submitted for 81400 through 81408 may require a unique test identifier. Please refer to the Unique Test Identifiers for Non-Specific Procedure Codes guideline for additional information.
• All claims received for 81400 through 81408 are subject to the applicable authorization requirements regardless of the specific test performed.

References
Introduction

These references are cited in this guideline.

Unique Test Identifiers for Non-Specific Procedure Codes

Procedures addressed

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<table>
<thead>
<tr>
<th>Procedures addressed by this guideline</th>
<th>Procedure codes</th>
</tr>
</thead>
<tbody>
<tr>
<td>MOPATH PROCEDURE LEVEL 1</td>
<td>81400</td>
</tr>
<tr>
<td>MOPATH PROCEDURE LEVEL 2</td>
<td>81401</td>
</tr>
<tr>
<td>MOPATH PROCEDURE LEVEL 3</td>
<td>81402</td>
</tr>
<tr>
<td>MOPATH PROCEDURE LEVEL 4</td>
<td>81403</td>
</tr>
<tr>
<td>MOPATH PROCEDURE LEVEL 5</td>
<td>81404</td>
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<tr>
<td>MOPATH PROCEDURE LEVEL 6</td>
<td>81405</td>
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<tr>
<td>MOPATH PROCEDURE LEVEL 7</td>
<td>81406</td>
</tr>
<tr>
<td>MOPATH PROCEDURE LEVEL 8</td>
<td>81407</td>
</tr>
<tr>
<td>MOPATH PROCEDURE LEVEL 9</td>
<td>81408</td>
</tr>
<tr>
<td>UNLISTED MOLECULAR PATHOLOGY</td>
<td>81479</td>
</tr>
<tr>
<td>UNLISTED MAAA</td>
<td>81599</td>
</tr>
<tr>
<td>CLINICAL CHEMISTRY TEST</td>
<td>84999</td>
</tr>
</tbody>
</table>

Description

This policy provides instruction on how to submit a unique test identifier when a procedure code is billed that does not adequately describe the performed molecular or genomic test referred to here as “non-specific procedure codes.”

Given the large and rapidly increasing number of molecular and genomic tests, many tests do not have unique procedure codes and are instead billed with non-specific procedure codes. These non-specific procedure codes generally fall into one of the following categories.

Tier 2 codes

Tier 2 Molecular Pathology codes (81400-81408) are a set of CPT codes designed to represent the level of technical and interpretive effort required for a large number of molecular and genomic tests that have not been assigned a unique CPT code.
(i.e., are not addressed by Tier 1, GSP, MAAA, etc. codes). Specific tests, or analytes, are assigned to these Tier 2 codes by the AMA a few times yearly and cannot be self-assigned by the laboratory.

The AMA publishes a set of gene abbreviations or analyte identifiers, called claim designation codes, for each test assigned to a tier 2 code. These codes are intended to provide billing transparency such that the combination of a tier 2 code and the applicable claim designation code on a claim form are reasonably specific to the test performed. Where the test is specific to a gene, the claim designation code is generally the standard gene name. The claim designation codes are published in the annual AMA CPT Professional codebook.

**Unlisted codes**

If a molecular or genomic test has not been assigned to any test-specific or Tier 2 CPT code, those tests are generally billed under one of the following unlisted codes:

- 81479: Unlisted molecular pathology procedure
- 81599: Unlisted multianalyte assay with algorithmic analysis
- 84999: Unlisted chemistry procedure

The proper unlisted code depends on the nature of the test, but most molecular tests are best described by 81479 or 81599.

There is no publicly-available, widely-adopted source of unique codes for tests billed under unlisted codes.

The Palmetto MolDX program requires that most molecular tests be registered with the program and obtain a unique identifier (McKesson Z-Code or Palmetto Test Indicator) for the purposes of claim processing. However, this identifier is both lab and test-specific and is currently primarily utilized by only certain Medicare jurisdictions.

**Criteria**

**Unique test identifier development**

**Tier 2 AMA claim designation codes**

For tests billed under a Tier 2 CPT code, the unique test identifier is the same as the original claim designation code published by the AMA when available, provided the claim designation code described only a single test assigned to that Tier 2 code. In the event that the same claim designation code described more than one test assigned to the same Tier 2 code, eviCore assigned a unique code (not the original AMA claim designation code) to at least one of these tests. When the AMA has not published a claim designation code, a unique code is developed by eviCore. No separate registration or notification process is required on the part of the laboratory.
Tier 2 special cases

Tier 2 code 81403 allows for known familial variant testing to be billed without specific gene assignment. The unique test identifier for known familial variants not otherwise specified is generally the AMA assigned claim designation code for the gene if one exists with the addition of “KFM” (e.g., FGFR3 and FGFR3KFM).

Unlisted codes

For tests billed under unlisted procedure codes, a unique code will be developed unrelated to the Tier 2 claim designation codes. No separate registration or notification process is required on the part of the laboratory.

Obtaining a unique test identifier

When a medical necessity review is performed for a test that will be billed under a non-specific procedure code, billing instructions will include the appropriate unique test identifier if required in the determination communication.

If a medical necessity review is not performed for a test that will be billed under a non-specific procedure code, a unique test identifier can be obtained by contacting eviCore through the phone number provided by the health plan. However, most non-specific procedure codes require medical necessity determination. If pre-service medical necessity determination is required and not obtained, that requirement will take precedence over any other billing requirements. A voluntary medical necessity determination request may also be submitted for any non-specific procedure code regardless of the plan’s requirements.

Billing tests using non-specific procedure codes

When a unique test identifier is provided in the medical necessity determination communication, it must be included on the claim regardless of medical necessity review requirements or determination outcome. Enter the unique test identifier in one of the following narrative fields based on the type of claim being submitted:

<table>
<thead>
<tr>
<th>Claim type</th>
<th>Electronic claim</th>
<th>Paper claim</th>
</tr>
</thead>
<tbody>
<tr>
<td>Professional</td>
<td>837P: Enter in the 2400 SV101-7 field (Line Item Description) associated with the non-specific CPT code. Each non-specific CPT code should have a unique identifier in the associated field.</td>
<td>CMS-1500: Enter in box 24 in the shaded line above the service line that contains the non-specific CPT code. Each non-specific CPT code should have a unique identifier entered above it. Each test identifier should have the qualifier “ZZ” appended at the beginning (e.g., ZZBRAF).</td>
</tr>
<tr>
<td>Claim type</td>
<td>Electronic claim</td>
<td>Paper claim</td>
</tr>
<tr>
<td>------------</td>
<td>-----------------</td>
<td>-------------</td>
</tr>
<tr>
<td><strong>Institutional</strong></td>
<td>837I: Enter in the 2400 SV202-7 field (Line Item Description) associated with the non-specific CPT code. Each non-specific CPT code should have a unique identifier in the associated field.</td>
<td>UB-04: Enter in box 80 (Remarks). Only a single non-specific CPT code should be billed per claim form due to the limitations of a single descriptive field. The test identifier should have the qualifier “ZZ” appended at the beginning (e.g., ZZBRAF).</td>
</tr>
</tbody>
</table>

**References**

<table>
<thead>
<tr>
<th>Term</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>adenoma</td>
<td>An ordinarily benign neoplasm of epithelial tissue. If an adenoma becomes cancerous, it is known as an adenocarcinoma.</td>
</tr>
<tr>
<td>adenomatous polyposis coli</td>
<td>Adenomatous polyposis coli (APC) is a gene located on chromosome 5q. Inherited APC gene mutations are associated with Familial Adenomatous Polyposis (FAP) and Attenuated FAP. Most colorectal cancer polyps have mutations in both copies of the APC gene, even in people that don't have FAP.</td>
</tr>
<tr>
<td>adjuvant therapy</td>
<td>When discussing cancer treatment, adjuvant therapy is given after a primary treatment (like surgery) to increase the chances of a cure. Adjuvant therapy may include chemotherapy, radiation therapy, hormone therapy, or biological therapy.</td>
</tr>
<tr>
<td>adverse drug reaction</td>
<td>A harmful or unpleasant reaction to a drug that generally means the drug should be prescribed differently or avoided.</td>
</tr>
<tr>
<td>aerobic exercise</td>
<td>Any physical activity that causes the heart to pump faster and harder and breathing to quicken. Strengthens the heart muscle and may also help lower high blood pressure and increase good cholesterol.</td>
</tr>
<tr>
<td>AFAP</td>
<td>Attenuated FAP (AFAP) is a form of FAP characterized by a less dramatic proliferation of polyps (between 20-99 cumulative polyps) and age of onset for colorectal cancer of approximately 50 years. Polyps generally localize to the proximal (right-sided) colon. The American Gastroenterological Association (AGA) recommends genetic testing once a person has developed 20 or more cumulative polyps.</td>
</tr>
<tr>
<td>AFP</td>
<td>Short for &quot;alpha-fetoprotein&quot;, a substance found in pregnant women's blood. High levels of AFP are associated with risk for spina bifida and abdominal wall defects.</td>
</tr>
<tr>
<td>amniotic fluid</td>
<td>The protective fluid that surrounds the developing baby. This fluid fills the amniotic sac, or &quot;bag of water&quot; inside the mother's uterus.</td>
</tr>
<tr>
<td>ancestry</td>
<td>Can be represented by a family tree showing how biological family members are related to each other. It is sometimes used interchangeably with &quot;lineage.&quot;</td>
</tr>
<tr>
<td>anemia</td>
<td>A condition caused by too little oxygen in the blood, usually caused by too little hemoglobin or too few red blood cells.</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>angina</td>
<td>Pain, pressure, or a feeling of indigestion in the chest caused by too little oxygen-rich blood reaching the heart. Usually caused by coronary artery disease.</td>
</tr>
<tr>
<td>anticipation</td>
<td>A way certain genetic diseases are inherited that causes them to get worse over the generations.</td>
</tr>
<tr>
<td>anticoagulant</td>
<td>Medications that prevent the blood from clotting -- often call “blood thinners.”</td>
</tr>
<tr>
<td>anticonvulsant drug</td>
<td>Medications used to prevent or treat seizures. Common anticonvulsant drugs include Dilantin, Zantoin, Klonopin, Valium, Tegretol, Depakote and others.</td>
</tr>
<tr>
<td>antidepressant</td>
<td>A medication used to prevent or treat depression. Current antidepressants categories include SSRIs, MAOIs, tricyclics, tetracyclics, and others.</td>
</tr>
<tr>
<td>antipsychotic</td>
<td>Medications used to treat schizophrenia, schizoaffective disorder, bipolar disorder and other conditions that distort a person's grasp of reality</td>
</tr>
<tr>
<td>antiretroviral</td>
<td>A medication used to treat a retrovirus infection, such as HIV</td>
</tr>
<tr>
<td>APOB</td>
<td>A gene for the protein that normally helps deliver LDL cholesterol to the liver to be broken down. An APOB gene mutation causes a person not to clear LDL from the body as well as usual and it builds up. APOB mutations are one cause of familial hypercholesterolemia, although LDLR mutations are the most common.</td>
</tr>
<tr>
<td>Apolipoprotein B100</td>
<td>ApoB100 is short for apolipoprotein B100. It is a normal protein that is a major part of “bad” cholesterol. High ApoB100 is a strong risk factor for heart disease.</td>
</tr>
<tr>
<td>aromatase inhibitor</td>
<td>A class of drugs used to treat postmenopausal women who have hormone-dependent breast cancer. AIs work by blocking the enzyme aromatase responsible for converting androgen to estrogen. This limits the amount of estrogen available to promote breast cancer growth.</td>
</tr>
<tr>
<td>arrhythmia</td>
<td>Any variation from the normal heart rate or rhythm. The heart might beat faster than usual (tachycardia), slower than usual (bradycardia), or with an unusual pattern.</td>
</tr>
<tr>
<td>artery</td>
<td>Blood vessels that carry oxygen-rich blood throughout the body. The coronary arteries carry blood to the heart muscle.</td>
</tr>
<tr>
<td>Ashkenazi Jewish</td>
<td>Jewish people whose ancestors are from Eastern Europe -- mostly Germany, Poland, Russia, and some parts of France. Whereas Sephardic Jewish people have ancestry from Spain, Portugal, parts of France, Italy, North Africa, and the Middle</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>East. Most American Jews are Ashkenazi.</td>
<td>Atherosclerosis is a disease caused by plaque buildup inside the arteries that limits blood flow. Also called hardening of the arteries.</td>
</tr>
<tr>
<td>autosomal dominant</td>
<td>A pattern of inheritance where only one gene from a pair isn't working properly and causes the condition. Anyone with an autosomal dominant condition has a 50% chance of passing on the nonworking gene -- and, therefore, the condition -- to each child.</td>
</tr>
<tr>
<td>autosomal recessive</td>
<td>Describes a pattern of inheritance where both genes from a pair must be working abnormally to cause the condition. People with one abnormal and one normally working gene don't have the condition and are called carriers. When both parents are unaffected carriers of a condition, there is a 25% chance to have an affected child with each pregnancy.</td>
</tr>
<tr>
<td>average woman</td>
<td>The “average woman” is someone picked at random from the general public.</td>
</tr>
<tr>
<td>Beta-thalassemia</td>
<td>An inherited blood disorder that causes anemia, which is a shortage of red blood cells. This disorder causes lower than usual amounts of oxygen in the blood.</td>
</tr>
<tr>
<td>b-hCG</td>
<td>Short for “beta-human chorionic gonadotropin”, this substance is known as the pregnancy hormone. It is produced by the placenta.</td>
</tr>
<tr>
<td>biopsy</td>
<td>The process of removing tissue from living patients for diagnostic evaluation.</td>
</tr>
<tr>
<td>black box warning</td>
<td>A warning required by the U.S. Food and Drug Administration (FDA) on the package inserts of some prescription drugs. These are the strongest warnings from the FDA about a significant risk for serious or life-threatening complications of a drug. Black box refers to the heavy black line surrounding the warning.</td>
</tr>
<tr>
<td>blood clot</td>
<td>Proteins change liquid blood into a solid blood clot usually in response to an injury to prevent further blood loss. Imbalance in the clotting proteins can lead to too little or too much clotting (thrombosis). When an abnormal clot forms, it can block blood flow and cause tissue damage or death.</td>
</tr>
<tr>
<td>blood clotting factor</td>
<td>Proteins and enzymes in the blood that control changing liquid blood into a solid blood clot. Imbalance of these factors can cause too little or too much clotting.</td>
</tr>
<tr>
<td>blood transfusion</td>
<td>Transferring blood or components of blood, such as blood plasma, into a patient.</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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<tr>
<td>blood vessel</td>
<td>The channels that carry blood throughout the body: arteries, veins and capillaries</td>
</tr>
<tr>
<td>bone marrow transplant</td>
<td>A procedure that replaces diseased or damaged bone marrow with healthy bone marrow. The damaged bone marrow may be destroyed by chemotherapy or radiation. The healthy bone marrow can come from the patient or a donor.</td>
</tr>
<tr>
<td>bowel preparation</td>
<td>Purging and cleansing of the bowel of fecal and other matter to assure clear evaluation of the bowel.</td>
</tr>
<tr>
<td>BRCA1</td>
<td>A gene located on chromosome 17 that normally produces a protein to help restrain cell growth. A harmful change in BRCA1 may predispose a person toward developing breast and/or ovarian cancer.</td>
</tr>
<tr>
<td>BRCA2</td>
<td>A gene located on chromosome 13 that normally produces a protein to help to restrain cell growth. A harmful change in BRCA2 may predispose a person toward developing breast and/or ovarian cancer.</td>
</tr>
<tr>
<td>breast MRI</td>
<td>MRI uses powerful magnets and radio waves to create detailed pictures of the breast and surrounding tissues. It provides clear pictures of parts of the breast that are difficult to see clearly on ultrasound or mammogram, but it’s not a replacement for mammography.</td>
</tr>
<tr>
<td>cancer</td>
<td>A disease where abnormal cells grow and divide without control. Cancer cells can invade nearby tissues and spread through the bloodstream and lymphatic system to other parts of the body (called metastasis).</td>
</tr>
<tr>
<td>carbohydrate</td>
<td>Carbohydrates are the most abundant nutrients we eat and are broken down by the liver into glucose (sugar) to provide energy.</td>
</tr>
<tr>
<td>carcinoma</td>
<td>A cancer that begins in the skin or tissues that line or cover internal organs.</td>
</tr>
<tr>
<td>cardiomyopathy</td>
<td>A heart muscle disease that usually leads to a weakened heart muscle and a reduced ability to pump blood effectively. Any damage to the heart muscle can cause cardiomyopathy. Recognized causes include genetic factors, heart attack, alcoholism, and certain viral infections.</td>
</tr>
<tr>
<td>carrier</td>
<td>A person who has one copy of a changed gene and one normal copy of that gene.</td>
</tr>
<tr>
<td>CBC</td>
<td>An abbreviation for “complete blood count”. A standard test that provides information including the white blood cell count, red blood cell count, amount of hemoglobin, platelet count and</td>
</tr>
<tr>
<td>Term</td>
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</tr>
<tr>
<td>CCR5-tropic</td>
<td>A form of HIV virus that uses a protein on the outside of a cell, called the CCR5 receptor, to enter and infect the cell.</td>
</tr>
<tr>
<td>CD4 cells</td>
<td>A kind of white blood cell, also called “helper T cells”, which help protect the body against infection. These are the cells that the HIV virus infects.</td>
</tr>
<tr>
<td>cell</td>
<td>The basic building block of the tissues and organs in the body. Most cells have a complete copy of our genetic code and all cells are made by copying existing cells.</td>
</tr>
<tr>
<td>chelation therapy</td>
<td>Treatment to remove iron from the body using a chemical that attaches to heavy metals inside the body to remove them.</td>
</tr>
<tr>
<td>chemoprevention</td>
<td>The administration of any chemical or drug to treat a disease or condition and limit its further progress, or to prevent the condition from ever occurring.</td>
</tr>
<tr>
<td>cholesterol</td>
<td>A waxy, fat-like substance used by the body to make hormones, vitamin D, and other important substances. Eating too much cholesterol increases the risk of heart disease.</td>
</tr>
<tr>
<td>chromosome</td>
<td>A threadlike strand of DNA that carries genes and transmits hereditary information. Each chromosome can contain hundreds or thousands of individual genes. The number of chromosomes in the normal human cell is 46 (23 pairs).</td>
</tr>
<tr>
<td>chromosome translocation</td>
<td>A genetic condition where material from one chromosome breaks off and sticks to another chromosome, or switches places with a part of another chromosome. There are different types of translocations, and they can have different effects on health and development.</td>
</tr>
<tr>
<td>CHRPE</td>
<td>Congenital Hypertrophy of Retinal Pigmented Epithelium - a benign eye abnormality common in those with FAP.</td>
</tr>
<tr>
<td>close relative</td>
<td>A close relative is defined as a mother, father, sister, brother or child.</td>
</tr>
<tr>
<td>colectomy</td>
<td>The surgical removal of the colon. A total colectomy is the surgical removal of the colon and rectum. A subtotal colectomy is the surgical removal of the colon or portions of the colon only (not rectum).</td>
</tr>
<tr>
<td>colon</td>
<td>Another name for the large intestine; the section of the large intestine extending from the cecum to the rectum. An adult colon is approximately five to six feet in length and is responsible for absorbing water and forming, storing, and expelling waste.</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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<tr>
<td>colonoscopy</td>
<td>A procedure that examines the entire rectum and colon. A colonoscope is a long, flexible, lighted tube with a tiny lens on the end used to directly examine the whole colon and look for the presence of growths. Colonoscopy is the most effective way to evaluate the inside of your entire colon for the presence of colorectal cancer or polyps. This procedure is considered “invasive,” because it requires sedation and the insertion of the colonoscope through the rectum.</td>
</tr>
<tr>
<td>colorectal cancer</td>
<td>Cancer that occurs in the rectum or the colon.</td>
</tr>
<tr>
<td>Comprehensive Analysis</td>
<td>Comprehensive Analysis is the most complete BRCA test. It looks at all the coding DNA of the BRCA1 and BRCA2 genes, to see if there are any changes or mutations. It can find: changes that are known to cause cancer, changes that are harmless, and changes whose link to cancer is unknown.</td>
</tr>
<tr>
<td>congenital heart defect</td>
<td>A problem with the structure of the heart, or the vessels connected to it, which is present from birth. Many types of heart defects exist. They can affect how the blood flows through the heart, or its rhythm.</td>
</tr>
<tr>
<td>corneal arcus</td>
<td>Also called “arcus cornealis”. An accumulation of cholesterol around the cornea (the clear front surface of the eye) that causes a grey ring around the colored part of the eye. May be a normal feature of aging, but may also be a sign of unusually high cholesterol levels.</td>
</tr>
<tr>
<td>CXCR4-tropic</td>
<td>A form of HIV virus that uses a protein on the outside of a cell, called the CXCR4 receptor, to enter and infect the cell.</td>
</tr>
<tr>
<td>CYP1A2</td>
<td>An enzyme involved in the metabolism of many drugs, including caffeine. Some people have a form of CYP1A2 that is particularly susceptible to tobacco smoke and may have adverse reactions when taking drugs metabolized by CYP1A2 while smoking.</td>
</tr>
<tr>
<td>CYP2C19</td>
<td>An enzyme involved in the metabolism of many drugs, including several ulcer and reflux drugs. Variants in the gene can cause adverse reactions to drugs metabolized by CYP2C19.</td>
</tr>
<tr>
<td>CYP2C9</td>
<td>An enzyme involved in the metabolism of many drugs, including warfarin and celecoxib, and several anti-inflammatory drugs. Variants in the gene can cause adverse reactions to drugs metabolized by CYP2C9.</td>
</tr>
<tr>
<td>CYP2D6</td>
<td>An enzyme involved in the metabolism of many drugs, including codeine, tamoxifen, and several antidepressants. Variants in the gene can cause adverse reactions to drugs</td>
</tr>
<tr>
<td>Term</td>
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<tr>
<td>metabolized by CYP2D6.</td>
<td></td>
</tr>
<tr>
<td>cytochrome P450</td>
<td>Cytochrome P450, abbreviated CYP450, is a large family of drug metabolizing enzymes, including CYP1A2, CYP2C9, CYP2C19, and CYP2D6.</td>
</tr>
<tr>
<td>de novo mutation</td>
<td>A mutation that is not running in the family yet, but occurs when a gene is damaged at conception. A de novo mutation can also then be passed on to one’s children.</td>
</tr>
<tr>
<td>Desmoid tumor</td>
<td>Fibrous growth identified generally in the abdominal area associated with FAP and AFAP.</td>
</tr>
<tr>
<td>detection rate</td>
<td>Also called “sensitivity”. Refers to the likelihood that a test will actually find the condition that it is looking for. If a test has a 90% detection rate, it will find 90% (9 out of 10) of people with the condition. Most tests don't have a 100% detection rate, so you should pay attention to detection rates to understand the limitations of any test you consider.</td>
</tr>
<tr>
<td>diabetes</td>
<td>A disease that causes you to have too much glucose (sugar) in your blood because of a problem with the hormone insulin. People with diabetes either can't make insulin (type I) or they can't use it well enough (type II).</td>
</tr>
<tr>
<td>DNA</td>
<td>Stands for “deoxyribonucleic acid”. The chemical inside the nucleus of the cell that encodes the genetic instructions passed from generation to generation. Genes are made of DNA.</td>
</tr>
<tr>
<td>DNA replication</td>
<td>The duplication process of genetic material.</td>
</tr>
<tr>
<td>drug interaction</td>
<td>When a drug reacts with another drug (prescribed, over-the-counter, herbs, supplements, etc.), food, or other environmental exposure to cause an altered response. The effect may be an increased or decreased response or an adverse drug reaction.</td>
</tr>
<tr>
<td>environment</td>
<td>When talking about what causes disease, environment refers to basically everything that isn't controlled by genetics. Environment can include what we eat, physical activity, medications we take, chemicals we are exposed to, our physical surroundings, and countless other factors.</td>
</tr>
<tr>
<td>enzyme</td>
<td>A protein made by the body that encourages a biochemical reaction. Humans make hundreds of different enzymes from the instructions in our genes. If any one enzyme isn't working normally, it can cause a disease.</td>
</tr>
<tr>
<td>epithelium</td>
<td>Membranous tissue constructed of one or more layers of cells that cover the internal and external surfaces of the body and</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>ethnic background</td>
<td>The geographical and racial identity of a person's ancestors.</td>
</tr>
<tr>
<td>ethnic group</td>
<td>A group of people whose ancestors lived in the same region of the world, and thus, who share a common genetic background.</td>
</tr>
<tr>
<td>ethnicity</td>
<td>A group of people who frequently share some common ancestry and are, therefore, more likely to share certain genetic traits or mutations. May be based on descending from the same geographical location, a shared religion, a tribal connection, or other cultural practices. People often belong to more than one ethnic group.</td>
</tr>
<tr>
<td>extensive metabolizer</td>
<td>Extensive metabolizers have two “normal” drug metabolism genes. They make the average amount of enzyme and usually have normal drug response. Most people are extensive metabolizers. People have many drug metabolism genes and can be different kinds of metabolizers for each.</td>
</tr>
<tr>
<td>false negative</td>
<td>A test result that is read as negative when the disease is present.</td>
</tr>
<tr>
<td>false positive</td>
<td>A test result that is read as positive when the disease is not present.</td>
</tr>
<tr>
<td>familial adenomatous polyposis</td>
<td>Familial Adenomatous Polyposis (FAP) is an inherited condition that causes the formation of hundreds to thousands of precancerous polyps within the colon, often before age 20. FAP is usually caused by an inherited mutation in one copy of the APC gene.</td>
</tr>
<tr>
<td>familial hypercholesterolemia</td>
<td>An inherited condition that causes people to have very high levels of LDL, or “bad”, cholesterol and a high risk for heart disease if not aggressively treated with cholesterol-lowering drugs.</td>
</tr>
<tr>
<td>family history</td>
<td>Family history may refer to whether or not you have any biological relative with a specific condition. It may also refer to the collective medical histories of all of your biological relatives. An accurate family history is one of the most important tools available to predict and prevent conditions that you may be at risk for.</td>
</tr>
<tr>
<td>FDA</td>
<td>U.S. Food and Drug Administration, a department of the federal government, that regulates drugs, foods, some tests, medical devices, and other things that may impact public health and safety.</td>
</tr>
<tr>
<td>fecal immunochemical</td>
<td>Fecal immunochemical test (FIT) is a test, similar to FOBT, to check for hidden blood in the stool. Blood may signal cancer</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>test</td>
<td>or one of many non-cancer related causes of bleeding.</td>
</tr>
<tr>
<td>fecal occult blood test</td>
<td>Fecal occult blood test (FOBT) is a test to check for hidden blood in the stool. The presence of blood in stool may be a sign of cancer or one of the many non-cancer related causes of bleeding (e.g. hemorrhoids).</td>
</tr>
<tr>
<td>fibrate</td>
<td>A group of drugs that work to lower your “bad” (LDL) cholesterol by reducing your triglycerides (another type of fat) and raising your “good” (HDL) cholesterol. Commonly prescribed fibrates include fenofibrate (brand name examples include: Antara, Fenoglide, Lipofen, Lofibra, TriCor, Triglide, and Lipidil) and gemfibrozil (brand name: Lopid).</td>
</tr>
<tr>
<td>flexible sigmoidoscopy</td>
<td>Procedure used to examine the rectum and lower third of the colon. A sigmoidoscope is a long, flexible, slender tube with a lens on the end used to visualize a portion of the colon to look for the presence of growths.</td>
</tr>
<tr>
<td>functional</td>
<td>Functional refers to genes or proteins that are not affected by genetic changes that disrupt their normal structure or behavior.</td>
</tr>
<tr>
<td>gastrointestinal tract</td>
<td>The digestive system, consisting of the esophagus, stomach, small intestine and large intestine.</td>
</tr>
<tr>
<td>gene</td>
<td>A piece of DNA that acts as an instruction to the body for how to make a specific protein (enzyme, hormone, etc.). Genes are inherited, passed from parent to child.</td>
</tr>
<tr>
<td>gene sequencing</td>
<td>A genetic test that is considered the gold standard for finding genetic changes known as mutations.</td>
</tr>
<tr>
<td>genetic</td>
<td>Refers to any trait that is inherited, or passed from generation to generation through genes. These traits may range from having specific diseases to our response to certain drugs to simply our physical characteristics, like eye and hair color.</td>
</tr>
<tr>
<td>genetic condition</td>
<td>A genetic condition is any disease, disorder, syndrome, or trait that is caused, at least in part, from alterations in genes or chromosomes.</td>
</tr>
<tr>
<td>genetic counseling</td>
<td>Genetic counseling is a process to help people learn about, cope with, and manage their risk of genetic disorders. This risk may be uncovered because the person is diagnosed with a condition, has a family history, has an affected child, and/or has an abnormal genetic test result.</td>
</tr>
<tr>
<td>genetic counselor</td>
<td>A healthcare professional with specialized training in how the science of genetics relates to medical care. A genetic counselor can evaluate your personal and family history,</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>genetic discrimination</td>
<td>Treatment or consideration based on genetic status or category rather than individual merit or actual conditions.</td>
</tr>
<tr>
<td>genetic modifier</td>
<td>A gene that changes how another gene is expressed.</td>
</tr>
<tr>
<td>genetic predisposition</td>
<td>Any condition in which genetic make-up leaves the individual more susceptible to disease.</td>
</tr>
<tr>
<td>genetic test</td>
<td>A specific type of laboratory test that is designed to find out if a person has a genetic disorder, is a carrier of a genetic disease, or has a predisposition to develop a genetic problem. Genetic testing can look at chromosomes, genes, or proteins -- depending on the specific condition being tested.</td>
</tr>
<tr>
<td>genomics</td>
<td>The study of the genome and its significance to pathology and disease.</td>
</tr>
<tr>
<td>genotype</td>
<td>The version of genes a person, organism, or cancer has.</td>
</tr>
<tr>
<td>genotyping</td>
<td>Tests that look specifically at the genetic information of a person, organism, or cancer. These tests may predict a certain characteristic (&quot;phenotype&quot;) but don't actually test for that characteristic.</td>
</tr>
<tr>
<td>glucose</td>
<td>A form of sugar made from carbohydrates we eat that the body uses for energy. Too much glucose in their blood may be a sign of diabetes.</td>
</tr>
<tr>
<td>HBB</td>
<td>A gene involved in making a piece of a protein called hemoglobin. Genetic changes, or mutations, in the HBB gene can cause sickle cell disease and beta-thalassemia.</td>
</tr>
<tr>
<td>HDL</td>
<td>High density lipoprotein cholesterol. Also called the “good” cholesterol. High HDL lowers the risk for heart disease.</td>
</tr>
<tr>
<td>HDL2</td>
<td>A subtype of HDL (the “good” cholesterol). HDL2 is the “best” cholesterol because high levels give you the most protection against heart disease -- even more than just high total HDL.</td>
</tr>
<tr>
<td>HDL3</td>
<td>A subtype of HDL (the “good” cholesterol). HDL3 is not as good for you as other types of HDL. Some studies show that high levels of HDL3 may actually increase your risk for heart disease.</td>
</tr>
<tr>
<td>heart</td>
<td>A muscular organ whose primary job is to pump blood to all parts of the body.</td>
</tr>
<tr>
<td>heart attack</td>
<td>When the blood supply to part of the heart muscle is suddenly blocked. The heart muscle may be damaged or start to die if the blood supply is not restored.</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>heart disease</td>
<td>A general term for any condition that threatens the heart's ability to function normally. Because coronary artery disease (plaque buildup that may cause a heart attack) is by far the most common type, it is often just called heart disease.</td>
</tr>
<tr>
<td>hemochromatosis</td>
<td>A condition in which too much iron builds up in the body, which can lead to organ damage.</td>
</tr>
<tr>
<td>hemoglobin</td>
<td>A protein found in red blood cells that carries oxygen throughout the body</td>
</tr>
<tr>
<td>hemoglobin analysis</td>
<td>A test that measures the different types of hemoglobin in the blood. It is used to diagnose diseases caused by abnormal hemoglobin, such as sickle cell anemia.</td>
</tr>
<tr>
<td>hereditary</td>
<td>Genetically transmitted -- or capable of being transmitted -- from parent to child.</td>
</tr>
<tr>
<td>hereditary nonpolyposis colorectal cancer</td>
<td>Hereditary non-polyposis colorectal cancer (HNPCC) is an inherited disorder in which there is a tendency to develop colorectal cancer without a significant number of polyp precursors. HNPCC is specifically associated with inherited mutations in five mismatch repair genes.</td>
</tr>
<tr>
<td>HFE gene</td>
<td>The HFE gene makes a protein that regulates how much iron your body absorbs from your diet.</td>
</tr>
<tr>
<td>high performance liquid chromatography</td>
<td>A laboratory procedure that can separate a liquid mixture into its individual compounds. As an example, this procedure is used to separate different kinds of hemoglobins in a person's blood.</td>
</tr>
<tr>
<td>HNPCC-related cancer</td>
<td>Other primary cancers included in an inherited cancer syndrome because of the increased prevalence in syndrome carriers. In addition to colon cancer, HNPCC-related cancers include cancer of the endometrium, ovary, stomach, kidney/urinary tract, brain, biliary tract, central nervous system and small bowel.</td>
</tr>
<tr>
<td>hormone</td>
<td>Chemical messengers made mostly in our glands that influence our growth and development, sexual function, reproduction, mood, and metabolism. Hormone medications include oral contraceptive pills, patches or rings; hormonal treatments for infertility; hormone replacement therapy; or serum estrogen modifiers (sometimes taken to treat or prevent certain forms of cancer).</td>
</tr>
<tr>
<td>human immunodeficiency</td>
<td>A retrovirus that attacks the human immune system, thus affecting the body's ability to fight off the organisms that cause disease.</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>virus</td>
<td>disease. HIV is the cause of acquired immune deficiency syndrome or AIDS.</td>
</tr>
<tr>
<td>hypertension</td>
<td>Blood pressure that stays at 140/90 mmHg or higher over a period of time. Average blood pressure is about 120/80 mmHg.</td>
</tr>
<tr>
<td>IDL</td>
<td>Intermediate density lipoprotein -- a type of “bad” cholesterol. High IDL increases the risk for heart disease even more than just high total LDL levels. IDL is under strong genetic control so close relatives of someone with high IDL should also consider testing.</td>
</tr>
<tr>
<td>in vitro fertilization</td>
<td>A laboratory procedure in which sperm fertilize eggs outside the body in a laboratory setting to facilitate pregnancy. The fertilized egg is then placed in the woman's uterus for implantation.</td>
</tr>
<tr>
<td>inherited</td>
<td>Any trait that is passed from generation to generation through our genes. These traits may range from having a specific disease to how we respond to certain drugs to simply our physical characteristics, like eye and hair color.</td>
</tr>
<tr>
<td>inhibin A</td>
<td>A substance made by the placenta during pregnancy and found in the mother's blood. Also abbreviated “DIA.”</td>
</tr>
<tr>
<td>insulin</td>
<td>A hormone that helps glucose, the sugar used by the body for energy, get into the cells that need it. When you don't make enough insulin or you can't use insulin effectively, you are likely to develop diabetes.</td>
</tr>
<tr>
<td>intermediate metabolizer</td>
<td>Intermediate metabolizers have a drug metabolism gene that doesn't work properly. They make less of the enzyme coded for by those genes, but usually make enough to process most drugs. People have many drug metabolism genes and can have be different kinds of metabolizers for each.</td>
</tr>
<tr>
<td>iron overload</td>
<td>A condition in which higher-than-usual amounts of iron collect in the tissues of the body. Over time, iron overload can damage organs like the liver and cause problems like diabetes.</td>
</tr>
<tr>
<td>K-RAS</td>
<td>A gene that when mutated contributes to converting a normal cell into a cancerous cell.</td>
</tr>
<tr>
<td>LDL</td>
<td>Low-density lipoprotein cholesterol. Also called the “bad” cholesterol. High LDL increases the risk of heart disease.</td>
</tr>
</tbody>
</table>
| LDLR                 | Stands for low density lipoprotein receptor. The LDLR gene normally makes a protein that helps to remove LDL ((bad≈ cholesterol) from the blood. An LDLR gene mutation causes a
<table>
<thead>
<tr>
<th>Term</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>person not to get rid of LDL as quickly and it builds up. LDLR mutations are the most common cause of familial hypercholesterolemia.</td>
<td></td>
</tr>
<tr>
<td>leukemia</td>
<td>A cancer that starts in blood-forming tissue, such as the bone marrow, and causes large numbers of abnormal blood cells to be produced and enter the bloodstream.</td>
</tr>
<tr>
<td>lifestyle</td>
<td>In talking about health conditions, lifestyle generally refers to factors within your control like diet, physical activity, smoking, alcohol use, and use of other preventive health measures.</td>
</tr>
<tr>
<td>lipid</td>
<td>A fat that acts as a source of energy and helps the body use certain vitamins. Cholesterol and triglycerides are examples of lipids. High lipid levels increase the risk for heart disease and diabetes and may be caused by eating too much fat, alcohol use, inactivity, inherited conditions, and certain medications and disease.</td>
</tr>
<tr>
<td>lipoprotein a</td>
<td>Lp(a) stands for lipoprotein a -- a type of “bad” cholesterol. High Lp(a) increases the risk of heart disease 2 to 10 times more than just high total LDL levels and may cause heart disease earlier than usual. Drug therapy is usually needed. Lp(a) is under strong genetic control so close relatives of someone with high Lp(a) should also consider testing.</td>
</tr>
<tr>
<td>liver</td>
<td>An organ involved in a wide range of functions, including helping with digestion and the detoxification of chemicals.</td>
</tr>
<tr>
<td>liver biopsy</td>
<td>A surgical procedure that removes a small piece of liver so it can be examined in a lab.</td>
</tr>
<tr>
<td>lymphoma</td>
<td>Cancer that begins in the cells of the immune system.</td>
</tr>
<tr>
<td>maintenance dose</td>
<td>The amount of drug that is needed over the long-term to reach a stable, therapeutic response.</td>
</tr>
<tr>
<td>malignant</td>
<td>Cancerous. Malignant tumors, or cancer, have the ability to invade adjacent tissues and spread throughout the body. Thus, malignant tumors can become life threatening.</td>
</tr>
<tr>
<td>mammogram</td>
<td>An X-ray picture of the breast. The x-ray images make it possible to detect tumors that cannot be felt. They can also find microcalcifications that may signal the presence of cancer.</td>
</tr>
<tr>
<td>maraviroc</td>
<td>The generic name of Selzentry, a drug used to treat HIV infection that only works in people whose HIV uses a specific receptor (CCR5) to infect the cell.</td>
</tr>
<tr>
<td>maternal serum screening test</td>
<td>A blood test that looks at the levels of certain substances in a pregnant woman’s blood. These tests are used to find the risk for having certain birth defects. They can’t tell for sure whether</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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<td>-------------------------------------------------------------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>a pregnancy has a birth defect.</td>
<td></td>
</tr>
<tr>
<td>MCH</td>
<td>An abbreviation for “mean corpuscular hemoglobin”. The average amount of hemoglobin in the average red blood cell. The normal range for the MCH is 27 - 32 picograms. MCH is a standard part of a CBC (complete blood count) test.</td>
</tr>
<tr>
<td>MCV</td>
<td>An abbreviation for “mean corpuscular volume”. The average size of a red blood cell. The normal range for the MVC is 80 - 100 femtoliters. MVC is a standard part of the CBC (complete blood count) test.</td>
</tr>
<tr>
<td>Mediterranean</td>
<td>Someone whose ancestors come from one of the countries bordering the Mediterranean Sea. These countries include but are not limited to: Spain, southern France, Italy, and Greece.</td>
</tr>
<tr>
<td>metabolic syndrome</td>
<td>Also called “insulin resistance”. A combination of factors (like abnormal cholesterol, abdominal obesity, high blood sugar, and high blood pressure) that increases the risk of getting both heart disease and diabetes.</td>
</tr>
<tr>
<td>metabolism or metabolize</td>
<td>The way drugs and other substances are broken down for use in the body and elimination.</td>
</tr>
<tr>
<td>metastasis</td>
<td>The spread of cancer from one part of the body to another.</td>
</tr>
<tr>
<td>methylation</td>
<td>A process by which a methyl group is added to the DNA base cytosine. This process often decreases the amount of gene product that is made. For example, tumor suppressor genes are often methylated which decrease their function and lead to cancer.</td>
</tr>
<tr>
<td>mhl1</td>
<td>A mismatch repair (MMR) gene located on chromosome 3. Mutations in MLH1 are associated with Lynch syndrome (also called HNPCC) and greatly increase the chance of cancer -- especially colon.</td>
</tr>
<tr>
<td>MMR gene</td>
<td>Mismatch repair gene, a gene that functions as a part of the “spell check” system of a cell. Mutations in MMR genes are involved in causing some hereditary cancer syndromes.</td>
</tr>
<tr>
<td>morbidity</td>
<td>A diseased state.</td>
</tr>
<tr>
<td>MSH2</td>
<td>A mismatch repair (MMR) gene located on chromosome 2. Mutations in MLH1 are associated with Lynch syndrome (also called HNPCC) and greatly increase the chance of cancer -- especially colon.</td>
</tr>
<tr>
<td>multifactorial inheritance</td>
<td>Conditions that are caused by an interaction between more than one gene and environmental (non-genetic) factors. Most common human diseases seem to be multifactorial, including diabetes, heart disease, mental illness, and most birth defects.</td>
</tr>
<tr>
<td>Term</td>
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</tr>
<tr>
<td>A family history of a multifactorial condition usually increases the risk for other relatives.</td>
<td></td>
</tr>
<tr>
<td>multiple myeloma</td>
<td>Cancer that begins in the cells of the immune system.</td>
</tr>
<tr>
<td>multisite</td>
<td>Multisite Testing looks for the three BRCA gene mutations that cause 80% to 90% of all hereditary breast and ovarian cancers in Ashkenazi Jewish people. This test gives you a clear result: either you have one of these three mutations, or you don't. If you don't, it is possible to have a different BRCA mutation that was not tested for.</td>
</tr>
<tr>
<td>mutation</td>
<td>A change in the DNA code that may cause a gene not to function in the normal way.</td>
</tr>
<tr>
<td>newborn screening</td>
<td>Testing that is done routinely after birth, to look for serious developmental, genetic and metabolic disorders. This testing is done so that important medical treatments or other actions can start before symptoms develop.</td>
</tr>
<tr>
<td>niacin</td>
<td>Also called “nicotinic acid”. Part of vitamin B3 found in foods like meat, fish, milk, eggs, green vegetables, and grains. Niacin supplements increase HDL, lower Lp(a), and to a lesser degree, lower LDL cholesterol. Common brand names include: Niacor, Niaspan, Nicolar, Nicotinex Elixir, and Slo-Niacin.</td>
</tr>
<tr>
<td>non-invasive procedure</td>
<td>Procedures that do not require insertion of an instrument or device through the skin or a bodily orifice for diagnosis or treatment.</td>
</tr>
<tr>
<td>Noonan syndrome</td>
<td>A genetic disorder that causes abnormal development of many parts of the body. It can be caused by a defect in one of four different genes (KRAS, PTPN11, RAF1, SOS1). Noonan syndrome may be inherited from a parent who has the condition, or may happen by chance in a pregnancy.</td>
</tr>
<tr>
<td>obesity</td>
<td>Having a high amount of body fat. Usually defined by a body mass index (BMI) of 30 or higher.</td>
</tr>
<tr>
<td>omega 3-fatty acid</td>
<td>Also called “fish oil”. Omega-3 fatty acids from eating oily fish or taking fish oil supplements may lower triglycerides, slow the buildup of plaque in the arteries, and raise HDL (“good”) cholesterol. Too much omega-3 fatty acid is dangerous, so you should always talk to your doctor before starting supplements.</td>
</tr>
<tr>
<td>organs</td>
<td>A grouping of tissue that works together to perform a common function. Examples of organs include: stomach, lungs, and liver.</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>osteoma</td>
<td>Benign, bony tumors often on the skull or mandible (sometimes a clinical finding with FAP patients).</td>
</tr>
<tr>
<td>over-the-counter</td>
<td>OTC or over-the-counter drugs can be bought without a prescription. OTC drugs still carry certain risks and may interact with other drugs.</td>
</tr>
<tr>
<td>P-53</td>
<td>A gene which normally regulates the cell cycle and protects the cell from damage to its genome. Mutations in this gene cause cells to develop cancer.</td>
</tr>
<tr>
<td>PAPP-A</td>
<td>Short for “pregnancy-associated plasma protein A”, a substance found in pregnant women's blood. Low levels of PAPP-A at 8-14 weeks of pregnancy have been associated with risk for Down syndrome and pregnancy complications.</td>
</tr>
<tr>
<td>pedigree</td>
<td>A diagram of biological relationships that usually includes information on each relative's medical history.</td>
</tr>
<tr>
<td>premenopausal</td>
<td>The time when a woman is entering menopause until it is complete -- often defined as from the time periods become irregular until 12 months after the last period.</td>
</tr>
<tr>
<td>phenotype</td>
<td>Characteristics that can be seen or measured and are often the result of genes and environment working together. Examples include things like eye color, weight, IQ, cholesterol levels, or drug response.</td>
</tr>
<tr>
<td>phenotyping</td>
<td>Tests that measure specific traits or characteristics that can be caused by genes and/or environmental factors. This is in contrast to genotype testing that only looks at genetic information.</td>
</tr>
<tr>
<td>placebo</td>
<td>A phony treatment or &quot;sugar pill&quot;. Researchers often compare people taking a drug with those taking a placebo to better measure the real effects of the drug.</td>
</tr>
<tr>
<td>placenta</td>
<td>Also called the afterbirth, the placenta is the tissue that connects the developing baby to the mother's uterus. It develops as part of the pregnancy and has the same DNA as the developing baby. The placenta allows for the exchange of nutrients, waste and gases between the developing baby and the mother.</td>
</tr>
<tr>
<td>plaque</td>
<td>Related to heart disease, plaque is the buildup of cholesterol, calcium, and other substances on the inside walls of the arteries causing the arteries to be more narrow and less flexible.</td>
</tr>
<tr>
<td>plasma</td>
<td>The liquid part of the blood that carries blood cells and other components.</td>
</tr>
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<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>polymorphism</td>
<td>Natural differences in a DNA sequence that are usually common and do not cause disease</td>
</tr>
<tr>
<td>polyp</td>
<td>A usually non-cancerous growth or tumor protruding from the lining of an organ, such as the colon. Left untreated, polyps have an increased risk of becoming cancerous.</td>
</tr>
<tr>
<td>poor metabolizer</td>
<td>Produce inactive drug metabolism enzyme or no enzyme at all. Poor metabolizers may have a reduced response or no response and may have increased side effects</td>
</tr>
<tr>
<td>poor metabolizer</td>
<td>Poor metabolizers have a pair of drug metabolism genes that don't work properly. They make very little or none of the enzyme coded for by that pair of genes. This causes slower metabolism or the inability to process certain drugs. People have many drug metabolism genes and can be different kinds of metabolizers for each.</td>
</tr>
<tr>
<td>postmenopausal</td>
<td>The time in a woman's life after menopause is complete -- often defined as starting 12 months after the last period.</td>
</tr>
<tr>
<td>pre-cancerous</td>
<td>Condition of the tissue, such as a polyp, that can turn into a cancer if not treated or removed.</td>
</tr>
<tr>
<td>preconception</td>
<td>Generally considered the period of time when a person is planning pregnancy but has not yet conceived (become pregnant).</td>
</tr>
<tr>
<td>pre-diabetes</td>
<td>Diagnosed when glucose (sugar) levels are higher than normal, but not high enough to make the diagnosis of diabetes -- usually a fasting glucose of 100 to 125 mg/dL or a glucose of 140 to 199 mg/dL after glucose tolerance test.</td>
</tr>
<tr>
<td>predisposition</td>
<td>Any condition, genetic or other, that renders an individual more susceptible to disease.</td>
</tr>
<tr>
<td>preimplantation genetic diagnosis</td>
<td>A technique used with in vitro fertilization to test early-stage embryos for disease-causing genes, so that embryos without the disease-causing genes can be implanted in the mother's uterus.</td>
</tr>
<tr>
<td>prenatal diagnosis</td>
<td>Testing for diseases in the fetus or embryo before it is born.</td>
</tr>
<tr>
<td>presymptomatic</td>
<td>The stage prior to an individual presenting with symptoms that are clinically relevant to the disease in question.</td>
</tr>
<tr>
<td>prophylactic bilateral mastectomy</td>
<td>A risk-reducing treatment where both breasts, as well as some of the surrounding tissue, are surgically removed in order to keep cancerous cells from forming.</td>
</tr>
<tr>
<td>prophylactic bilateral</td>
<td>A risk-reducing treatment where ovaries are surgically removed in order to keep cancerous cells from forming;</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>oophorectomy</td>
<td>recommended after childbearing is complete.</td>
</tr>
<tr>
<td>protein</td>
<td>Large, complex molecules made of amino acids that form body structures, enzymes, hormones, and antibodies. Proteins are all made based on the instructions in our genes. The amino acids we need to make new proteins are consumed in the protein we eat or made by the body.</td>
</tr>
<tr>
<td>protein(s)</td>
<td>The molecules that form the body, allow it to grow, and regulate how it works. Our bodies make the proteins we need using the instructions from our genes.</td>
</tr>
<tr>
<td>receptor</td>
<td>A protein on the surface of a cell that only binds with certain other molecules. When this happens, a cellular process can occur.</td>
</tr>
<tr>
<td>rectum</td>
<td>The last portion of the digestive tract, at the end of the colon.</td>
</tr>
<tr>
<td>red blood cells</td>
<td>A cell in the blood that carries oxygen to all parts of the body. Also called an erythrocyte.</td>
</tr>
<tr>
<td>risk factor</td>
<td>Anything that increases the chance of developing a certain disease or having a child with a specific condition. Risk factors might include your family history, lifestyle, other health conditions, blood test results, age, gender, and countless other factors.</td>
</tr>
<tr>
<td>sarcoma</td>
<td>A cancer that begins in bone, cartilage, fat, muscle, blood vessels, or other connective or supportive tissues.</td>
</tr>
<tr>
<td>screening</td>
<td>In medicine, screening generally refers to a test or exam that is reasonably simple, inexpensive, and harmless that can be given to a large group of people in order to find a smaller group with a higher-than-average chance for a certain condition. These people will sometimes have more specific testing or be treated early before symptoms appear.</td>
</tr>
<tr>
<td>selective estrogen receptor modulator</td>
<td>Selective Estrogen Receptor Modulator (SERM) is a hormone-like drug that affects multiple tissues by interacting with receptors for the hormone estrogen. A particular SERM may have estrogen-like effects in some tissues and anti-estrogen effects in others.</td>
</tr>
<tr>
<td>Selzentry</td>
<td>The brand name of maraviroc, a drug used to treat HIV infection that only works in people whose HIV uses a specific receptor (CCR5) to enter the cell.</td>
</tr>
<tr>
<td>sequencing</td>
<td>A lab method that looks at each DNA nucleotide (A,T,G, and C) in a piece of DNA for differences (mutations) from the usual DNA sequence. A more labor intensive and expensive test that is often used when the specific mutations that cause a disease</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
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</tr>
<tr>
<td>aren't known.</td>
<td></td>
</tr>
<tr>
<td>serum CA-125</td>
<td>A blood test used in an effort to detect ovarian cancer.</td>
</tr>
<tr>
<td>serum ferritin</td>
<td>A protein your body makes when it stores iron.</td>
</tr>
<tr>
<td>siblings</td>
<td>Brothers and/or sisters.</td>
</tr>
<tr>
<td>sickle cell disease</td>
<td>An inherited disorder in which the red blood cells have an abnormal crescent shape that affects blood flow. This disorder causes anemia because the abnormal blood cells don't survive long.</td>
</tr>
<tr>
<td>sickle/beta-thalassemia</td>
<td>A disease that occurs when someone inherits a sickle-cell anemia gene mutation from one parent and a beta-thalassemia gene mutation from the other parent. Symptoms are usually very similar to sickle cell disease.</td>
</tr>
<tr>
<td>side effect</td>
<td>An unintended and usually undesired reaction to a drug or treatment.</td>
</tr>
<tr>
<td>Single Site</td>
<td>Single Site Testing looks for just one BRCA mutation. This test can only be done for people who know the DNA sequence of a BRCA mutation that is running in their family. This test gives you a clear result: Either you have the mutation that was tested for or you don't.</td>
</tr>
<tr>
<td>southeast Asian</td>
<td>Someone whose ancestors come from one of the countries south of China and east of India. These countries include but are not limited to: Vietnam, Cambodia, Laos, Burma, or Indonesia.</td>
</tr>
<tr>
<td>spleen</td>
<td>An organ in the abdomen that supports the immune system, destroys and filters out old blood cells, and holds a reserve of blood cells. People can live without a spleen.</td>
</tr>
<tr>
<td>sporadic</td>
<td>In reference to cancer, this means a cancer not caused by hereditary genetic mutations. Most cancers are sporadic.</td>
</tr>
<tr>
<td>statin</td>
<td>A group of drugs that lower the amount of cholesterol made naturally by the liver. When diet and exercise changes aren't enough, statins are often the first choice for drug therapy. Commonly prescribed statins include: Lovastatin (Mevacor, Altoprev), Pravastatin (Pravachol), Simvastatin (Zocor), Fluvastatin (Lescol), Atorvastatin (Lipitor), and Rosuvastatin (Crestor).</td>
</tr>
<tr>
<td>Stevens-Johnson syndrome</td>
<td>An allergic reaction to a drug or infection that causes flu-like symptoms, skin wounds, and may affect other organs like the eyes and mouth.</td>
</tr>
</tbody>
</table>
| stroke                      | Caused by a sudden lack of blood supply and oxygen to the...
<table>
<thead>
<tr>
<th>Term</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>brain</td>
<td>Usually happens because either a blood clot blocks a blood vessel in the brain (ischemic stroke) or a blood vessel breaks and bleeds into the brain (hemorrhagic stroke).</td>
</tr>
<tr>
<td>symptom</td>
<td>Any sign that a person has a condition or disease. Symptoms, like headache, fever, fatigue, nausea, vomiting, and pain, may not be specific but together point to an underlying cause.</td>
</tr>
<tr>
<td>symptoms</td>
<td>Changes or signs that are caused by or accompany a disease or condition. Symptoms are the evidence of that underlying disease or condition. Symptoms can be used to help diagnose a problem.</td>
</tr>
<tr>
<td>tamoxifen</td>
<td>A drug commonly used to treat patients with breast cancer, certain other cancers, and those at high risk for breast cancer. It works by interfering with the activity of the hormone estrogen, which feeds the growth of many, but not all breast cancers.</td>
</tr>
<tr>
<td>toxic epidermal necrolysis</td>
<td>A life-threatening allergic reaction started by certain drugs, infections, illnesses, and unknown factors. TEN can cause large areas of the skin to peel away, flu-like symptoms, and other complications. The condition gets worse quickly and usually requires hospitalization.</td>
</tr>
<tr>
<td>transferrin saturation</td>
<td>The percentage of transferrin (a protein that carries iron in the blood) that is currently carrying iron.</td>
</tr>
<tr>
<td>translocation</td>
<td>A genetic condition where material from one chromosome breaks off and sticks to another chromosome, or switches places with a part of another chromosome. There are different types of translocations, and they can have different effects on health and development.</td>
</tr>
<tr>
<td>transvaginal ultrasound</td>
<td>A type of ultrasound done by inserting an ultrasound probe into the vagina. This allows a view of a woman's reproductive organs, including the uterus, ovaries, cervix, and vagina.</td>
</tr>
<tr>
<td>triglycerides</td>
<td>A type of energy-rich fat. High triglycerides (over 200mg/dL) increase the risk for heart disease and stroke.</td>
</tr>
<tr>
<td>tropism</td>
<td>The specific cell types that a virus can recognize and infect.</td>
</tr>
<tr>
<td>tumor</td>
<td>An abnormal mass of tissue that results from excessive cell division. Tumors may be benign (not cancerous) or malignant (cancerous).</td>
</tr>
</tbody>
</table>
| Turner syndrome             | A genetic condition in which a girl or woman does not have the usual pair of two X chromosomes. Instead, some or all of her cells are missing an X chromosome, or part of an X chromosome. Symptoms are variable but usually include short
<table>
<thead>
<tr>
<th>Term</th>
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</tr>
</thead>
<tbody>
<tr>
<td>stature and infertility</td>
<td></td>
</tr>
<tr>
<td>ultra metabolizer</td>
<td>Have more than two functional copies of a drug metabolism gene, and produce a larger-than-normal amount of enzyme. Ultra metabolizers may have a reduced or no response and may have increased side effects</td>
</tr>
<tr>
<td>ultrarapid metabolizer</td>
<td>Ultrarapid metabolizers have extra copies of a gene involved in drug metabolism, so they make more enzyme than the average person. This results in faster metabolism of drugs processed by that enzyme.</td>
</tr>
<tr>
<td>umbilical cord</td>
<td>The cord that connects the developing baby to the placenta, which is attached to the mother's uterus. The umbilical cord carries oxygen- and nutrient-rich blood to the developing baby.</td>
</tr>
<tr>
<td>unconjugated estriol</td>
<td>One of the three main estrogens produced by the body. Low levels of this substance are associated with risk for certain birth defects, including Down syndrome and trisomy 18. Also abbreviated “uE3.”</td>
</tr>
<tr>
<td>variant</td>
<td>Gene variations contribute to diversity and make people unique. When a certain form of a gene is seen in at least 1% of people, but not most people, it is called a variant. Variants may also increase or decrease a person's risk for certain genetic diseases but usually don't cause the disease themselves.</td>
</tr>
<tr>
<td>vein</td>
<td>Blood vessels that carry blood low in oxygen back to the heart.</td>
</tr>
<tr>
<td>virtual colonoscopy</td>
<td>A method of examining the colon by taking a series of X-rays (called a CT scan) and using a high-powered computer to reconstruct 2-D and 3-D pictures of the interior surfaces of the colon from these X-rays.</td>
</tr>
<tr>
<td>VKORC1</td>
<td>A gene that tells the body how to make vitamin K epoxide reductase (VKOR), an enzyme important in forming blood-clotting factors. A common VKORC1 gene variant (-1639G&gt;A) puts people at increased risk for complications when taking warfarin at standard doses.</td>
</tr>
<tr>
<td>VLDL</td>
<td>Very low density lipoprotein -- a type of “bad” cholesterol. High VLDL increases the risk for plaque buildup in the arteries and heart disease.</td>
</tr>
<tr>
<td>VLDL3</td>
<td>A subtype of VLDL (a “bad” cholesterol). High VLDL3 increases heart disease risk the most and is a risk factor even when total cholesterol levels are normal. Diet and exercise changes are very effective for lowering VLDL3.</td>
</tr>
<tr>
<td>warfarin</td>
<td>The most commonly prescribed drug for preventing harmful</td>
</tr>
<tr>
<td>Term</td>
<td>Definition</td>
</tr>
<tr>
<td>--------------------</td>
<td>------------------------------------------------------------------------------------------------------</td>
</tr>
<tr>
<td>blood clots</td>
<td>blood clots from forming or from growing larger. Belongs to a class of drugs called anticoagulants or “blood thinners.”</td>
</tr>
<tr>
<td>white blood cells</td>
<td>A cell found in the blood whose primary job is to defend the body against infection.</td>
</tr>
<tr>
<td>xanthoma</td>
<td>Fat buildup that looks like a yellow lump under the skin, most commonly on the heels, hands, elbows, other joints, feet, and buttocks. Especially common in people with inherited high cholesterol like familial hypercholesterolemia.'</td>
</tr>
</tbody>
</table>